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Research Paper

Experimental investigation and oral bioavailability enhancement of nano-sized curcumin by using supercritical anti-solvent process

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ABSTRACT

The biomedical applications of curcumin (CUR) are limited due to its poor oral bioavailability. In this work, CUR nanoparticles were successfully prepared by combining the supercritical anti-solvent (SAS) process with Tween 80 as a solubilizing agent and permeation enhancer. Different processing parameters that can govern the mean particle size and size distribution of nanoparticles were well investigated by manipulating the types of solvents, mixing vessel pressure, mixing vessel temperature, CO_2 flow rate, solution flow rate and solution concentration. Solid state characterization was done by Fourier Transform infrared spectroscopy, differential scanning calorimetry, dynamic light scattering, scanning electron microscopy, and powder X-ray diffraction study. Solubility and dissolution profile of SAS-processed CUR were found to be significantly increased in comparison with native CUR. Further, a validated ultra-performance liquid chromatographic method with quadrupole-time of flight-mass spectrometry was developed to investigate the pharmacokinetic parameters after a single oral dose (100 mg/kg) administration of CUR (before/after SAS-processed CUR was found to be increased approximately 11.6-fold (p < 0.001) as compared to native CUR.

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1. Introduction

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Abbreviations: µm, micrometer; °C, degree Celsius; Ac, acetone; AcN, acetonitrile; AUC, area under curve; BCS, Biopharmaceutics Classification System; CC, calibration curve; CFR, CO₂ flow rate; cm, centimeter; C_{max}, maximum plasma concentration; CO₂, carbon dioxide; CUR, curcumin; DLS, dynamic light scattering; DSC, differential scanning calorimetry; FT-IR, Fourier Transform infrared spectroscopy; g, gram; h, hour; HPLC, high performance liquid chromatography; HQC, high quality control; IS, internal standard; KV, kilovolt; L, liter; LQC, low quality control; Me, methanol; min, minute; mL, millilitre; MPa, millipascal; MQC, middle quality control; ng, nanogram; nm, nanometer; P, pressure; pA, picoampere; Pa, pascal; Pc, critical pressure; PDI, poly dispersity index; P-gp, P-glycoprotein; QC, Quality Control; Q-TOF-MS/MS, quadrupole-time of flight-mass spectrometry; RESS, rapid expansion of supercritical solutions; s, second; S.D., standard deviation; S. No., serial number; SAS, supercritical anti-solvent; SC, solution concentration; ScCO₂, super critical carbon dioxide; SFR, solution flow rate; SGF, simulated gastric fluid; T, temperature; T_c , critical temperature; T_{max} , time to get maximum plasma concentration; UPLC, ultra performance liquid chromatography; USP, United States Pharmacopoeia; V, volt; v, version.

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For orally-administered drugs, aqueous solubility and intestinal permeability are the two predominant rate-limiting steps for deciding their bioavailability. Restriction on the use of Biopharmaceutics Classification System (BCS) class IV drug mole-54 cules is due to both their poor aqueous solubility and low perme-55 ability. Various formulation approaches (for example, polymeric 56 prodrugs [1,2], co-crystallization [3,4], solid dispersion [5–7], salt 57 formation [8,9], cyclodextrin complexation [10,11], micronization 58 [12,13], and nanocrystalization [14,15]) have been studied for this 59 class of molecules to enhance their oral bioavailability. Curcumin 60 (1,7-bis (4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione) 61 is a polyphenol extracted from the rhizome of Curcuma longa, 62 belonging to the family Zingiberaceae [16]. Apart from its tradi-63 tional use as a spice, natural coloring and flavoring agent, recent 64 preclinical and clinical studies have proved its therapeutic benefits 65 as an antispasmodic, anticoagulant, antimicrobial, anti-oxidant, 66 anti-inflammatory, anti-tumor, anti-HIV and hypocholesterolemic 67 agent for the treatment of various chronic disorders, such as neo-68 plastic, neurological, hepatic, cardiovascular, pulmonary, 69

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M. Anwar et al. / European Journal of Pharmaceutics and Biopharmaceutics xxx (2015) xxx-xxx

