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Original article

N,O-Bis(trimethylsilyl)acetamide/*N*-hydroxysuccinimide ester (BSA/NHS) as coupling agents for dipeptide synthesis

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ABSTRACT

A method using *N,O*-bis(trimethylsilyl)acetamide/*N*-hydroxysuccinimide ester (BSA/NHS) as coupling agents for dipeptide synthesis is descried. The coupling reaction between *N*-hydroxysuccinimide (NHS) esters and amines could be performed under mild conditions with *N,O*-bis(trimethylsilyl)acetamide (BSA) as coupling reagent and no additional acid/base is required. All byproducts and excessive reactants are water soluble or hydrolysable and easy to eliminate through water-washing at the purification stage. Moreover, all the reactants are inexpensive and widely used in conventional drug production.

1. Introduction

Since du Vigneaud published the first solution-phase synthesis of oxytocin in 1953 [1], it has been found that peptides participate in various biocatalytic processes [2]. With assistance of recent advances in synthetic [3] and drug delivery technologies [4], peptide-based anti-cancer [5], anti-diabetic [6], anti-microbial [7], anti-fungal [8] drugs and intrinsic hormonal analogues have been discovered [9]. Therefore, it is imperative to develop large scale synthetic approaches for the production of complex peptides.

Traditional solution-phase methodologies are the most widely used approaches for industrial production of approved peptide-based pharmaceuticals. In general, they have no scale limitations, and each step can be monitored precisely. However, significant shortcomings of solution-phase methodologies, such as consumption of a large amount of organic solvents, tedious protection/de-protection steps and difficult purification processes, still hamper the development of scale-up routes of peptide syntheses. Solid-phase methodologies, in which the peptides of relatively complex sequences could be synthesized using automated and rapid synthesis/workup procedures, were pioneered by Bruce Merrifield to overcome the problems in traditional solution-phase methodologies [10]. However, after decades of optimization, shortcomings of solid-phase synthesis still limited its applications in commercial-scale manufacture. For example, lack of scalability, inadequate in-process controls and low purity of the final products still need improvement [11]. Alternative approaches consisting of soluble polymer-tagged liquid-phase reactions have also been developed to ensure both high reaction rate and real time reaction monitoring [12]. In this method, peptide syntheses could be performed in the solution phase but work-up is accomplished in a manner similar to solid-phase synthesis where most of the excess organic reagents and activated amino acids are removed by simple solidification and filtration procedures. However, the high-cost of soluble polymers, the time-consuming process of solidification/crystallization in each coupling cycle and the inherent scale limitation as observed in solid-phase methodologies represent serious challenges for large scale synthesis of peptides.

Here we describe a simplified dipeptide synthesis strategy using the *N*-hydroxysuccinimide (NHS) ester of amino acids and a silylation agent *N,O*-bis(trimethylsilyl)acetamide (BSA) as a coupling agent (Fig. 1) The coupling reactions between NHS esters and amines occur efficiently under mild conditions with BSA only and no additional acid/base is required. Besides, excessive BSA and other byproducts are water soluble or hydrolysable [13] and easy to remove by simple water-washing at the purification stage. Consequently, less racemization, fewer units of operation, and simpler purification processes can be achieved compared to other solution-phase methodologies.

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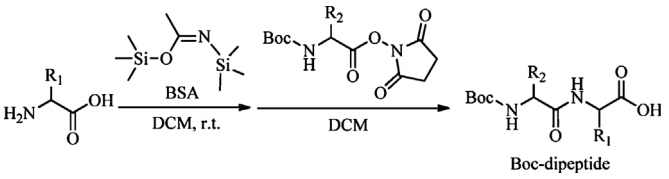

Fig. 1. Synthesis of *t*-butyloxycarbonyl (Boc)-protected dipeptide using BSA and NHS ester as coupling agents.

2. Experimental

All reactions were performed under a nitrogen atmosphere using anhydrous techniques unless otherwise noted. ^{1}H NMR (300 MHz) on a Varian Mercury 300 spectrometer was recorded in DMSO-d_6 or $CDCl_3$. Chemical shifts are reported in δ (ppm) units relative to the internal standard tetramethylsilane (TMS). All the reactions were monitored by thin-layer chromatography (TLC) analysis on pre-coated silica gel G plates at 254 nm under UV lamp or HPLC analysis.

