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Perspective

Influence of sodium dodecyl sulfate coating on adsorption of methylene blue by biochar from aqueous solution

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ABSTRACT

Biochar is regarded as a promising new class of materials due to its multifunctional character and the possibility of effectively coupling different properties. With increasing introduction into the environment, environmental chemicals such as surfactants will load onto the released biochar and change its physicochemical characteristics and adsorption behavior toward pollutants. In this study, sodium dodecyl sulfate (SDS), as one type of anionic surfactant, was coated onto biochar with different loading amounts. The influence of SDS loading onto biochar's physicochemical properties were investigated by Fourier transform infrared (FT-IR) spectroscopy, elemental analysis, zeta potential and Brunauer–Emmett–Teller (BET) surface area and pore size distribution analysis. Results showed that the pore size of the biochar decreased gradually with the increase of SDS loading because of the surface-adsorption and pore-blocking processes; the pH of the point of zero charge (pH_{PZC}) decreased with increasing SDS loading. Although surface-coating with SDS decreased the pore size of the biochar, its adsorption capacity toward methylene blue (MB) significantly increased. The biochar-bound SDS introduced functional groups and negative charges to the biochar surface, which could thus enhance the adsorption of MB via hydrogen bonding and electrostatic interaction. The results can shed light on the underlying mechanism of the influence of anionic surfactants on the adsorption of MB by biochar.

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Introduction

Biochar is a carbon-rich aromatic material produced by pyrolyzing biomass through various thermochemical processes

(i.e., pyrolysis, hydrothermal carbonization, flash carbonization, and gasification) under oxygen-limited conditions and at relatively low temperatures ($<700^\circ\text{C}$) (Tan et al., 2017). The specific properties of biochar include high specific surface area, porous structure, and copious amounts of

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59 surface functional groups (Safaei Khorram et al., 2016; Wang
60 et al., 2017). Although its specific surface area and micropore
61 volume are much lower than that of activated carbon, biochar
62 is still regarded as a good replacement for commercial
63 activated carbon because of its abundant functional groups
64 and lower cost (Zhou et al., 2016). Recently, biochar has been
65 used widely as an effective adsorbent for removal of heavy
66 metals and organic compounds from wastewater (Chen et al.,
67 2008; Guo et al., 2014; Jiang et al., 2017a, 2017c; Pan et al., 2013;
68 Shen et al., 2017; Zafar et al., 2017). Among these pollutants,
69 much attention has been paid to the adsorption of dyes by
70 biochar due to their toxic effects and high concentrations in
71 wastewaters. For instance, Nautiyal et al. (2016) used *Spirulina*
72 *platensis* algae to prepare biochar which exhibited excellent
73 adsorption performance toward Congo red. Ding et al. (2016)
74 investigated the simultaneous adsorption of methyl red and
75 methylene blue onto woody biochar in different concentra-
76 tion combinations. Sewu et al. (2017) examined the adsorp-
77 tion ability of crystal violet onto biochar produced from
78 Korean cabbage, and the Langmuir maximum adsorption
79 capacity was as high as 1304 mg/g.

80 Surfactants, as a key component of industrial and household
81 waste, are routinely deposited in numerous ways on land
82 and into water systems (Emmanuel et al., 2005; Metcalfe et al.,
83 2008). These commonly are organic substances with amphiphilic
84 structures, indicating that they contain both hydrophobic groups
85 and hydrophilic groups, which allow them to adsorb various
86 pollutants such as heavy metal ions and organic molecules
87 (Flores et al., 2017; Gamba et al., 2017; Shah et al., 2017; Stofela
88 et al., 2017; Zhang et al., 2017). As reported, surfactants can
89 interact with geosorbents and impact their adsorption behavior
90 toward pollutants (Koh and Dixon, 2001; Park et al., 2011;
91 Richards and Bouazza, 2007; Xi et al., 2010). Surfactants also
92 can bind to released nanomaterials, change their physicochem-
93 ical characteristics, and then affect their adsorption mechanism
94 (Liu et al., 2014; Rajesh and Ramagopal, 2016). It was reported
95 that a surfactant (i.e., cetylpyridinium chloride) can coat onto
96 activated carbon and mineral substances to improve their
97 removal performance for heavy metals from water (Choi et al.,
98 2009; Lin et al., 2011; Sánchez-Martín et al., 2008). Furthermore,
99 several papers have demonstrated that dissolved surfactants
100 could impact the adsorption of organic pollutants by biochar.
101 For example, Han et al. (2013) reported the influence of
102 a cationic surfactant on pentachlorophenol adsorption by
103 biochar; and the cationic surfactant cetyltrimethyl ammoni-
104 um bromide could be coated onto the biochar surface *via*
105 ion-exchange with abundant exchangeable cations, such as
106 Mg^{2+} , Na^+ , and K^+ , and then impacted the pentachlorophenol
107 adsorption.

