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Bioluminescent and micro-computed tomography imaging of bone repair induced by fibrin-binding growth factors

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In this work we have evaluated the capacity of BMP-2 and fibrin-binding PDGF-BB to support cell growth and induce bone regeneration using two different imaging technologies to improve understanding of structural and organizational processes participating in tissue repair. Human mesenchymal stem cells from adipose tissue (hAMSCs) expressing two luciferase genes, one under the control of the CMV promoter and the other under the control of a tissue-specific promoter (osteocalcin or PECAM), were seeded in fibrin matrices containing BMP-2 and fibrin-binding PDGF-BB, and further implanted intramuscularly or in a mouse calvarial defect. Then, cell growth and bone regeneration were monitored by bioluminescence imaging (BLI) to analyze the evolution of target gene expression, indicative of cell differentiation towards the osteoblastic and endothelial lineages. Non-invasive imaging was supplemented with micro-computed tomography (microCT) to evaluate bone regeneration and high-resolution microCT of vascular casts. Results from BLI showed hAMSC growth during the first week in all cases, followed by a rapid decrease in cell number; as well as an increment of osteocalcin but not PECAM-1 expression 3 weeks after implantation. Results from microCT show that the delivery of BMP-2 and PDGF-BB by fibrin induced the formation of more bone and improves vascularization, resulting in more abundant and thicker vessels, in comparison with controls. Although inclusion of hAMSCs in the fibrin matrices made no significant difference in any of

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