2.1. General procedure for the preparation of N-Boc protected dipeptide

Under argon protection, BSA (2.2 equiv.) was added to a solution of amino acid (1.1 equiv.) in anhydrous dichloromethane. After the mixture was stirred for 1–8 h at 23 °C, a solution of *N*-Boc protected NHS ester (1 equiv.) in dichloromethane was added. The reaction mixture was stirred at 23 °C under argon until all active ester was consumed as judged by TLC analysis. The reaction mixture was washed with brine, dried over Na_2SO_4 and concentrated *in vacuo* to provide a white solid. The isolated product was recrystallized from diethyl ether/*n*-hexane to yield the targeted dipeptide as a white solid.

N-Boc-L-phenylalanine-L-proline (Boc-Phe-Pro-OH): ESI-MS (*m/z*): 363.2 $[M+H]^+$; 307.1 $(M-(CH_3)_2C=CH_2)$. ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.26 (m, 5*H*), 5.39 (d, 1*H*, *J* = 8.6 Hz), 4.69–4.49 (m, 2*H*), 3.63–3.52 (m, 1*H*), 3.01 (m, 3*H*), 2.28–2.16 (m, 1*H*), 2.10–1.98 (m, 1*H*), 1.86 (m, 2*H*), 1.39 (s, 9*H*).

N-Boc-L-alanine-L-proline (Boc-Ala-Pro-OH): ESI-MS (*m/z*): 287.2 $[M+H]^+$; 231.1 $(M-(CH_3)_2C=CH_2)$. ^{1}H NMR (400 MHz, $CDCl_3$): δ 4.60 (dd, 1*H*, *J* = 8.1, 3.9 Hz), 4.49 (s, 1*H*), 3.73 (q, 1*H*, *J* = 8.0 Hz), 3.59 (m, 1*H*), 2.31–2.00 (m, 4*H*), 1.43 (s, 9*H*), 1.34 (d, *J* = 6.9 Hz, 3*H*).

N-Boc-L-alanine-L-phenylalanine (Boc-Ala-Phe-OH): ESI-MS (*m/z*): 337.2 $[M+H]^+$; 281.1 $(M-(CH_3)_2C=CH_2)$. ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.32–7.10 (m, 5*H*), 6.85 (d, 1*H*, *J* = 7.5 Hz), 5.36–5.06 (m, 1*H*), 4.82 (q, 1*H*, *J* = 6.5 Hz), 4.21 (s, 1*H*), 3.20 (dd, 1*H*, *J* = 14.0, 5.5 Hz), 3.03 (dd, 1*H*, *J* = 14.3, 6.4 Hz), 1.43 (s, 9*H*), 1.26 (s, 3*H*).

N-Boc-L-leucine-L-phenylalanine (Boc-Leu-Phe-OH): ESI-MS (*m/z*): 379.2 $[M+H]^+$; 323.2 $(M-(CH_3)_2C=CH_2)$. ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.29–7.11 (m, 5*H*), 6.97–6.78 (m, 1*H*), 5.13 (d, 1*H*, *J* = 8.8 Hz), 4.92–4.73 (m, 1*H*), 4.21 (d, 1*H*, *J* = 8.2 Hz), 3.26–3.10 (m, 1*H*), 2.98 (dd, 1*H*, *J* = 13.9, 6.4 Hz), 1.59 (m, 2*H*), 1.44 (m, 10*H*), 0.89 (t, 6*H*, *J* = 7.0 Hz).

N-Boc-L-isoleucine-L-valine (Boc-Ile-Val-OH): ESI-MS (*m/z*): 331.2 $[M+H]^+$; 275.2 $(M-(CH_3)_2C=CH_2)$. ^{1}H NMR (400 MHz, $CDCl_3$): δ 6.87 (d, 1*H*, *J* = 8.7 Hz), 5.34 (d, 1*H*, *J* = 9.0 Hz), 4.62 (dd, 1*H*, *J* = 8.6, 4.8 Hz), 3.97 (t, 1*H*, *J* = 8.2 Hz), 2.00 (m, 2*H*), 1.43 (s, 9*H*), 1.22 (m, 2*H*), 0.93 (m, 12*H*).

