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Pharmacokinetics and Preventive Effects of Sulfo-Albumin as a Novel Macromolecular Hydrogen Sulfide Prodrug on Carbon Tetrachloride-Induced Hepatic Injury

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ABSTRACT

Hydrogen sulfide (H₂S) has been recently recognized as a gaseous signaling molecule that controls various biological activities. In the present study, we developed sulfo-albumin as a macromolecular H₂S prodrug for therapeutic use, in which polysulfide groups (source of H₂S) were conjugated with bovine serum albumin through a covalent linkage. In an *in vitro* study on H₂S release in phosphate buffered saline solution, we found that H₂S was released from sulfo-albumin in the presence of 5-mM glutathione but not in its absence. Furthermore, sulfo-albumin was taken up by RAW 264.7 cells, and it released H₂S in cells but not in plasma. These results indicate that H₂S can be selectively released from sulfo-albumin in cells. ¹¹¹In-labeled sulfo-albumin predominantly accumulated in the liver, dependent upon the number of sulfide groups, after intravenous injection in mice. In a carbon tetrachloride-induced acute liver injury mouse model, sulfo-albumin significantly suppressed the increase in plasma aspartate aminotransferase and alanine aminotransferase activities, which are indicators of hepatocyte injury, after intravenous injection. These findings indicate that sulfo-albumin is a promising compound for the treatment of hepatic injuries.

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Introduction

Although hydrogen sulfide (H₂S) has been considered as a toxic pollutant, it has recently been found that H₂S is endogenously generated from various synthases such as cystathionine β-synthase, cystathionine γ-lyase, and 3-mercaptopyruvate sulfur-transferase in the body.^{1,2} H₂S plays an important role in various biological activities, including anti-oxidative stress, anti-inflammation, anti-apoptosis, and vascular smooth muscle relaxation.³⁻⁷ Therefore, H₂S is recognized as a third gaseous signaling molecule in biological systems, following nitric oxide (NO) and carbon monoxide (CO).^{7,8} As H₂S has a protective effect against reactive oxygen species (ROS)-mediated diseases including inflammatory and ischemia/reperfusion

injuries,^{3,4} the delivery of H₂S to the site where ROS are generated has been expected for the treatment of ROS-mediated diseases.

To date, some researchers have attempted the use of H₂S to treat ROS-mediated injury.^{9,10} Because it is difficult to handle and administer H₂S directly, H₂S prodrugs that generate H₂S *in vivo* have been developed for efficient administration of H₂S into the body.¹¹⁻¹³ However, the tissue distribution of H₂S prodrugs has not been studied, despite the fact that the tissue distribution of H₂S prodrugs requires optimization to obtain the maximal therapeutic effect of H₂S. Of the various strategies available, the use of macromolecular carriers appears to be a good approach to control the release and distribution of H₂S. This is because the macromolecular carriers have several functional groups whereby sulfide groups (source of H₂S), targeting ligands, and functional moieties can be chemically conjugated. To control the delivery of H₂S, the tissue distribution of the sulfide group-modified macromolecular carrier should be examined.

In the present study, to provide basic information for controlled delivery of H₂S *in vivo*, we developed a novel macromolecular H₂S prodrug and examined its physicochemical properties, pharmacokinetics, and therapeutic potential in the ROS-mediated injury. We

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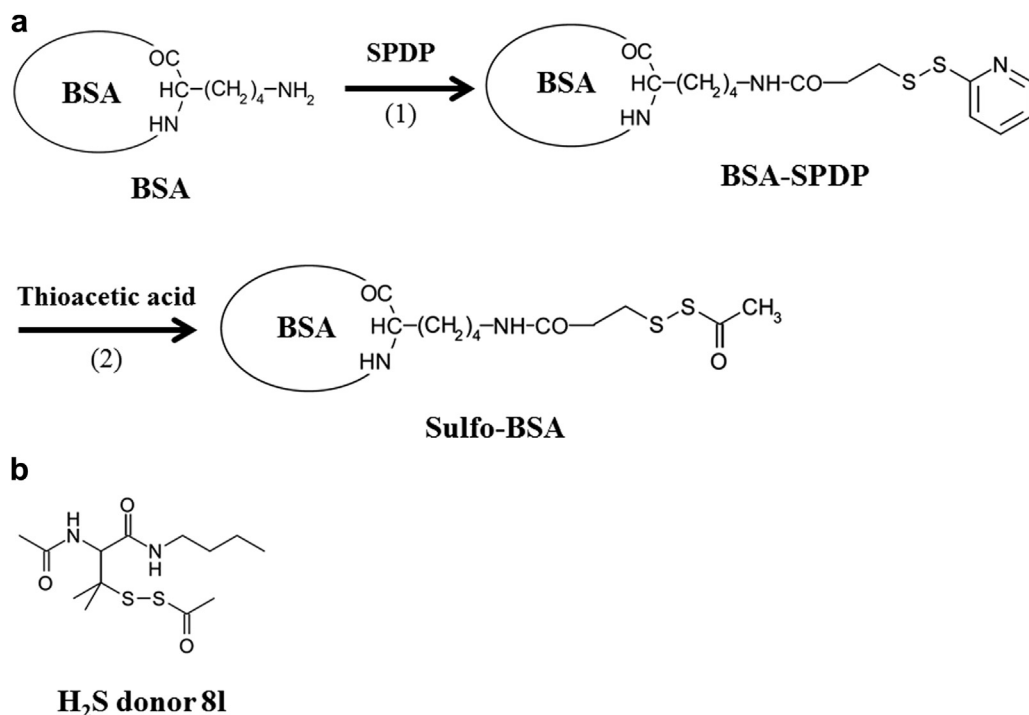


