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Rational design of cancer-targeted selenium nanoparticles to antagonize multidrug resistance in cancer cells

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6 Abstract

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Multidrug resistance is one of the greatest challenges in cancer therapy. Herein we described the design and synthesis of folate (FA)-7 conjugated selenium nanoparticles (SeNPs) as cancer-targeted nano-drug delivery system for ruthenium polypyridyl (RuPOP), which 8 exhibits strong fluorescence, which allows the direct imaging of the cellular trafficking of the nanosystem inside the cancer cells. This 9 nanosystem could effectively antagonize against multidrug resistance in liver cancer. FA surface conjugation significantly enhanced the 10cellular uptake of SeNPs by FA receptor-mediated endocytosis through nystain-dependent lipid raft-mediated and clathrin-mediated 11 pathways. The nanomaterials overcame the multidrug resistance in R-HepG2 cells through inhibition of ABC family proteins expression. 12 Internalized nanoparticles triggered ROS overproduction and induced apoptosis by activating p53 and MAPKs pathways. Moreover, FA-13 SeNPs exhibited low in vivo acute toxicity, which verified the safety and application potential of FA-SeNPs as nanodrugs. This study 14 15 provides an effective strategy for the design of cancer-targeted nanodrugs against multidrug resistant cancers.

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17 Key words: Multidrug resistance; Nanodrug delivery; Cancer targeting; Selenium nanoparticles

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19 Background

Multidrug resistance is becoming one of the most important 20obstacles for cancer therapy. P-glycoprotein (P-gp or ABCB1), 21adenosine triphosphate (ATP)-dependent active efflux pump, is 22often overexpressed in the plasma membrane of most multidrug 23 resistant cancer cells.¹⁻³ One of the most common malignancies 24in the world is hepatocellular carcinoma. Importantly, a serious 25obstacle for the successful treatment of liver cancer is the 26development of drug resistance. Nowadays, doxorubicin 27(DOX)-based combination chemotherapy is the main therapeutic 28strategy for hepatocellular carcinoma, but it failed to treat drug 29

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resistance cancers.⁴ What is more, some drugs, such as 30 cyclosporin-A (a calcium channel blocker)⁵ and verapamil (an 31 immunosuppressive peptide)⁶ are the two most studied agents to 32 reverse the drug resistance. However, they are not effective and 33 specific to P-gp overexpressing cancer cells. Until now, no 34 effective treatment is available for end-stage hepatocellular 35 carcinoma.⁷ Therefore, developing new therapeutic agents 36 which can overcome drug resistance for hepatocellular carcino- 37 ma cancer patient is urgently needed. 38

In order to overcome the multidrug resistant and reduce the 39 side effect, targeted nanodrug delivery systems were widely used 40 by improving the stimuli-triggered drug release and cancer-41 targeted drug delivery to minimize the side effects.^{8,9} Significantly, 42 the cancer targeting ligands could bind to their receptor on the 43 cancer cell membrane, which could enhance the selective 44 accumulation and uptake of the nanoparticles in the tumor-45 bearing organs, and reduce the toxicity toward normal cells at the 46 same time. Nanotechnology is now widely used, as a drug carrier for 47 cancer therapy.^{8,10} So far, many nanosystems with different 48 functions had been reported for cancer therapy, such as oxides, 49 metals, polymers, mesoporous silica and semiconductors.¹⁰⁻¹³ 50 Among them, selenium nanoparticles (SeNPs) receive more and 51 more attention as nanocarriers due to their biocompatibility, 52

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straightforward synthesis, low-toxicity, degradability in vivo, 53excellent antioxidant activity and chemopreventative effects, such 54as SeNPs used as 5-fluorouracil (5-FU) and DOX carriers.^{12,14} We 55had also reported cancer-targeted nanoparticles enhanced anticancer 56effects.^{10,14} But there was no report about multidrug resistance in 57our previous work. The folate receptor (FAR) is frequently 58overexpressed on cancers cells and has been used for targeted 59delivery of FA inked liposomes to cancer cells in vitro.¹⁵ FAR 60 transports the captured drugs into the cell by receptor-mediated 61 endocytosis and this has found use in cancer therapy by enhancing 62 the concentration of drugs in the cancer cells.¹⁶ Studies also found 63 the FA-targeted drugs could target specific cancer cells and 64 non-targeted the normal cells.¹⁷ Therefore, FA could be linked to 65 SeNPs to target FAR-overexpressing cancers. 66