70 metabolic and psychological diseases [17–19]. To overcome the 71 above-mentioned bioavailability problems for a BCS class IV mole-72 cule, various nanoparticle-based drug delivery approaches for cur-73 cumin (CUR) have been reported by research scientists [20-22]. 74 Recently, the use of a supercritical fluid as a solvent (rapid expan-75 sion of supercritical solutions; RESS) or an anti-solvent (supercrit-76 ical anti-solvent; SAS) for poorly-aqueous soluble drugs has been 77 set up as a feasible technique for their size reduction to micro or 78 nanoscale. Supercritical carbon dioxide (ScCO₂) is commonly used 79 as a supercritical fluid in SAS and RESS processes due to its advan-80 tages, such as mild critical temperature and pressure (T_c = 31.1 °C, P_c = 7.38 MPa), low viscosity, low surface tension, low cost, 81 non-toxicity, non-flammability, strong solvent power and safety 82 in environmental considerations. 83

84 Due to low solubility of most drug molecules in ScCO₂, use of 85 RESS process is mostly limited, whereas, amorphization [23,24], 86 recrystallization [25,26], micronization [27–29] and nanonization 87 [23,24,30] through SAS process have been successfully achieved. 88 SAS is a one-step efficient process leading to completely dry, smaller particles with narrow size distribution, controlled crystals with 89 90 preferred morphology and better flow-ability, and organic 91 solvent-free product justifying their industrial applications. By 92 reducing the size and increasing the surface area, SAS process 93 can improve a drug's solubility, dissolution rate, and stability in 94 physiological fluid, which may ultimately reduce the dose, dosing 95 frequency and associated toxic effects along with bioavailability 96 enhancement of the drug. CUR also undergoes photodegradation 97 when exposed to light, in solution as well as in solid form [17], 98 placing difficulty in its handling during formulation development 99 process in an open environment. As SAS process operates in a 100 closed environment, it can solve the above-mentioned formulation 101 problems.

102 Our research aimed to improve the oral bioavailability of CUR 103 by reducing its particle size to the nanometric range using SAS pro-104 cess, on combining with Tween 80 as a solubilizer and permeation 105 enhancer [31,32]. In this study, the effect of different organic sol-106 vents and process variables of SAS process on the particle size 107 and size distribution of CUR nanoparticles was investigated.

108 2. Materials and methods

109 2.1. Materials

110 Curcumin (purity > 99%) was purchased from Sigma–Aldrich, St. 111 Louis, MO, USA. Acetone (LC/MS grade), methanol (HPLC grade), 112 acetonitrile (LC/MS grade), sodium lauryl sulfate (extra pure grade) 113 and Tween 80 (extra pure grade) were obtained from S.D. Fine 114 Chemicals Pvt. Ltd. (Mumbai, India). Carbon dioxide (CO₂) (purity 115 99.8%) was purchased from Laser Gases Pvt. Ltd. (New Delhi, 116 India). Salbutamol (IS; internal standard) was gifted by Ranbaxy 117 Laboratories Ltd., Gurgaon, India. Solvents used for the 118 ultra-performance liquid chromatographic (UPLC) method were 119 purchased from Merck (Darmstadt, Germany). MS grade ammo-120 nium acetate was obtained from Fluka Analytical (Sigma-Aldrich, 121 The Netherlands). Milli-Q water was produced in-house in the lab-122 oratory by Milli-Q water purification system (MA, USA). All chem-123 icals were used as received without further purification.

