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Short communication

Variability of *in vivo* reference point indentation in skeletally mature inbred rats

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ABSTRACT

Reference point indentation (RPI) has emerged as a novel tool to measure material-level biomechanical properties *in vivo*. Human studies have been able to differentiate fracture *versus* non-fracture patients while a dog study has shown the technique can differentiate drug treatment effects. The goal of this study was to extend this technology to the *in vivo* measurement of rats, one of the most common animal models used to study bone, with assessment of intra- and inter-animal variability. Seventy-two skeletally mature male Sprague-Dawley rats were subjected to *in vivo* RPI on the region between the tibial diaphysis and proximal metaphysis. RPI data were assessed using a custom MATLAB program to determine several outcome parameters, including first cycle indentation distance (ID-1st), indentation distance increase (IDI), total indentation distance (TID), first cycle unloading slope (US-1st), and first cycle energy dissipation (ED-1st). Intra-animal variability ranged from 13% to 21% with US-1st and Tot Ed 1st-L being the least variable properties and IDI the most highly variable. Inter-animal variability ranged from 16% (US-1st) to 25% (ED-1st and IDI). Based on these data, group size estimates would need to range from 9 to 18/group to achieve sufficient power for detecting a 25% difference in a two-group experiment. Repeat tests on the contralateral limb of a small cohort of animals ($n=17$) showed non-significant differences over 28 days ranging from −6% to −18%. These results provide important data on RPI variability (intra- and inter-animal) in rats that can be used to properly power future experiments using this technique.

1. Introduction

Assessment of biomechanical properties has long been confined to pre-clinical studies and, more specifically, *ex-vivo* mechanical tests. Recent technology, termed reference point indentation (RPI), has made it possible to assess biomechanical properties *in vivo* (Hansma et al., 2008). *in vivo* studies have shown that RPI can differentiate between patients who have fractured *versus* non-fracture patients (Diez-Perez et al., 2010) as well as patients who have been treated with bisphosphonates *versus* those who were treatment naïve (Güerri-Fernández et al., 2012). *in vivo* testing of dogs has shown RPI can differentiate raloxifene treatment from controls after six months of clinically relevant dosing (Aref et al., 2013). In addition, a related device (Osteoprobe) that operates using slightly different technology revealed significant differences in the material properties of patients with diabetes *versus* healthy controls (Farr et al., 2014). Collectively, these data show promise for RPI technology to allow minimally invasive measures of material-level biomechanical properties.

Rodents represent the most commonly used animal model to study bone and are often the model first used to evaluate novel interventions (Kalu, 1991; Thompson et al., 1995). Although several studies have assessed biomechanical properties of rodent bone *ex vivo*, there have been no reports of *in vivo* assessment of rodents. The goal of this study was to determine the intra- and inter-animal variability, as well as the variability over time (in order to understand potential variability that might occur in control animals in future intervention studies), for *in vivo* measures with RPI in skeletally mature rats. These data will be essential to understand the practicality of the technique for use in rats as well as to provide variability data to help design adequately powered experiments.

2. Methods

2.1. Experimental design

Seventy-two skeletally mature male (6 month old) Sprague Dawley rats were purchased (Harlan) and acclimatized for one week prior to reference point indentation (RPI) testing. A subset of animals ($n=17$) underwent a second RPI test session 28 days after the first test. These repeat test sessions were performed on

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the contralateral limb to avoid any local tissue damage caused by the first test session. Following each testing session, animals were returned to their cages. These animals were part of a larger experiment that is outside the scope of this current report. All procedures were approved by the Indiana University School of Medicine Animal Care and Use Committee prior to the start of the study.

2.2. Reference point indentation (RPI)

Material-level mechanical properties of the anterior surface of the tibial cortex were assessed *in vivo* using RPI (Biodent Hfc, Active Life Scientific, Santa Barbara, CA). This site was chosen as it has been utilized previously in human and dog *in vivo* studies, and its limited soft tissue coverage facilitates easy access to the bone surface. The cortical thickness in this region is around 4 mm thick. Rats were placed under general anesthesia using inhalation isoflurane, and a local anesthetic was injected just beneath the skin in the region of testing. Skin overlying the region was pierced with a sterile BP1 probe contained within the measurement head unit (MHU) attached to a modified holder apparatus (Fig. 1). The MHU was lowered vertically, normal to the surface of the bone, until the probe assembly rested on the bone surface. As opposed to previous *in vivo* work in humans and dogs, we did not scrape the periosteum prior to testing due to challenges working in the small target area. Following positioning of the reference probe, a reference force of ~13 Newtons was applied to stabilize the MHU, and the measurement protocol was initiated. Measurements began with a series of four preconditioning cycles (1N force at 5 Hz) followed by a series of 10 testing cycles (10 N at 2 Hz). This force was chosen to match *in vivo* levels used previously in humans and dogs. To achieve our goal of three usable tests for each animal, between three and seven measurements, within a few mm of each other, were collected. For the multiple tests on each animal an average was taken for a given parameter and that data-point was used to compare that parameter across animals. All animals were conscious and mobile ~10 min post-testing. There was no sign of post-test pain or discomfort as assessed by visual inspection of animals during normal cage activity.

