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Short communication

Intracranial pressure changes during mouse development

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ABSTRACT

During early stages of postnatal development, pressure from the growing brain as well as cerebrospinal fluid, i.e. intracranial pressure (ICP), load the calvarial bones. It is likely that such loading contributes to the peripheral bone formation at the sutural edges of calvarial bones, especially shortly after birth when the brain is growing rapidly. The aim of this study was to quantify ICP during mouse development. A custom pressure monitoring system was developed and calibrated. It was then used to measure ICP in a total of seventy three wild type mice at postnatal (P) day 3, 10, 20, 31 and 70. Retrospectively, the sample in each age group with the closest ICP to the average value was scanned using micro-computed tomography to estimate cranial growth. ICP increased from 1.33 ± 0.87 mmHg at P3 to 1.92 ± 0.78 mmHg at P10 and 3.60 ± 1.08 mmHg at P20. In older animals, ICP plateaued at about 4 mmHg. There were statistically significant differences between the ICP at the P3 vs. P20, and P10 vs. P20. In the samples that were scanned, intracranial volume and skull length followed a similar pattern of increase up to P20 and then plateaued at older ages. These data are consistent with the possibility of ICP being a contributing factor to bone formation at the sutures during early stages of development. The data can be further used for development and validation of computational models of skull growth.

1. Introduction

During early stages of postnatal development, intracranial pressure (ICP), from the growing brain and cerebrospinal fluid load calvarial bones and sutures (Moss, 1954; Cohen, 1993; Opperman, 2000; Herring, 2008). It is likely that such loading contributes as an epigenetic factor to the peripheral bone formation at the edges of calvarial bones just after birth. Once forceful mastication starts, muscles also load the craniofacial system and presumably influence cranial growth (Nakata, 1981; Rafferty and Herring, 1999; Al Dayeh et al., 2013). While there is a large ongoing effort to understand the genetic causes of various craniofacial developmental disorders (e.g. Morriss-Kay and Wilkie, 2005; Richtsmeier and Flaherty, 2013; Cox et al., 2013), understanding epigenetic factors such as biomechanical loading based on ICP during normal and abnormal development is crucial too. A broad understanding of the various factors involved in the development of the craniofacial system can in the long term enhance the treatment of various congenital diseases such as craniosynostosis and Treacher Collins syndrome.

Quantifying ICP in infants is clearly challenging, but animal models can provide invaluable insights. In particular, accurate invasive bur accurate methods can be employed, rather than non-invasive methods that are safer for children but inadequate for this study (Silasi et al., 2009; Raboel et al., 2012; Murtha et al., 2012; Uldall et al., 2014). Mice are particularly useful in that like other mammals, they have many similarities to humans in terms of calvarial morphology and genome (Morriss-Kay and Wilkie, 2005), their genetics is well characterized and there are models available to investigate the pathogenesis of various craniofacial deformities. Despite a long-standing interest in skull development in mice (e.g. Fong et al., 2003; Henderson et al., 2005) and rats (e.g. Jones et al., 1987), to the best of our knowledge, intracranial pressure during normal mouse development has not been quantified previously.

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Such data can be used to enhance our understanding of the biomechanics of normal calvarial growth and possibly, ultimately, management of related congenital diseases. Therefore, the aims of this study were to develop a suitable ICP measurement system and quantify ICP during wild type mouse development. To highlight morphological changes during development one sample per age group was scanned and analyzed.

2. Materials and methods

A pressure monitoring system was developed to measure ICP in mice of 5 age groups. ICP was recorded while animals were anesthetised. Following the recording animals were decapitated while still under anesthesia. Then, the sample with closest ICP to the average ICP for each age group was selected for morphological analysis.

2.1. Pressure monitoring system

A 22-gauge needle (outer diameter 0.70 mm; length 6 mm) was connected via luer-lock to silicone tubing (outer diameter 4 mm; length 250 mm) which was then connected to a differential pressure sensor (TruStability® Board Mount Pressure Sensors: HSC Series, Honeywell, NJ, USA). The measurement range of the sensor was ± 18.68 mmHg with total error band of 0.19 mmHg. The signal, i.e. changes in the voltage due to external pressure at the tip of the needle, was acquired at 100 Hz using a custom program written in LabVIEW 2013 (National Instruments Corp, Austin, TX, USA). The pressure measurement system was calibrated using tubes with 50, 70, 100, 120 and 150 mm of water, with each test repeated five times.

2.2. In vivo recording of ICP

A total of 73 inbred wild type mice (*Mus musculus*, C57BL/6J - Jackson Labs, Bar Harbor, Maine, USA) at postnatal (P) day 3 (2.23 ± 0.27 g), 10 (5.05 ± 1.1 g), 20 (9.06 ± 1.48 g), 31 (17.75 ± 1.91 g) and 70 (22.46 ± 4.01 g) were used. Sex was not recorded for the younger groups, but a retrospective statistical analysis comparing the ICP between males and females at P31 did not show a significant difference. The P70 mice were all female. All protocols were approved by the Institutional Animal Care and Use Committees of the University of Washington and Seattle Children's Research Institute. Mice were anesthetized using isoflurane in a non-rebreathing custom set-up. During testing heat support was provided via a warm water pad. Once the animals did not respond to toe pinch, a sagittal incision was made over the calvaria. The needle was inserted through the left parietal bone ca. 2 mm lateral to the sagittal suture and 2 mm anterior to the lambdoid suture. With care it was possible to penetrate the bone with the needle even in older animals, but it was important not to enlarge the hole beyond its diameter. The needle was inserted to a depth calculated to position it in the subarachnoid space, which is filled with cerebrospinal fluid. No external pressure was applied to the skull once the needle had been inserted. It was held in place until ICP reached a maximum (typically 1–2 min); when ICP began to drop, or after several minutes if it did not drop, the needle was removed and the maximum recorded pressure was reported. Once recording was completed, the animals were decapitated while still under anaesthesia.

