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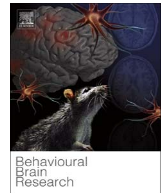

Short communication

Acute inescapable stress alleviates fear extinction recall deficits caused by serotonin transporter abolishment

Pieter Schipper[a], Marloes J.A.G. Henckens[a], Dora Lopresto[a], Tamas Kozicz[b], Judith R. Homberg[a,*]

[a] Department of Cognitive Neuroscience, Centre for Neuroscience, Donders Institute for Brain, Cognition and Behaviour, Radboudumc, Geert Grooteplein 21 (Route 126), 6525 EZ Nijmegen, The Netherlands

[b] Anatomy Department, Centre for Neuroscience, Donders Institute for Brain, Cognition and Behaviour, Radboudumc, Geert Grooteplein 21 (Route 109), 6525 EZ Nijmegen, The Netherlands

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ABSTRACT

Life stress increases risk for developing post-traumatic stress disorder (PTSD), and more prominently so in short-allele carriers of the serotonin transporter linked polymorphic region (5-HTTLPR). Serotonin transporter knockout (5-HTT$^{-/-}$) rats show compromised extinction (recall) of conditioned fear, which might mediate the increased risk for PTSD and reduce the therapeutic efficacy of exposure therapy. Here, we assessed whether acute inescapable stress (IS) differentially affects fear extinction and extinction recall in 5-HTT$^{-/-}$ rats and wildtype controls. Surprisingly, IS experience *improved* fear extinction recall in 5-HTT$^{-/-}$ rats to the level of wildtype animals, while wildtypes were unaffected by this IS. Thus, whereas 5-HTT$^{-/-}$ rats evidently were more responsive to the stressor, the behavioral consequences presented themselves as adaptive.

Severe life adversity has been linked to increased risk for developing post-traumatic stress disorder (PTSD) [1]. A large body of evidence suggests that the serotonergic system plays a role in mediating these detrimental effects of stress. Genetic variation in serotonin transporter (5-HTT) expression is known to alter stress sensitivity in humans, non-human primates and rodents, with genetic variants conferring a reduction in function (such as the 5-HTTLPR s-allele) exacerbating the effects of stressful life experiences on the incidence of PTSD [2]. Critically, traumatic life events modulate the strength and neural basis of fear acquisition and extinction in a 5-HTT dependent manner, which may underlie the increased vulnerability to psychopathology [3,4]. As fear acquisition and extinction processes are key in both the development and treatment of PTSD [5], understanding 5-HTT by stress interactions is essential for the development of therapeutic interventions attuned to these individuals.

5-HTT knockout (5-HTT$^{-/-}$) rodents are characterized by a behavioral profile of generalized anxiety (e.g. [6], and impaired fear extinction memory recall (e.g. [7])), modeling symptoms of stress-related psychopathology. While 5-HTT abolishment results in a wide array of anatomical and physiological changes and adaptations in the brain, perhaps the most prominent of these is a constitutive sevenfold increase in extracellular serotonin levels [8]. This is relevant, given that acute inescapable stress (IS), an experimental stressful life experience, impairs fear extinction by increasing dorsal raphe nucleus (DRN) serotonergic signaling and subsequent serotonin release in the basolateral amygdala (BLA) [9]. Expression of conditioned fear is associated with phasic elevation of BLA serotonin [10], and terminating serotonergic inputs into the amygdala reduces its expression, but only in repeated inescapable stress (IS) experienced mice [11], implicating a critical role for serotonin in mediating the behavioral fear phenotype induced by IS. Combining these findings with the constitutively increased extracellular serotonin levels in 5-HTT$^{-/-}$ rats raises the expectation that IS-induced fear extinction impairment is exacerbated in those with inherited 5-HTT down-regulation, explaining the 5-HTTLPR related clinical findings for PTSD.

