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Oral delivery of an anti-diabetic peptide drug via conjugation and complexation with low molecular weight chitosan

Q13 Sukyung Ahn^a, In-Hyun Lee^a, Eunhye Lee^b, Hyungjun Kim^c, Yong-Chul Kim^c, Sangyong Jon^{a,*}

^a KAIST Institute for the BioCentury, Department of Biological Sciences, Korea Advanced Institute of Science and Technology, Daejeon 305-701, Republic of Korea

5 ^b Utah-Inha DDS & Advanced Therapeutics Research Center, Incheon 406-840, Republic of Korea

6 ^c School of Life Sciences, Gwangju Institute of Science and Technology, Gwangju 500-712, Republic of Korea

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ABSTRACT

Despite the therapeutic potential of exendin-4 as a glucagon-like peptide-1 (GLP-1) mimetic for the treatment of 23 type 2 diabetes, its utility has so far been limited because of the low level of patient compliance due to the 24 requirement for frequent injections. In this study, an orally available exendin-4 was produced by conjugating 25 it to low molecular weight chitosan (LMWC). Conjugation between the LMWC and cysteinylated exendin-4 26 was carried out using a cleavable linker system in order to maximize the availability of the active peptide. The 27 LMWC-exendin-4 conjugate formed a nanoparticle structure with a mean particle size of 101 \pm 41 nm through 28 complexation between the positively charged LMWC backbone and the negatively charged exendin-4 of individ- 29 ual conjugate molecules. The biological activity of the LMWC-exendin-4 conjugate was evaluated in an INS-1 cell 30 line. The LMWC-exendin-4. From the pharmacokinetic study after oral administration of the conjugate, a C_{max} value of 32 344 pg/mL and a T_{max} of 6 h were observed, and the bioavailability, relative to the subcutaneous counterpart, 33 was found to be 6.4%. Furthermore, the absorbed exendin-4 demonstrated a significantly enhanced hypoglyce- 34 mic effect. These results suggest that the LMWC-exendin-4 conjugate could be used as a potential oral anti- 35 diabetic agent for the treatment of type 2 diabetes.

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42 1. Introduction

Patients with type 2 diabetes have greatly impaired incretin-mediated 43 insulin secretion, mainly owing to decreased secretion of glucagon-like 44 peptide-1 (GLP-1) [1]. Exendin-4 is a GLP-1 mimetic peptide comprising 45 46 39 amino acids that has been used in the treatment of type 2 diabetes as it has a significantly enhanced half-life in vivo compared to endoge-47 nous GLP-1, and similar gluco-regulatory activity [2,3]. Even though 48 exendin-4 has been widely used for the treatment of type 2 diabetes, 4950patients are required to undergo frequent subcutaneous injections, resulting in poor patient compliance in addition to side effects such as 51infection at the sites of injection [4,5]. Since oral delivery is expected 5253 to result in drastically enhanced patient compliance, many attempts have been made to develop oral delivery systems for exendin-4 [6-8]. 54 However, orally administered peptides encounter formidable barriers 5556to absorption into the blood stream. These typically include physical barriers, such as viscous mucous layers and tight junctions of aligned 57enterocytes in the gastrointestinal (GI) track; chemical barriers, such 58as low stomach pH; and biological barriers, such as enzymatic degrada-5960 tion. Overcoming these problems is essential for improving the level of absorption of orally administered peptide-based drugs [9,10]. There 61

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have been a limited number of successful studies into the development 62 of systems for oral exendin-4 delivery. It has been shown that site- 63 specific covalent attachment of biotin to exendin-4 facilitated receptor- 64 mediated intestinal absorption of the peptide, resulting in approximately 65 3.95% oral bioavailability (BA) [7]. Very recently, a relatively high oral BA 66 of ~14.0% was attained using a nanoparticulate system formed by electro- 67 static complexation between negatively charged exendin-4 and positively 68 charged chitosan, and subsequent coating with anionic poly(γ -glutamic 69 acid) [8]. Unlike previous approaches, we have sought to overcome such 70 oral delivery barriers using chemical conjugation between the therapeutic 71 agent of interest and a mucoadhesive polymer. In the last few years, a 72 large number of mucoadhesive materials have been investigated for oral 73 delivery of peptides and proteins, including chitosan, methacrylate, 74 and alginate, based polymers [11–16]. Rekha and Sharma reported 75 that anionic/hydrophobic modification of chitosan (LSC) enhanced 76 mucoadhesivity of the nanoparticles and orally administered FITC- 77 insulin loaded LSC particles to diabetic rats showed enhanced insulin 78 absorption and transported insulin across the enterocytes efficiently. 79 We have shown that intestinal absorption of the therapeutic agent 80 could be significantly enhanced by conjugating it to low molecular 81 weight chitosan (LMWC) [17-19]. Such high absorption is attributed 82 to unique features of LMWC, including its high mucoadhesiveness and 83 its ability to open tight cell-cell junctions, facilitating paracellular trans- 84 port of drugs [20-24]. Our previous results suggest that the conjugation 85

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^{*} Corresponding author. Tel.: +82 42 350 2634; fax: +82 42 350 4450. *E-mail address:* syjon@kaist.ac.kr (S. Jon).