2.3. Statistical analysis

Statistical analysis was performed in SPSS (IBM SPSS, NY, USA). One-way analysis of variance (ANOVA) with post-hoc Bonferroni and Tukey tests was carried out, with Levene's test used to test for equal variances. The significance level was set at $p < 0.05$.

2.4. Ex vivo micro-computed tomography

The specimen with measured ICP closest to the average ICP value of each age group was scanned using an X-Tek HMX 160 micro-CT scanner (XTek Systems Ltd. Hertfordshire, UK) with a voxel size of 0.01 mm in x, y, and z directions. AVIZO (FEI Visualization Sciences Group, Merignac Cedex, France) was used to reconstruct three dimensional models. The scans were automatically aligned with respect to each other in AVIZO based on minimization of the root mean square distance between the nodes forming the triangulated surfaces of the skull (i.e., Procrustes method) using an iterative closest point algorithm. Each skull surface was typically consisted of about 300,000 nodes. Skull length, width and intracranial volume (ICV) were measured using the software.

3. Results

3.1. Sensor calibration

As the needle was gradually moved down the tube of water, voltage gradually increased, plateauing at the bottom of the tube. Similarly, upon removal from the water, voltage decreased to its baseline value (Fig. 1A). Calibration of the sensor at various heights of water (each repeated five times) showed that the corresponding voltage changes were stable, repeatable and linear (Fig. 1B). Note the error bars corresponding to one standard deviation (of five repeats) are shown in Fig. 1B. These values were in the range of 0.004–0.01 V. These calibration data were used to convert the voltage changes during ICP measurement to mmHg.

3.2. ICP measurements

ICP was 1.33 ± 0.87 mmHg at P3, increasing to 1.92 ± 0.78 mmHg at P10, 3.60 ± 1.08 mmHg at P20, 3.81 ± 1.14 mmHg at P31 and 4.11 ± 0.83 mmHg at P70. There were statistically significant differences between P3 vs. P20, P31, P70, and P10 vs. P20, P31, P70, but not between P20, P31 and P70 (Fig. 2).

3.3. Morphological changes

Skull length was 13 mm in the P3 skull and 17, 19, 20 and 22 mm at P10, P20, P31 and P70 respectively. Skull width increased to a lesser extent from 8 mm at P3 to 11 mm at P70. ICV

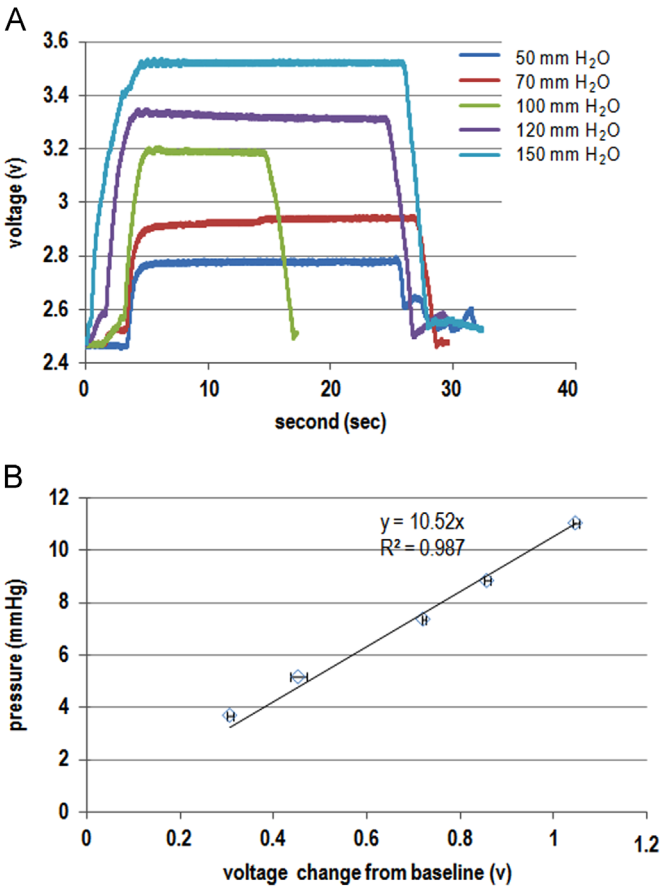

Fig. 1. (A) Testing of the pressure sensor with varying heights of water. The needle was slowly inserted to the bottom of a tube of water, held there for 10–25 s, and then slowly removed. (B) Calibration of the pressure sensor showed the response was linear. Small brackets indicate the SD of measurements.

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