To investigate how the effects of IS on fear extinction are modulated by 5-HTT genotype, we here assessed fear extinction and extinction recall in both naïve and IS-experienced 5-HTT$^{-/-}$ rats and their wildtype (5-HTT$^{+/+}$) counterparts [8]. We first subjected a substantial group of adult males of both genotypes to IS consisting of one session of 100 unpredictable tail shocks of randomized duration under restraint (n_5-HTT$_{-/-}$ = 20, n_5-HTT$_{+/+}$ = 19), or a control manipulation (n_5-HTT$_{-/-}$ = 20, n_5-HTT$_{+/+}$ = 16), followed by cued fear conditioning 24 h later. This stressor (albeit given after conditioning) was previously shown to increase freezing during extinction [12]. Animals were then re-exposed to the fear conditioned stimulus to measure fear extinction

* Corresponding author at: Behavioural Neurogenetics Group, Department of Cognitive Neuroscience (126), Donders Institute for Brain, Cognition and Behaviour, Radboudumc, Geert Grooteplein 21, 6525 EZ Nijmegen, The Netherlands.
E-mail address: judith.homberg@radboudumc.nl (J.R. Homberg).

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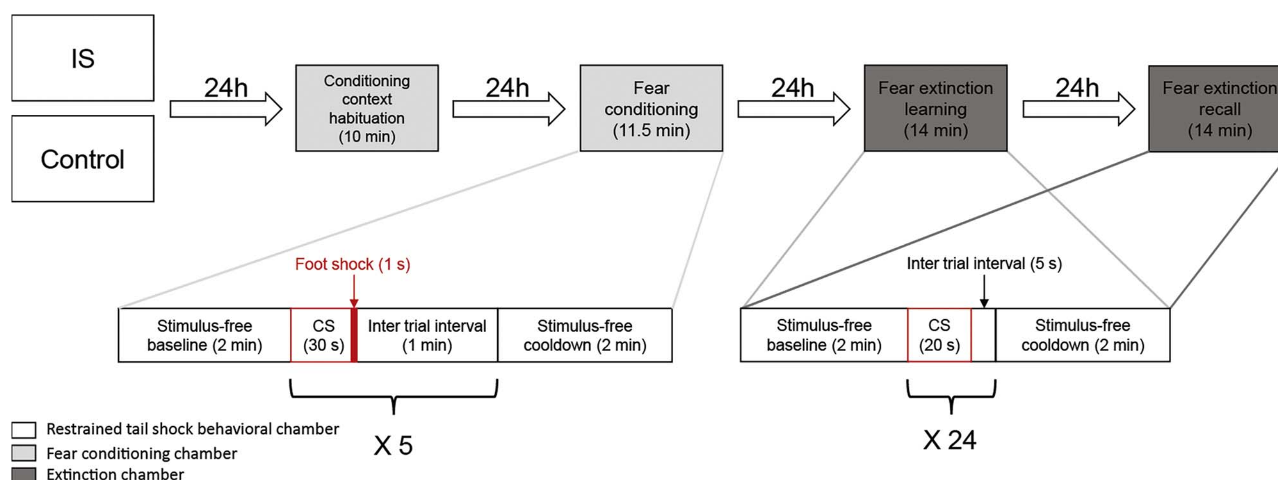

Fig. 1. Experimental outline.
All animals underwent habituation to the fear conditioning apparatus, fear conditioning, fear extinction learning and fear extinction recall testing respectively 24, 48, 72 and 96 h after IS, which consisted of 100 unpredictable tail shocks under restraint, or a control manipulation consisting of two hours of mild restraint in the behavioral apparatus used for tail shock administration.

learning and subsequent recall, by means of behavioral freezing (see Fig. Figure 1 for the experimental timeline).

Serotonin transporter knockout rats (Slc6a41Hubr) were generated on a Wistar background by N-ethyl-N-nitrosurea (ENU)-induced mutagenesis. Experimental animals were derived from crossing heterozygous 5-HT transporter knockout (5-HTT$^{+/-}$) rats that were outcrossed for at least twelve generations with wild-type Wistar rats obtained from Harlan Laboratories (Horst, The Netherlands). Ear punches were taken at the age of 21 days after weaning for genotyping, which was done by Kbiosciences (Hoddesdon, United Kingdom). We tested male adult 5-HTT$^{-/-}$ and 5-HTT$^{+/+}$ rats which ranged from 16 to 24 weeks of age. The animals were housed in pairs, in open cages. All animals had *ad libitum* access to food and water. A 12-h light-dark cycle was maintained, with lights on at 08.00 A.M. All behavioral experiments were performed between 08.00 A.M. and 18:00 P.M. All experiments were approved by the Committee for Animal Experiments of the Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands, and all efforts were made to minimize animal suffering and to reduce the number of animals used.