124 2.2. LC/MS instrumentation and conditions

125 2.2.1. Ultra-performance liquid chromatographic condition

Ultra-performance liquid chromatography (UPLC) was per-126 127 formed with a Waters ACQUITY UPLC[™] system (S. No. F09 UPB 128 920 M; Model Code# UPB; Waters, MA, USA) equipped with a bin-129 ary solvent manager, an auto-sampler, column manager, and a tunable MS detector (Synapt; S. No. JAA272, Micromass Ltd., UK). 130 Chromatographic separation was performed on a Waters 131 ACQUITY UPLCTM BEH C18 (2.1 mm \times 100 mm; 1.7 µm) column 132 at a temperature of 40 °C. The mobile phase for UPLC analysis, con-133 sisting of acetonitrile:10 mM ammonium acetate (90:10 v/v), was 134 degassed and isocratic elution was performed at a flow rate of 135 0.2 mL min⁻¹. Ten microliter of the sample solution was injected 136 in each run with a total chromatographic run time of 4.0 min. 137 Data acquisition, data handling, and instrument control were per-138 formed by Empower[®] Software v 1.0. 139

2.2.2. Preparation of stock solution, calibration standards and Quality Control (QC) samples

The required amount of CUR and IS was dissolved in methanol 142 to prepare the standard stock solution having a concentration of 143 $50 \,\mu g \,m L^{-1}$. Different working concentrations of CUR were pre-144 pared by diluting the stock solutions with methanol. Calibration curve (CC) was prepared by spiking 2% (w/v) aqueous analyte (CUR) in a concentration range of 50 $-0.048 \ \mu g \ mL^{-1}$ in blank plasma (20 µL CUR in 980 µL blank plasma) to yield final concentration ranging from 1000 to 0.97 ng mL⁻¹. Quality control samples 149 were prepared independently in the same way as discussed above at three levels: 500 ng mL^{-1} (HQC, high quality control), 200 ng mL⁻¹ (MQC, middle quality control), and 1 ng mL⁻¹ (low 152 quality control (LQC)). All the solutions were stored at -20 °C until 153 analysis. 154

2.2.3. Plasma sample preparation

A fixed concentration (50 ng mL⁻¹) of IS (50 μ L) was mixed with 500 μ L of plasma sample and vortexed with 200 μ L of 10 mM 157 ammonium formate solution at 300 rpm for 5 min. To this mixture, 5 mL of extraction mixture (ethyl acetate:chloroform: 9:1 (v/v)) was added and kept in a reciprocating shaker for 20 min at 100 rpm. Further, it was centrifuged at 1500 rpm for 10 min at 4 °C. Supernatant (4 mL) was carefully transferred into a glass test tube and evaporated to dryness using a centrifugal evaporator (Model MV100, S. No. 026258, Tomy Medico Co., Ltd., Japan). 164 Then, the dried residue was reconstituted with 500 uL of mobile phase and a 10 µL aliquot was injected into the LC/MS system for the quantification of CUR.

2.2.4. Quadrupole-time of flight-mass spectrometry (Q-TOF-MS/MS) conditions

MS detection was performed on a Waters Q-TOF Premier 170 (Micromass MS Technologies, Manchester, UK) mass spectrometer. 171 The nebulization gas was set to $500 \text{ L} \text{ h}^{-1}$, the cone gas was set to 172 50 L h⁻¹ and the source temperature was set to 100 °C. The capil-173 lary voltage and sample cone voltage were set to 2.5 KV and 174 50 V, respectively. The Q-TOF Premier[™] was operated in V mode 175 with resolution over 32,000 mass with 1.0 min scan time, and 176 0.02 s inter-scan delay. Argon was employed as the collision gas 177 at a pressure of 5.3×10^{-5} Torr. Quantitation was performed using 178 Synapt Mass Spectrometry (Synapt MS). The transitions occurred 179 at m/z values of 367.0 \rightarrow 217.1 and 240.1 \rightarrow 148.0 for CUR and 180 salbutamol (IS), respectively. The optimum values for 181 compound-dependent parameters such as trap collision energy 182 and transfer collision energy were set to 17 and 4 V, respectively, 183 for fragmentation. The accurate mass and composition for the pre-184 cursor ions and for the fragment ions were calculated using the 185 MassLynx v 4.1 software. 186

2.3. Supercritical anti-solvent process

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The schematic presentation of SAS process (Waters SFC, USA 188 (412) 967-5655, S. No. 3636971) is shown in Fig. 1(A). It consists 189 of a heat exchanger (TharSFC, USA, model 4.2A, 1000VA, S. No. 190

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