



Original article

Chitosan treatment for skin ulcers associated with diabetes

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ARTICLE INFO

Article history:

Received 2 March 2016

Revised 10 March 2017

Accepted 29 March 2017

Available online xxxxx

Keywords:

Chitosan

Ulcer

Diabetes mellitus

Gel

Film

Biomaterials

ABSTRACT

Infections, ulcerations, gangrene and, in severe cases, extremity amputation, are common complications among diabetic subjects. Various biomaterials have been utilized for the treatment of these lesions. Chitosan is an amino sugar with a low risk of toxicity and immune response. In this study, we evaluated chitosan topical gel and film treatments for subjects with diabetic ulcerations and wounds associated with diabetes mellitus. In a pre-experimental design, we described the result of chitosan gel and film treatment for wounds and skin ulcers among patients with long-standing diabetes mellitus. We studied 8 diabetic patients with wounds and skin ulcers (long duration and Wagner degree 1–2). Initially, most lesions had some degree of infection, tissue damage and ulceration. At the end of the treatment (topical chitosan) period, the infections were cured. All patients experienced a significant improvement in the initial injury and developed granulation tissue and a healthy skin cover. This report represents one of the few published clinical experience regarding the chitosan for the treatment of skin lesions among diabetic subjects. These results are relevant and promising for the treatment of this disease.

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1. Introduction

Diabetic foot is a common complication among patients with diabetes mellitus (DM) (Boada, 2012). Diabetic foot is one of the heaviest and most debilitating financial burdens, not only for the sufferer, but for the family and health system (Brito-Zurita et al., 2013; Odhayani et al., 2015). Some of the complications associated with diabetic foot include infections, ulcers, gangrene and extremity amputation (in severe cases) (Rubio et al., 2012). For mild infections, *Staphylococcus aureus* is present in 80–90% of lesions, whereas *Enterococcus spp.* and *Pseudomonas spp.* are responsible for moderate-to-severe infections (Dai et al., 2010). The treatment options for diabetic foot include substances with local effects, such as hyper-oxygenated fatty acids, silicone, alginates, different antiseptics, carbon dressings, silver, and collagen.

Peer review under responsibility of King Saud University.



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The use of synthetic acellular biomaterials for replacing skin has recently been introduced (Dieckmann et al., 2010). Among these biomaterials, chitosan, which is a polysaccharide composed of amino sugars, has a low risk of toxicity and immune response (Dieckmann et al., 2010). Chitosan is a biodegradable linear polymer that is non-toxic to mammals and is obtained by partial deacetylation of chitin (Sánchez-Duarte et al., 2012). Chitosan is a naturally occurring compound and is present in the coating of some mollusks and insects. Chitosan is comprised of glucosamine and N-acetyl-D-glucosamine linked by β -(1 → 4) glycosidic bonds (Aytekin et al., 2011) and is insoluble in water and soluble in acidic solutions (pH < 6.5). The properties of chitosan depend on molecular weight, deacetylation degree and ionic strength of the molecules (Dutta et al., 2009). Due to its cationic nature, chitosan has antimicrobial activity against a wide range of microorganisms, such as *Escherichia coli*, *Salmonella typhimurium*, *Staphylococcus aureus* and *Listeria monocytogenes* (Rodríguez-Núñez et al., 2014). Among the properties observed in the tissues where it has been used, chitosan allows blood to clot quickly and has recently gained approval in the United States for use in bandages and other hemostatic agents (Rabea et al., 2003).

Several animal studies have been performed using chitosan, and this product has proven superior to others in eliminating infection

<http://dx.doi.org/10.1016/j.sjbs.2017.03.017>

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Please cite this article in press as: Escárcega-Galaz, A.A., et al. Chitosan treatment for skin ulcers associated with diabetes. Saudi Journal of Biological Sciences (2017), <http://dx.doi.org/10.1016/j.sjbs.2017.03.017>

and promoting tissue healing compared to frequently used compounds by physicians (Dai et al., 2010). However, few clinical studies have been performed using chitosan for the treatment of skin ulcers. Valentine et al. (2010) tested 40 subjects who underwent chitosan treatment for hemostasis and wound repair subsequent to endoscopic surgery. The authors observed decreased bleeding times from the wounds. Later, Velazco et al. (2012) reported the application of dressings made with chitosan on patients with diabetic foot complications. The granulation process begins at eight days after dressing placement and at 22 days there was decrease in the size of the lesion. At 45 days, the total closure of the ulcer was observed. Furthermore, the dressings were completely biodegradable. Due to the current evidence, we evaluated chitosan topical gel and film treatments for diabetic patients with wounds and ulcers associated with diabetes mellitus.

2. Materials and methods

Utilizing a pre-experimental study design, we describe the results of administering chitosan gel and film to wounds and ulcers to patients with long-standing diabetes mellitus.