IS tail shocks were given in a triadic chamber (large) measuring 18.3 × 11.4 × 18.5 cm with grid floors (Med Associates, St. Albans, VT, USA). The grid floors were covered with vinyl to minimize injury to the animal. Shocks were delivered by a shock generator (model ENV-412, Med Associates). A 30.5 cm × 24.1 cm × 21 cm operant conditioning chamber (Model VFC-008, Med Associates) was used for fear conditioning and sham conditioning. The box was housed within a sound-attenuating cubicle and contained a white LED stimulus light, a white and near infrared house light as well as a speaker capable of producing an 85 dB 2.8 kHz tone. The metal grid floor of the apparatus was connected to a scrambled shock generator (model ENV-412, Med Associates) configured to deliver shocks at 0.6 mA intensity. Fear extinction and extinction recall were tested in a novel context. The novel context consisted of a 25 cm × 25 cm × 30 cm Plexiglas cage, the bottom of which was covered in a +/−0.5 cm thick layer of black bedding. In this context, 85 dB (measured at the center of the floor) 2.8 kHz auditory stimuli were delivered through a set of external speakers.

Animals in the IS group were restrained by the tail in the triadic chamber using disposable finger electrodes, under which electrolytic gel was applied. 100 shocks of increasing intensity (30 shocks at 0.8 mA, 30 shocks at 1.0 mA, 40 shocks at 1.2 mA) and of randomized duration (1 – 30 s, 5 s average) were given on a variable interval schedule ranging from 50–70 seconds (60 s average). The IS procedure took 2 h. Control animals were restrained by the tail (while they were still able to move all limbs) for 2 h in the apparatus using disposable finger electrodes, but were not given shocks. 24 h after IS or the control procedure, animals were habituated to the fear conditioning environment for 10 min. The house light was on during habituation and conditioning. For the fear conditioning itself, after a 2 min habituation period, a 30 s 85 dB 2.8 kHz auditory stimulus co-terminated with a 1 s 0.6 mA foot shock, followed by a 1 min inter-trial interval. A total of 5 of these tone – shock pairings were given. 24 h and 48 h after conditioning, fear extinction and extinction recall were tested, respectively. After a 2 min habituation period, 24 20-s presentations of the auditory stimulus were given, with an inter-trial interval of 5 s. Conditioning and extinction sessions were recorded and freezing was manually assessed by a trained observer who was blind to genotype and treatment. For the IS or control procedures, the conditioning and the habituation to the fear conditioning chamber, the apparatus was cleaned before and after each animal using a tissue slightly dampened with 70% EtOH. Water was used for cleaning during the extinction and extinction recall. Due to equipment malfunction, the conditioning session could be recorded only for half the animals of each group.

All statistical analyses were performed using IBM SPSS Statistics version 20.0 (SPSS Inc., Chicago, Illinois, USA). Data are presented as mean ± standard error of the mean (SEM). Effects of genotype and treatment on freezing during conditioning and extinction were analyzed using a 2-way repeated measures ANOVA (F); development of freezing behavior was assessed across extinction sessions and trial blocks within the extinction recall session. Significant genotype x treatment interactions were further explored using *post hoc* Student's *t*-tests. Probability p-values below 0.05 were considered significant.

When assessing freezing during the stimulus free 2-minute period preceding the tone-shock pairings in the conditioning session through 2-way ANOVA, we found an effect of genotype ($F_{(1,36)} = 4.591$, $p = 0.039$), with 5-HTT$^{-/-}$ spending more time on freezing. No effect of IS ($F_{(1,36)} = 0.155$, $p = 0.696$), nor a genotype x IS interaction effect ($F_{(1,36)} = 0.123$, $p = 0.728$) was found. Analyzing total time spent freezing during cue presentation in the fear conditioning session using repeated measures 2-way ANOVA analysis yielded no effect of genotype ($F_{(1,36)} = 0.021$, $p = 0.884$), IS ($F_{(1, 36)} = 0.707$, $p = 0.406$), or genotype x IS interaction ($F_{(1, 36)} = 0.1358$, $p = 0.716$) (Fig. Figure 2A).

Analysis of time spent freezing during the stimulus free baseline-period preceding the extinction sessions revealed no significant effect of genotype, IS or genotype x IS interaction in the extinction learning session ($F_{(1,74)} = 2.153$, $p = 0.147$; $F_{(1,74)} = 3.592$, $p = 0.062$; and

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