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Parasitology International xxx (2015) xxx-xxx



Contents lists available at ScienceDirect

Parasitology International



journal homepage: www.elsevier.com/locate/parint

Q3 Chemoprophylactic and therapeutic efficacy of thymol in murine 2 cystic echinococcosis

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7 ARTICLE INFO

Article history: Received 16 March 2015 Received in revised form 15 June 2015 10 Accepted 17 June 2015 11 12 Available online xxxx 13Kevwords: 14 Cystic echinococcosis Echinococcus granulosus 15Essential oils 16Thymol 17 Albendazole $\frac{1}{29}$ 36

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Q9 1. Introduction

Cystic echinococcosis, an infection with the larval form of the dog tapeworm *Echinococcus granulosus*, is characterized by cystic lesions, most commonly in the liver and lungs [1]. This parasitic infection is a chronic, complex, and still neglected disease [2].

Currently four treatment approaches are in use: surgery, PAIR (punc-39 ture, aspiration, injection of protoscolicidal agent, reaspiration), chemo-40 therapy, and watch and wait for inactive, clinically silent cysts [3]. The 41 drugs commonly used against cystic echinococcosis are benzimidazoles, 42 such as albendazole (ABZ) and mebendazole [4]. Unfortunately, 20%-43 40% of cases do not respond favorably to such chemotherapy and 44 these drugs produce stabilization, rather than cure in the majority of 45 patients [5]. With regard to these difficulties, novel therapeutical tools 46 are needed to optimize treatment of cystic echinococcosis. 47

The control of helminthosis, and generally of all parasitic diseases, is usually made with synthetic anthelmintics. These anthelmintic drugs have some important disadvantages, such as cost and improper use leading to drug resistance, environmental pollution, and food residues [6]. Consequently, the search of new therapeutic alternatives such as the use of traditional medicinal plants has been increased. Aromatic plants have pharmaceutical properties that are in part

Aromatic plants have pharmaceutical properties that are in part attributed to essential oils [7]. For their hydrophobic character, the

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http://dx.doi.org/10.1016/j.parint.2015.06.005 1383-5769/© 2015 Published by Elsevier Ireland Ltd.

ABSTRACT

Cystic echinococcosis is a zoonotic disease caused by the larval stage of the cestode *Echinococcus granulosus*. The 19 drugs commonly used against cystic echinococcosis are benzimidazoles. Unfortunately, 20%–40% of cases do not 20 respond favorably to such chemotherapy. Consequently, the search of new therapeutic alternatives such as the 21 use of traditional medicinal plants has been increased. The aim of the current experimental work was to investi-22 gate the chemoprophylactic and clinical efficacy of thymol on mice infected with *E. granulosus* metacestodes. 23 Thymol (40 mg/kg) was administered under two different therapeutic schemes: dosing every 24 h over 24 20 days and treatment every 12 h for 10 days. Thymol demonstrated efficacy against experimental murine cystic Q6 echinococcosis. The chemoprophylactic and therapeutic effect of thymol was comparable to that of albendazole. Q7 Due to the lack of toxicity observed in mice at the tested doses; we consider that thymol is a potential alternative 27 to be applied for the treatment of human hydatid disease. 28

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essential oils and their components have a considerable potential of 56 pharmacological applications as antimicrobial agents [7,8]. The role of 57 essential oils against parasitic helminths has been studied by several au-58 thors [9–12]. However, there are few publications about the effect of 59 these substances on *E. granulosus*. The in vitro effect of the essential 60 oils of *Rosmarinus officinalis* (rosemary), *Mentha pulegium*, *M. piperita*, **Q10** *Pistacia khinjuk* (pistachio) and *Trachyspermum ammi* (ajowan) has 62 been demonstrated against protoscoleces of *E. granulosus* [13–16]. 63

Thymol (2-isopropyl-5-methylphenol) is one of the major compo-64 nents of the essential oils of *Thymus vulgaris* and *Origanum vulgare* and 65 is a widely known anti-microbial agent [17]. The in vitro and in vivo ac-66 tivities of thymol against *Leishmania panamensis* [18] and *Anisakis simplex* larvae [19] were proven. Moreover, several encouraging findings have been reported using thymol in vitro on *E. granulosus* protoscoleces, 69 microcysts and metacestodes [20,21]. Thymol had anthelmintic effect 70 against the larval stage, altering the viability, structure and ultrastructure of the cestode. 72

The aim of the current experimental work was to investigate the 73 chemoprophylactic and clinical efficacy of thymol on mice infected 74 with *E. granulosus* metacestodes. 75

2. Materials and methods

2.1. Chemicals 77

ABZ suspension (2.1 mg/ml) was prepared by dissolution of ABZ pure 78 standard (Pharmaceutical grade, Parafarm, Buenos Aires, Argentina) in 79

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deionized water (pH = 7.0) under shaking (12 h). ABZ suspension was vigorously shaken before its intragastric administration to mice.

Thymol was obtained from Sigma-Aldrich (USA). Thymol solution (2.4 mg/ml) was prepared by dissolution of the drug in olive oil and maintained under refrigeration (3 to 5 °C).

85 2.2. Protoscolex collection

Protoscoleces of *E. granulosus* were collected aseptically from liver and lung hydatid cysts of infected cattle slaughtered in an abattoir located in the southeast of Buenos Aires province, Argentina. The area where the cattle came from is known to include only the G1 strain of *E. granulosus* [22]. Viability was assessed as previously described [23].