3. Results and discussion

In order to avoid racemization under alkaline conditions for the deprotection step, all the *N*-terminus of NHS ester were protected by Boc group. Most *N*-Boc protected NHS ester could be purchased, those commercially unavailable *N*-Boc protected NHS esters were readily obtained by coupling the corresponding *N*-Boc protected amino acids with *N*-hydroxysuccinimide (NHS-OH) in the presence of *N*,*N*-dicyclohexylcarbodiimide (DCC) [14]. The resulting byproducts containing the dicyclohexyl urea could be removed by filtration though a short pad of silica gel. After concentration of the filtrate, pure active esters could be recrystallized from various solvent systems. The solid *N*-Boc protected NHS esters were stable at −4 °C for a long period of time.

Table 1
Synthesis of *N*-Boc protected dipeptides *via* the BSA/NHS method[a].

Entry	Product	Time (h)	Solvent	BSA/AA/NHS ester[b]	Yield (%)[c]
1	Boc-Phe-Pro-OH	48	DCM	−/1.1/1	Trace
2	Boc-Phe-Pro-OH	8	DCM	2.2/1.1/1	94.3
3	Boc-Phe-Pro-OH	12	DCM	4.4/1.1/1	64.4
4	Boc-Phe-Pro-OH	24	DCM	1.1/1.1/1	58.7
5	Boc-Phe-Pro-OH	8	THF	2.2/1.1/1	44.5
6	Boc-Phe-Pro-OH	24	THF	2.2/1.1/1	78.8
7	Boc-Phe-Pro-OH	8	DMF	2.2/1.1/1	63.2
8	Boc-Phe-Pro-OH	24	DMF	2.2/1.1/1	83.2
9	Boc-Leu-Phe-OH	10	DCM	2.2/1.1/1	82.1
10	Boc-Ala-Phe-OH	10	DCM	2.2/1.1/1	81.5
11	Boc-Ala-Pro-OH	8	DCM	2.2/1.1/1	91.1
12	Boc-Ile-Val-OH	16	DCM	2.2/1.1/1	85.4

[a] Reaction conditions: unprotected amino acid (AA) reacted with BSA at room temperature, followed by the addition of *N*-Boc protected NHS ester (NHS ester).
[b] The molar ratio of BSA to amino acid and NHS ester.
[c] Isolated yield.

The coupling conditions and purification processes were optimized first through the synthesis of various dipeptides (Table 1). When no BSA was added, the coupling product was hardly detected (entry 1). Through screening various reaction conditions for the synthesis of Boc-Phe-Pro-OH, we found that the coupling efficiency and yield were optimal when 1.1 equiv. of unprotected proline reacted with 2.2 equiv. of BSA first in dichloromethane (DCM) at room temperature, followed by the addition of 1 equiv. of *N*-Boc protected Phe NHS ester (enrty 2). The ratio of BSA was important that either excessive (entry 3) or insufficient (entry 4) BSA would reduce the coupling yield significantly. Meanwhile, the ratio of each reagent was important not only for coupling efficiency, but also for purification process. NHS esters were insoluble in water, but soluble in organic solvents. In contrary, unprotected amino acid and BSA were either soluble in water or easily hydrolyzed in water. Therefore, the molar quantity of the NHS ester should be slightly lower than the unprotected amino acid and BSA to guarantee the NHS ester to be exhausted completely. Then the excessive unprotected amino acids and BSA could be easily removed by simply washing with water.

All the unprotected amino acids are insoluble in organic solvents. So the addition order of reagents is quite important and unprotected amino acids should react with BSA first to increase its solubility and nucleophilicity. According to our data, the solubility of most amino acids improved significantly after being silylated with BSA. Among them, the silylation of proline was the fastest and it became soluble in dichloromethane after just 1 h reaction with BSA. But it took hours for other unprotected amino acids to be silylated and dissolve in dichloromethane. Therefore, the slightly lower yield when C-terminal was unprotected amino acids other than proline may be owing to their relatively poorer solubility. For the same reason, when C-terminal was several unprotected hydrophilic amino acids, such as aspartic acid, glutamic acid and cysteine, nearly no dipeptide products were obtained. Besides, unprotected basic amino acids that have two amino groups, such as arginine, lysine and histidine, are also unsuitable for the BSA/NHS method. The solubility of silylated amino acid in THF and DMF was