Raw data output from the RPI analysis software (version 2.0) were imported into a customized MATLAB code (Mathworks) (Aref et al., 2013). Primary variables of interest from the MATLAB program include first cycle indentation distance (ID-1st), which represents the depth the probe penetrated on the initial cycle; first cycle energy dissipation (ED-1st), which represents the energy dissipated in the first cycle; first cycle unloading slope (US-1st) which represents material stiffness (damage modulus) for the first cycle; indentation distance increase (IDI), which represents the penetration depth between the first and 10th cycle; total indentation distance (TID) which represents the distance from the bone surface to the depth of penetration after the 10th cycle; and total energy dissipation (Tot ED) which represents the total energy dissipation summed over all 10 cycles (Fig. 1). Our previous work has shown that parameters analyzed by the MATLAB software that were also generated by the manufacturer software yielded correlation coefficients of >0.96 (Aref). The advantage of the MATLAB program over the manufacturer software is that additional data, specifically cycle-by-cycle and energy data are generated.

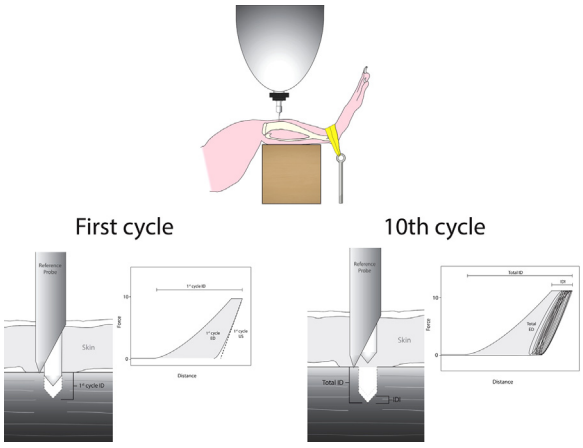

Fig. 1. *in vivo* testing set up and outcome parameters for RPI in skeletally mature rats. (A) The animals lower limb was flexed at the knee joint and placed on an elevated support so that the proximal tibial plateau was perpendicular to the testing probe. The foot was secured in place at the ankle and then a series of 10 cyclic indents were initiated where the test probe penetrates to a force of 10 N and then retracts. (B) Following the first cycle of the cyclic test, key outcomes of 1st cycle indentation distance (1st cycle ID), 1st cycle unloading slope (1st cycle US) and 1st cycle energy dissipation (1st cycle ED) can be calculated. Additional parmeters are obtained after the 10th cycle, including total indentation distance (Total ID), indentation distance increase (IDI) and energy dissipation (Total ED).

2.3. Data analyses

Intra-animal variability was assessed by calculating the coefficient of variation (CV) for all tests within an animal. Inter-animal variation was assessed by calculating CVs for each outcome parameter across all animals. Paired *t*-test analyses were used to compare baseline and 28 day data.

3. Results

A total of 319 tests were conducted in the 72 animals. Of these, 49 tests were deemed unsuccessful during testing based on the operators noting various problems with the tests. These included the test having a negative IDI (probe final position is above original reference position), decreasing displacement in first few cycles (resulting in a negative loading slope), or the measurement unit shifting during test. Upon removal of these unsuccessful tests, 71 animals had between 2 and 5 measures, and these were used for subsequent analyses (one animal was removed because it had only one acceptable measure).

Intra-animal variation of RPI parameters ranged from between 13.3% and 20.6% (Table 1, Fig. 2A). The least variable parameters within animals were US 1st and Tot ED 1st-L, each with a coefficient of variation of 13%. The most variable parameter within animals was IDI a CV of 20.6%. Inter-animal variation ranged from 16% to 25% (Table 2, Fig. 2B). The least variable parameter among animals was US-1st (CV=16%), while both ED-1st and IDI had the largest CVs of 25%.

One month following the initial RPI tests, a subset of animals ($n=17$) underwent a second RPI test on the contralateral limb. In this smaller dataset, intra-animal variation ranged from 45% to 74% with TID and ID-1st being the least variable parameters and US-1st the most variable. The inter-animal variability in this data set ranged from 13% to 23% with the least variable parameter being US-1st and the most variable being TID and ID-1st (data not shown).

Changes between baseline and one month measures were calculated to determine variability over time in untreated animals. All six parameters were, on average, lower at the second measurement relative to the first, with decreases ranging from −6% to −18% (Table 3). For each parameter, there was a wide range of responses with some animals increasing, some decreasing, and others unchanged (Fig. 3). There was no significant difference in any parameter between baseline and day 28.

Table 1
Intra-animal variation of RPI in skeletally mature male rats.

	1st Cycle Indentation Distance (ID 1st)	1st Cycle Energy Dissipated (ED 1st)	1st Cycle Unloading Slope (US 1st)	Indentation Distance Increase (IDI 1st-L)	Total Indentation Distance (TID 1st-L)	Total Energy Dissipated (Tot ED 1st-L)
Mean CV within animal, %	17.4	14.1	13.3	20.6	16.9	13.4
Standard deviation, %	10.4	10.8	13.0	14.9	10.2	9.3

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