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S. Ahn et al. / Journal of Controlled Release xxx (2013) xxx-xxx

of chemical or peptide-based drugs to LMWC may be an efficient strategy for enabling a high level of absorption into the blood stream [18].

In this regard, here we report the development of an oral delivery method for exendin-4 by conjugating it to LMWC. We describe the synthesis and characterization of the conjugate, its activity in the induction of *in vitro* cellular insulin secretion, and the pharmacokinetic parameters and glucoregulatory effect after oral administration in a diabetic mouse model.

2. Materials and methods

2.1. Materials

96 Custom synthesized cysteinylated exendin-4 (exendin-4-cys; N'-HGEGTFTSDLSKQMEEEAVRLFIEWLKNGGPSSGAPPPSC-C') was purchased 97 from AnyGen Co. (Gwangju, South Korea). Low molecular weight chito-98 san (LMWC) was purchased from KITTOLIFE (Seoul, South Korea) and 99 further purified by ultrafiltration before use. N-succinimidyl 3-(2-100 pyridyldithio)-propionate (SPDP) and tris(2-carboxyethyl)phosphine hy-101 drochloride (TCEP) immobilized bead for disulfide reduction were pur-102 chased from Pierce (Rockford, IL). All other solvents and reagents were 103 obtained from Sigma-Aldrich (St. Louis, MO) unless otherwise indicated. 104 105 Male C57BL/6 *db/db* mice (7–8 weeks old) were supplied by the Korean Research Institute of Bioscience and Biotechnology (Daejeon, South 106 Korea) and maintained in pathogen-free conditions in the animal facility 107 at Gwangju Institute of Science and Technology (GIST). The animal exper-108 iments were approved by the GIST Animal Care and Use Committee. The 109 110 exendin-4 ELISA kit was purchased from Phoenix pharmaceuticals, Inc. (Burlingame, CA) and insulin ELISA kits were purchased from Mercodia 111 (Uppsala, Sweden). The One-touch blood glucose meter (CodeFree) 112 was purchased from SD Biosensor, Inc. (Suwon, South Korea). The 113 114 glucose-sensitive pancreatic cell line (INS-1) was kindly donated by 115Prof. Kang Choon Lee (Sung Kyun Kwan University, Suwon, South Korea) and was cultured in RPMI 1640 medium (Life Technologies, 116 Grand Island, NY) containing 10% heat-inactivated fetal bovine serum 117 (FBS) (Life Technologies, Grand Island, NY), 100 units/mL penicillin, 118 and 100 µg/mL streptomycin at 37 °C in a humidified atmosphere of 119 120 5% CO₂.

121 2.2. Synthesis of the LMWC-PDP conjugate

122 A solution of SPDP (30 mg) dissolved in anhydrous DMF (5.6 mL, 50 mM) was added to a solution of LMWC (168 mg, average $M_{w} =$ 123 9 kDa) dissolved in 2.8 mL of sodium tetraborate buffer (10 mM, pH 9). 124 The mixture was stirred in an oil bath at 37 °C for 5 h. Distilled water 125(33.6 mL) was then poured into the reaction mixture and unreacted 126127SPDP was subsequently extracted using ethyl acetate (42 mL, 5 times). The aqueous layer was collected and dialyzed against distilled water 128 using a dialysis membrane tube with a MWCO of 5 kDa (Spectrum 129Laboratories, Rancho Dominguez, CA). After freeze drying for 2 days, 130LMWC-PDP (120 mg) was obtained as light yellow powder in 69% yield 131 132and further characterized using ¹H NMR (400 MHz, Jeol, Tokyo, Japan) 133 and UV spectrophotometry (Varian, Palo Alto, CA).

