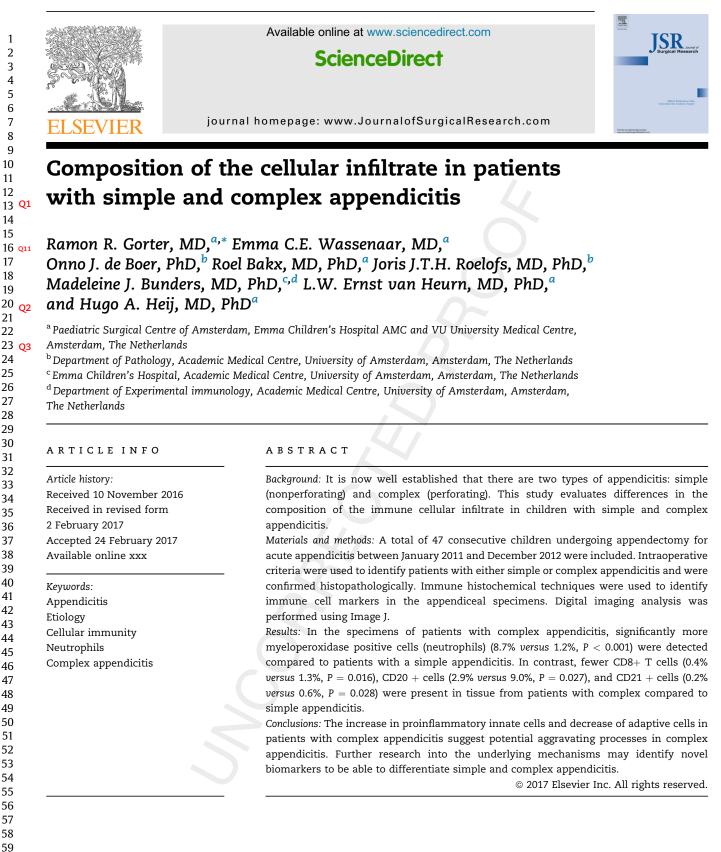
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131 132 Introduction

Recent studies indicate that acute appendicitis is not an 134 irreversibly progressive disease but that two distinct types of 135 appendicitis can be identified: simple (uncomplicated or 136 137 nonperforating appendicitis) and complex (complicated or 138 perforating) appendicitis. Simple appendicitis can be treated 139 with antibiotics only, whereas the second requires 140 an appendectomy in most cases.^{1,2} Initial nonoperative 141 treatment has been investigated for acute simple appendicitis 142 both in the adult and pediatric population with good results.³⁻⁸ 143 Approximately, in 60%-85% of the adult population and 144 62%-81% of the pediatric population, an appendectomy was 145 avoided at 1-y follow-up after treatment with antibiotics 146 only.³⁻⁸ Selection of patients in these studies was based on 147 148 clinical and radiological variables and did not include 149 biochemical markers. Accurate identification of patients with 150 simple or complex appendicitis will prevent unnecessary 151 surgical interventions in patients with simple appendicitis, 152 while making sure patients with complex appendicitis receive 153 the surgical treatment that they require.

154 To identify biomarkers, which help to distinguish simple 155 and complex appendicitis, it is essential to better understand 156 the individual underlying pathogenesis of both types. These 157 are predisposing factors for appendicitis in general, such as 158 appendicular obstruction, infection, diet, and ischemia.⁹ All 159 160 the previously mentioned factors eventually lead to invasion 161 of the appendiceal wall by intraluminal bacteria and activate 162 innate immune cells such as macrophages, whereas danger 163 signals due to ischemia-induced cell death can further 164 augment immune responses.9-11 For example, increase of 165^{Q4} extracellular ATP due to cell death in ischemic tissue is a 166 strong inducer for innate cells such as macrophages and 167 dendritic cells (DCs), which together with Toll like receptors 168 (TLRs) triggering results in interleukin (IL)-1b production.^{12,13} 169 T cells activated by DC are subsequently recruited to the site 170 171 of inflammation and can further contribute to inflammation 172 or by means of T regulatory cells decrease inflammation. 173 However, it is unknown why in one person this inflammation 174 is impeded while in others leads to complex appendicitis.

175 Studies focusing on the composition of the cellular 176 infiltrate in patients with appendicitis are scarce.^{14,15} It has 177 been noted that in all patients with appendicitis, there is an 178 influx of neutrophils in the lamina propria.¹⁴ Only one study 179 compared the composition of the cellular infiltrate in patients 180 with perforated appendicitis and nonperforated appendicitis 181 and detected increased numbers of cluster of differentiation 182 183 (CD)-8+ T cells in the specimens of the appendix of patients 184 with perforated appendicitis.¹⁵ Subsequent studies mainly 185 focused on the systemic immune response and investigated 186 the cytokine profiles in blood samples showing elevated levels 187 of IL-6, IL-17, and interferon α (INF- α) in patients with complex 188 appendicitis compared to simple appendicitis.^{16,17} 189

The aim of this study is to evaluate differences in the composition of the infiltrate of mononuclear immune cells in the appendix between patients with simple and complex appendicitis. The results will help identify critical cell types distinguishing between these phenotypes and can be used to design further studies to identify new biomarkers.

Materials and methods

Study population

All children aged 0-17 y who underwent an appendectomy for suspected acute appendicitis in the Academic Medical Center, Amsterdam between January 2011 and December 2012 were included. Patients with a noninflamed appendix, a parasitic infection, those who underwent an appendectomy as a routine procedure (for instance in case of malrotation), or as an interval appendectomy after initial nonoperative treatment of appendicitis were excluded. The medical charts from the included patients were reviewed, and the specimens from the original histopathological examination of the appendix were collected, and additional staining procedures, as described in the following, were performed. Patients without histopathological specimens, missing data, or those unsuitable for staining were also excluded. Study approval was obtained from the medical ethics review board before the start of this study.

Patients were allocated into either the simple or complex appendicitis group according to the following definitions for simple and complex appendicitis.^{18,19}

Simple appendicitis is diagnosed on the basis of (1) intraoperative findings: inflamed appendix without signs of gangrene, perforation, purulent fluid, contained phlegmone, or intra-abdominal abscess and (2) histopathological examination confirming the diagnosis of appendicitis without necrosis or perforation. Complex appendicitis is diagnosed on the basis of (1) intraoperative findings: signs of a gangrenous appendix with or without perforation, intra-abdominal abscess, appendicular contained phlegmone, or purulent free fluid and (2) histopathology confirming the diagnosis based on extensive necrotic tissue in the muscular layer of the appendix or signs of perforation. In case of discrepancies between clinical and pathological findings, discussion was held by the pathologist (J.R.) and one member of the surgical team (R.G.). In case of disagreement, a third reviewer was approached.

Medical chart review

A standardized data extraction form was used to review the medical charts (Appendix 1).

Staining and scanning

The original specimens of appendices were handled using a standardized protocol developed by the Department of Pathology, Academic Medical Center of Amsterdam. The hematoxylin and eosin—stained sections and tissue blocks were retrieved from the archive and re-evaluated to identify the most severely affected appendiceal segment based on microscopic examination by three of the authors (E.W., R.G., and J.R.), which was then selected for additional staining. Based on the literature, we decided to use the immune histochemical stains with antibodies specific for immune cells from the innate and adaptive immune response, and these are listed in Table 1.

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