2.1. Obtaining chitosan

Chitosan was obtained from the deacetylation of chitin using the methodology proposed by Correa-Murrieta et al. (2012) according to the following procedure. First, the raw chitin was isolated by lactic acid fermentation of the shrimp head. For its purification, the chitin was desmineralized with chlorhydric acid 1 M (1:15 w/v) at 27 °C for 4 h. Then, the solid was desproteinized with NaOH 4.5% at 65 °C for 4 h. Finally, the chitin was deacetylated with NaOH 45% at 110 °C for 1:30 h. All steps were with constant agitation. The moisture content was adjusted by drying the sample in an oven (Felisa, Zapopan, Jalisco, Mexico) at 60 °C for 5 h until achieving a constant weight. The ash content was determined on a Barnstead Thermolyne muffle (Madison, Iowa, USA) at 550 °C for 5 h until obtaining a gray powder. The protein content was determined by a spectrophotometric method based on adding the colorant Comassie Blue G-250 to the proteins. The deacetylation degree (DD) was determined by UV spectrophotometry according to the method proposed by Liu et al. (2006) using glucosamine and N-acetyl glucosamine standards. The viscosity was determined with an Ostwald viscometer (Vineland, Nueva Jersey, EE. UU.) in a water bath at 21 ± 1.0 °C. The molecular weight determination was performed according to the method proposed by Solis et al. (2007). For this determination, chitosan solution with a 0.0014 g/mL concentration was calculated by the method of Mark-Houwink (Wagner, 1985).

2.2. Chitosan gel and film preparation

Chitosan gel was prepared at 2%. For this, two grams of chitosan were added to 100 mL of acetic acid 1% and were stirred for 12 h.

Chitosan films were prepared by casting method according to the methodology proposed by Rodríguez-Núñez et al. (2014) with minor modifications. Those were made to base from chitosan gel 2%, specifically, 4 ml this solution were poured onto polypropylene plates and dried at 60 °C for 12 h. Finally, the films was removed from the mold and stored in sterile plastic bags (Whirl-Pak®).

2.3. Samples and subjects

A group of subjects with wounds and ulcerations associated with diabetes mellitus attended the sports medicine clinic at the Instituto Tecnológico de Sonora in Ciudad Obregón, Sonora (NW

Mexico) between January and December 2014. Subjects had wound or skin ulceration with a 2nd and 3rd degree risk according to the classification proposed by Heus van Putten et al. (1996). These patients were referred by friends and supporters of the university, and they freely requested to receive treatment. All patients were questioned about their epidemiological history related to their illness. Subjects with a risk greater than the 3rd degree (according to the Wagner classification) were excluded in order to avoid exposing them to any unnecessary risk. In addition, consent was obtained from all participants.

2.4. Healing protocol

The injured tissue was washed with a physiological saline solution at 0.9% (Pisa, Guadalajara, Jalisco, Mexico) and dried with a sterile gauze. Subsequently, some chitosan gel 2% was spread on the wound using a sterile syringe, allowed to rest for 1 min, and a chitosan film was then immediately placed on the wound. Finally, a band was applied to hold the chitosan film in place. The procedure was repeated every 48 h for the duration of the study. The follow-up period lasted 3 months for each subject in this study.

2.5. Clinical response analysis

Visual evidence regarding the wound was obtained at each visit when removing the healing material, any variations in the shape and size of the ulcer were recorded. The evaluation was qualitative, and improvement was regarded as a decrease in ulcer size, presence of granulated tissue and absent or decreased wound infection. Moreover, healing was regarded as wound scarring with an apparent absence of infection or necrotic tissue.

3. Results and discussion

3.1. Characterization of chitosan

The physicochemical composition of chitosan is related to its quality and functional properties. The ash, protein and moisture content were 0.15 ± 0.03%, 0.25 ± 0.01% and 13.69 ± 0.03%, respectively. Supplementary characteristics of chitosan included the deacetylation degree (97.02 ± 0.30%), molecular weight (153.15 kDa) and viscosity (range 1119.62–3655.09 cP).

The purity level of chitosan affects its biological properties, solubility and stability (Szymanska and Winnicka, 2015). For biomedical applications the chitosan should be of high purity. Knowledge of the degree of deacetylation allows to predict the biological functionality of the chitosan. As the degree of deacetylation increases, the rate of inflammatory response in the ulcer decreases. Also, this parameter usually has an impact on the immunogenic behavior of the polymer (Kurita et al., 2000). The molecular weight depends mainly on the extraction process and can be determine its antimicrobial properties. It has been reported that lower molecular weight chitosan is able to penetrate into the cell and destroy the microorganism (Hosseinnejad and Jafari, 2016). The chitosan of this research has low molecular weight chitosan, which may induce the destruction of the microorganisms that generate infection in ulcers. Pillai et al. (2009) reported viscosity values in a range of 200–2000 cps and this variation is related to the molecular weight and degree of deacetylation of chitosan.

3.2. Characterization chitosan films

The films were a light yellow color, smooth at the surfaces, uniform and with an average transparency value of 1.40 ± 0.10. The

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