134 2.3. Synthesis of the LMWC-exendin-4 conjugate

Exendin-4-cys (5 mg, 0.12 mmol) dissolved in distilled water (1 mL) 135was treated with a 10-fold molar excess of TCEP gel as a reducing agent 136at ambient temperature for 30 min. The excess TCEP gel was then 137 removed before further reaction. The pre-treated exendin-4-cys was 138 added to a solution of LMWC-PDP (48.8 mg) in PBS-EDTA buffer 139(pH 7.4) and reacted for 1 h at ambient temperature. Following the reac-140 tion, distilled water (8 mL) was poured into the reaction mixture, the 141 pyridine-2-thione leaving group was extracted using ethyl acetate 142 (10 mL, 5 times), and the solvent was evaporated using a centrifugal vac-143 144 uum system (Hanil, Seoul, Korea). The resulting viscous material was purified using reverse phase high performance liquid chromatography 145 (RP-HPLC; Shimadzu, Kyoto, Japan) on a C 18 column (150 mm \times 146 4.6 mm Symmetry, Waters, Milford, MA) with a mobile phase consisting 147 of 0.1% trifluoroacetic acid (TFA) in water and acetonitrile. The mobile 148 phase was run with a linear gradient from 5 to 65% for 30 min at a 149 flow rate of 1 mL/min and the detection wavelength was 230 nm. 150

2.4. Characterization of the LMWC-exendin-4 conjugate nanoparticles 151

The particle size and zeta potential of LMWC-exendin-4 were 152 measured before and after freeze drying using electrophoretic light 153 scattering apparatus (ELS 8000, Otsuka Electronics, Japan). The morphology was examined using transmission electron microscopy 155 (TEM) using a TECNAI F 20 electron microscope (Philips Electronic 156 Instruments Corp., Mahwah, NJ).

2.5. Proteolytic stability test

The LMWC-exendin-4 conjugate or exendin-4-cys (200 μ g/mL ¹⁵⁹ each) was mixed with an equal volume of trypsin solution (Life Technologies, Grand Island, NY) at 37 °C, and further incubated at 37 °C ¹⁶¹ for predetermined times. The reactions were then stopped by adding ¹⁶² 200 μ L of 1% TFA in H₂O. An aliquot of the reaction solution was taken ¹⁶³ and centrifuged at 1500 g for 5 min. The residual amount of each ¹⁶⁴ exendin-4 species in the supernatant was analyzed using RP-HPLC. ¹⁶⁵

2.6. In vitro biological activity test

The biological activity of the LMWC-exendin-4 conjugate was determined through the measurement of insulin secretion from the glucose 168 sensitive pancreatic-cell line (INS-1) after treatment with the compound. 169 The cells were maintained in complete RPMI 1640 medium (11.1 mM 170 glucose) containing 10% (v/v) FBS, 50 μ M 2-mercaptoethanol, 10 mM 171 HEPES, 2 mM glutamine, 1 mM sodium pyruvate, 100 units/mL penicil-172 lin, and 100 μ g/mL streptomycin and incubated at 37 °C under 5% CO₂ 173 atmosphere. *In vitro* biological activity was evaluated by incubating the 174 INS-1 cells in 500 μ L of Krebs-Ringer-HEPES (KRH) buffer (11.1 mM 175 glucose) containing exendin-4 or the LMWC-exendin-4 conjugate (1–10 nM) for 1 h or 6 h. Levels of insulin released were measured 177 using an insulin ELISA kit. 178

2.7. In vivo pharmacokinetics

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The pharmacokinetic profile of exendin-4-cys and the LMWC- 180 exendin-4 conjugate was assessed as follows. Either exendin-4-cys 181 (50 µg/kg, s.c.) or LMWC-exendin-4 (400 µg/kg, p.o.) was adminis- 182 tered to C57BL/6 mice (n = 5). Blood samples were withdrawn from 183 the retro-orbital sinus and centrifuged (700 g at 4 °C for 20 min). The 184 supernatant was filtered by using SEP-Column (RK-SEPCOL-1, phoenix 185 pharmaceuticals Inc., Burlingame, CA) to remove plasma proteins. 186 The filtered plasma was treated with TCEP bead for 1 h to cleave the 187 disulfide linkage between LMWC and exendin-4-cys and centrifuged 188 (700 g at 4 °C for 20 min). The concentration of exendin-4-cys in the 189 collected supernatant was measured using an exendin-4 ELISA kit and 190 calculated based on a standard curve for exendin-4-cys. The pharmaco- 191 kinetic parameters were estimated using a WinNolin software package. 192 The relative BA of LMWC-exendin-4 conjugate after oral administration 193 was calculated using the following formula: $[(AUC_{oral} \times Dose_{sc}) / 194]$ $(AUC_{sc} \times Dose_{oral}) \times 100$ [25]. 195

2.8. Intraperitoneal glucose tolerance test (IPGTT)

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The *in vivo* anti-diabetic (hypoglycemic) activity of LMWC-exendin-4 197 conjugate was measured in *db/db* mice after oral drug administration 198 (n = 5) by IPGTT [26]. The average body weight and blood glucose levels 199 were the same for every group. Briefly, mice fasted for 18 h were orally 200

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