91 2.3. Experimental animals and infection

Animal procedures and management protocols were approved by 92the Institutional Animal Care and Use Committee (act 2555-07-14) of 93 94 the Faculty of Exact and Natural Sciences, National University of Mar del Plata, Mar del Plata, Argentina and carried out in accordance with 95 the 2011 revised form of The Guide for the Care and Use of Laboratory 96 Animals published by the U.S. National Institutes of Health. Unnecessary 97 animal suffering was avoided throughout the study. Female CF-1 mice 98 99 $(n = 100; body weight 25 g \pm 5)$ were infected by intraperitoneal inoculation with 1500 E. granulosus protoscoleces/animal, suspended in 100 0.5 ml of medium 199 (Gibco). The animals were housed in a tempera-101 ture controlled (22 ± 1 °C), light-cycled (12-h light/dark 174 cycle) 102 room. Food and water were given ad libitum. 103

104 2.4. Experimental design

Two different experimental designs were conducted: a chemopro phylactic efficacy study (which simulates a cyst rupture during surgical
practice and the concomitant drug treatment) and, a clinical efficacy
study (simulating an experimental secondary hydatidosis).

109 2.4.1. Chemoprophylactic efficacy study

At the time point of infection, the animals were allocated into 5 110 111 experimental groups (n = 10) and treated as follows: a) distilled water control group, animals receiving distilled water as a placebo; 112113b) oil control group, animals receiving olive oil as a placebo; c) ABZ group, animals were treated with ABZ suspension at the dose rates of 114 012 25 mg/kg every 24 h for 30 days; d) thymol group I, animals were treated with thymol at the dose rates of 40 mg/kg every 24 h for 20 days; and 013 e) thymol group II, animals were treated with thymol (40 mg/kg) at 117 12-h intervals over 10 days. Treatments were performed by intragastric 118 administration. Six months after infection, mice were euthanized, and 119 necropsy was carried out immediately thereafter. 120

121 2.4.2. Clinical efficacy study

Protoscoleces of E. granulosus can differentiate to hydatid cysts if 122released within (or passaged into) an intermediate host. Cystic differen-123tiation is completed after 2-3 months [24]. At 6 months post-infection, 124mice were allocated into the following experimental groups (10 125animals/group) and treated as follows: a) distilled water control 126group, animals receiving distilled water as a placebo; b) oil control 127group, animals receiving olive oil as a placebo; c) ABZ group, animals 128 were treated with ABZ suspension at the dose rates of 25 mg/kg every 129Q14 24 h for 30 days; d) thymol group I, animals were treated with thymol Q15 at the dose rates of 40 mg/kg every 24 h for 20 days; and e) thymol group II, animals were treated with thymol (40 mg/kg) at 12-h intervals 132over 10 days. Treatments were performed by intragastric administra-133 tion. At the end of the treatment period, animals were euthanized, and 134135 necropsy was carried out immediately thereafter.

2.5. Determination of parasite weight and morphologic study

At necropsy in both the prophylactic and efficacy studies, the perito-137neal cavity was opened, and the hydatid cysts were carefully removed.138The weight of the cysts collected from each individual animal was re-139corded using an analytical scale. Samples of cysts recovered from each140mouse were processed for scanning and transmission electron micros-141copy (SEM and TEM) as described by Elissondo et al. [25].142

2.6. Statistical analysis

Weight of cysts (reported as mean \pm SD) were compared by means 144 of Kruskal–Wallis (non-parametric ANOVA) followed by Dunn's multiple comparison test. A value of *P* < 0.05 was considered statistically 146 significant. The statistical analysis was performed using the Instat 3.0 147 software program (GraphPad Software, San Diego, CA). 148

3. Results

All of the infected mice (10/10) from the untreated control groups, 151 ABZ-treated group and thymol-treated group I developed metacestodes 152 in the abdominal cavity. On the contrary, in 1 out of the 10 thymol-153 treated group II mice, the infection did not progress. Statistically significant differences (ABZ group: P < 0.001; thymol group I: P < 0.01; thymol group II: P < 0.05) were observed in the weights of cysts recovered from 156 untreated mice compared with those from the treated groups. On the 157 other hand, no statistically significant differences were found between 158 the weights of cysts recovered from the two thymol-treated groups 159 and ABZ-treated group (P > 0.05) (Table 1).

All cysts removed from control mice appeared turgid, showing no 161 observable collapse of the germinal layer, and no changes in ultrastruc- 162 ture were detected by SEM (Fig. 1a). TEM analysis of cysts recovered 163 from the untreated control group revealed the typical features of 164 *E. granulosus* metacestodes, with a distinct acellular outer laminated 165 layer and a germinal layer without alterations (Fig. 1b). In contrast, 166 the ultrastructural study of cysts developed in mice treated with ABZ 167 and thymol revealed a normal laminated layer, but some alterations 168 on the germinal layer were detected (Fig. 2). Studies by SEM revealed 169 that the germinal layer of treated cysts lost the feature multicellular 170 structure (Fig. 2a, c and e). TEM analysis of cysts from treated mice revealed the presence of some signs of degeneration as an increment in 172 the number of vacuoles and lipid droplets (Fig. 2b, d and f).

3.2. Clinical efficacy study

Table 2 summarizes the cyst weights (mean \pm SD) recorded after 175 treatments on the different experimental groups (unmedicated control 176 and treated groups) involved in the clinical efficacy study. Both thymol 177

Table 1

	$\label{eq:chemoprophylactic efficacy study} \hline $Wet weight (g) of cysts$ Mean \pm SD$
Unmedicated control group (olive oil)	12.3 ± 4.2
Unmedicated control group (distilled water)	10.6 ± 2.9
ABZ suspension	$1.3 \pm 0.9^{***}$
Thymol group I	$2.9 \pm 2.3^{**}$
Thymol group II	$3.7 \pm 2^*$

 $\label{eq:transform} \begin{array}{l} *P < 0.05, **P < 0.01, ***P < 0.001; statistically significant differences between treated group to the transformed state of the transformation of transformation of transformation of the transformation of trans$

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