108 Considering its increasing production and application,
109 biochar has inevitably been released into the environment.
110 Understanding the adsorption behavior of biochar toward
111 toxic dye in the presence of surfactant has multiple important
112 environmental implications, including assessing (i) the feasi-
113 bility of using biochar as an adsorbent in engineered treatment
114 of toxic dye, (ii) the fate and transport of toxic dye by biochar in
115 the environment, and (iii) the potential environmental and
116 health risks of biochar-adsorbed toxic dye in ecological systems.
117 In this work, therefore, the physicochemical characteristics of
118 biochar with surfactant coating such as sodium dodecyl sulfate

(SDS) were investigated. In addition, the effect of the presence of
SDS on the adsorption behaviors of methylene blue (MB) onto
biochar was also examined.

1. Material and methods

1.1. Materials

The biomass used in this work was peanut shell obtained
locally from the farms in Changsha City, Hunan Province,
China. MB (3,7-bis{dimethylamino}-phenazathionium chlo-
ride, $C_{16}H_{18}ClN_3S \cdot 3H_2O$, molecular weight 373.9) was ob-
tained from Tianjin Hengxing Chemical Reagent Co., Ltd.
(Tianjin, China). SDS was purchased from Shanghai Yabang
Chemical Reagent Co., Ltd. (Shanghai, China). All other
chemicals including HCl, NaOH and NaCl were provided by
Guangdong Xilong Chemical Co., Ltd. All chemicals employed
in this experiment were analytical reagent grade and used
without any further purification. The water used in this work
was Milli-Q water (18.25 M Ω /cm) obtained from a Millipore
Milli-Q water purification system.

1.2. Preparation of adsorbent

The biochar was prepared as described by a previously published
paper (Jiang et al., 2015). Briefly, 10-g biomass was added to
a 250-mL beaker with 100-mL $Zn(NO_3)_2$ solution (1 mol/L).
Then, the mixture was stirred continuously in a temperature-
controlled water bath shaker at 40°C for 24 hr. After being
dried, the mixture was pyrolyzed by a lab-scale tubular reactor
(SK-G08123K, China) in a N_2 environment (400 mL/min) at 550°C
for 1 hr. After cooling, the resulting biochar was washed with
water, and dried in an oven at 65°C. After that, 1 g of prepared
biochar was added into 100 mL of SDS solution (3, 1 and 0.6 mg,
respectively). The mixture was also shaken in a temperature
controlled water bath shaker at 120 r/min for 24 hr. Then
the SDS-coated biochar was collected by filtration and dried
at 65°C for 12 hr. The products were named 0.6 SDS-biochar,
1 SDS-biochar, and 3 SDS-biochar, respectively. Finally, the
prepared samples were stored in a desiccator for further use
(Mi et al., 2016).

1.3. Adsorbent characterization

Fourier transform infrared (FT-IR) spectroscopy measurements
were performed using a Fourier Transform Infrared Spectrometer
(Nicolet 5700 Spectrometer) using the KBr pellet technique in
the range of 4000–400/cm. The Brunauer–Emmett–Teller (BET)
surface area and the pore size distribution were determined
by using N_2 adsorption–desorption (Quantachrome, USA) at 77 K
over a relative pressure range from 0.0955 to 0.993. C, H, O,
N and S contents of samples were determined using an ele-
mental analyzer (Elementar, Vario EL III). Thermal analysis of
the precursor was done by thermogravimetry-differential scan-
ning calorimetry (TG–DSC) on a Netzsch STA 449C instrument
(conditions: air atmosphere, temperature ramp from 25 to 900°C,
rate 20°C/min). The zeta potential of samples was obtained
using a zeta potential meter (Zetasizer Nano-ZS90, Malvern) by
adjusting the solution pH from 2.0 to 11.0.

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