Figure 1. Structures and synthetic routes of sulfo-albumin (Sulfo-BSA) (a) and structure of H₂S prodrug 81 (b).

developed sulfo-albumin, in which multisulfide groups were conjugated to serum albumin through a covalent linkage, as a macromolecular H₂S prodrug (Fig. 1a). Next, the H₂S release and tissue distribution of sulfo-albumin were examined. Finally, the therapeutic effect of sulfo-albumin was investigated in carbon tetrachloride (CCl₄)-induced acute hepatic injury model mice.

Materials and Methods

Chemicals

Bovine serum albumin (BSA) and fluorescein isothiocyanate (FITC) isomer were purchased from Sigma-Aldrich (St. Louis, MO). N-Succinimidyl 3-(2-pyridyldithio) propionate (SPDP), diethylenetriaminepentaacetic acid anhydride, HSip-1, and N-Butyl-N²-acetyl-S-acetylsulfanyl-DL-penicillamine amide (low-molecular-weight thiol-activated H₂S prodrug 81; Fig. 1b) were purchased from Dojindo Laboratory (Kumamoto, Japan). Fetal bovine serum was obtained from Biosera (Ringmer, UK). Thioacetic acid, dimethyl sulfoxide, zinc acetate, N,N-dimethyl-*p*-phenylenediamine, and CCl₄ were purchased from Wako Pure Chemical Industries (Osaka, Japan). ¹¹¹Indium chloride was kindly supplied by Nihon Medi-Physics (Takarazuka, Japan). All other chemicals were of reagent grade.

Table 1
Physicochemical Properties of Sulfo-albumin

Compound	Molecular Weight ^a	Number of Sulfide Groups ^b (mol/mol)	Diameter (nm)	Zeta Potential (mV)
BSA	66,300	0	7.29 ± 0.42	-12.57 ± 0.12
Sulfo (5)-BSA	67,300	4.83	9.20 ± 0.06	-19.47 ± 0.78
Sulfo (10)-BSA	68,100	9.72	8.24 ± 0.68	-22.23 ± 1.37
Sulfo (30)-BSA	71,900	30.9	7.21 ± 0.70	-30.87 ± 0.75

^a Molecular weight was measured via MALDI TOF-MS.

^b The average number of sulfide groups was estimated by measuring the molecular weight via MALDI TOF-MS. Results are expressed as the mean ± SD of 3 experiments.

Animals

Male ddY mice (25 g) were purchased from Japan SLC Inc. (Shizuoka, Japan). Animals were maintained under conventional housing conditions, and all animal experiments were conducted in accordance with principles and procedures outlined in the NIH Guide for the Care and Use of Laboratory Animals. The protocols for animal experiments were approved by the Animal Experimentation Committee of the Kyoto Pharmaceutical University.

Synthesis of Sulfo-Albumin

Sulfo-albumins with one of 3 different degrees of sulfide group modifications (Sulfo (5)-BSA, Sulfo (10)-BSA, and Sulfo (30)-BSA) were prepared by reacting different amounts of SPDP and thioacetic acid with BSA. Typically, to synthesize Sulfo (30)-BSA, 30 mg of BSA was dissolved in 5 mL of 0.1 M phosphate buffer (pH 7.4), and 8.48 mg of SPDP was dissolved in 50 μL of dimethyl sulfoxide and added to the BSA solution. The reaction mixture was stirred at room temperature for 2 h. Then, 9.57 μL of thioacetic acid was added to the reaction mixture, and stirring continued for 1.5 h at room temperature.¹¹ Next, the mixtures were dialyzed using dialysis membrane (prewetted RC tubing [molecular weight cutoff: 15kD]; Spectrum Laboratories, Inc., Dominguez, CA) against distilled water

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