The major limitation of cisplatin is the side effects in normal 67 tissues, which include neurotoxicity, ototoxicity, nausea and 68 vomiting, and especially nephrotoxicity.¹⁸ The serious limita-69 tions of cisplatin-based treatments have spurred scientists to 70 search for alternative metal-based anticancer drugs.^{19,20} Spe-71 cially, ruthenium (Ru) displays several favorable properties 72suitable for drug design and medicinal applications. Studies have 73shown that Ru complexes exhibited low cytotoxicity toward 74normal cells and high activity against tumor metastasis.²¹⁻²³ Till 75now, a number of Ru complexes have been synthesized and 76 identified as novel anticancer agents.²⁴ Among them, KP109 and 77 NAMI-A have already entered clinical trials.^{21,23,25,26} Previous-78ly we found the Ru complex RuPOP exhibited higher anticancer 79 activity and lower toxicity than cisplain.²¹ However, the use and 80 development of the RuPOP complex were limited by its poor 81 aqueous solubility. Therefore this study aimed to construct a 82 drug delivery system for hydrophobic Ru complexes conquering 83 their drawbacks. Following our investigation, we found that 84 pluronic F-127 was a good surface modification agent. The 85 pluronic is a group of block copolymers consisting of propylene 86 oxide and ethylene oxide with central hydrophobic poly 87 (propylene oxide) flanked by two hydrophilic chains of poly 88 (ethylene oxide).^{27,28} Pluronics could absorb hydrophobic drugs 89 by intermolecular forces and thus increase the loading rate of 90 91RuPOP. Our results confirmed that as-synthesized FA-SeNPs 92nanosystem could be used as a cancer-targeted carrier of RuPOP to enhance anticancer efficacy against multidrug resistant cancer 93 cells. The underlying mechanisms of FA-SeNPs were also 9495elucidated. Taken together, this study may provide an effective strategy for the design and development of nanodrugs against 96 multidrug resistant cancers. 97

98 Methods

99 Preparation of FA-SeNPs

100 The solution of 20 mM vitamin C, 5 mM of sodium selenite (Na₂SeO₃) solution and 0.8 mg/mL of chitosan (CS) solution 101 was freshly prepared before the experiment. Pluronic F-127 102 (2.5 g) was activated by 4-nitrophenyl chloroformate (4-NPC) 103(125 mg) in dichloromethane (10 ml) at room temperature for 104 12 h.²⁹ The reaction mixture was dialyzed against distilled water 105(DW) for 24 h and freeze-dried. Then added thioglycolic acid 04 (150 μ L) to the amine terminated pluronic in DW for 12 h with 107

stirring. The resultant solution was dialyzed against DW for 24 h 108 and freeze-dried. The preparation of RuPOP-loaded SeNPs was 109 as described in our previous works.¹⁴ 2.5 g thiolated pluronic 110 dissolved in 1 mL methanol and 500 μ L 0.1 M CS-FA were 111 added in stirring for 12 h at room temperature then dialyzed 112 against DW for 24 h. 113

Characterization of FA-SeNPs 114

The FA-SeNPs were characterized by different methods 115 including Zetasizer particle size, Fourier transform infrared 116 spectroscopy (FT-IR), transmission electron microscope (TEM), 117 UV-vis spectroscopy, and fluorescence spectroscopy analysis. 118 The TEM images were obtained at an accelerating voltage at 119 80 kV on Hitachi (H-7650). Zetasizer Nano ZS particle analyzer 120 (Malvern Instruments Limited) was used to measure size 121 distribution and zeta potential of the nanoparticles. 122

Determination of loading rate of RuPOP in FA-SeNPs 123

The concentration of Se and RuPOP was determined by 124 ICP-AES analysis. 125

Hemolysis activity examinations

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The hemolysis properties of FA-SeNPs were examined by 127 spectrophotometry as reported. 30,31 For the studies of erythro- 128 cyte agglutination, each sample was treated to a hemolysis assay 129 for 1 h (5 μM SeNPs, 5 μM RuPOP and 5 μM FA-SeNPs), 130 placed on a glass slide, covered by a cover slip and analyzed by a 131 phase contrast microscope (Life technologies, EVOS FL auto). 132

Cell lines and cell culture 133

HepG2 hepatocellular carcinoma cells, L02 human hepatic 134 cells, and R-HepG2 drug-resistant hepatocellular carcinoma cells 135 were purchased from American Type Culture Collection (ATCC, 136 Manassas, VA, USA). HepG2 and L02 were incubated in DMEM, 137 but R-HepG2 were incubated in 1640 with 100 U/ml penicillin, 138 50 U/ml streptomycin and 10% fetal bovine serum (FBS) in a 139 humidified incubator at 37 °C with 5% CO₂ atmosphere. 140

The effects of FA-SeNPs on a series of cancer cells were 142 examined by MTT assay. 32

In vitro cellular uptake of FA-SeNPs 144

The *in vitro* cellular uptake of FA-SeNPs was quantitatively 145 determined by measuring the fluorescence intensity of RuPO- 146 P-loaded nanoparticles inside the cells by using Spectra Max M5 147 Microplate reader (Bio-Tek).¹⁴ 148

Folate competing assay 149

FA-SeNPs and excess amount of FA competed for binding 150 FARs on R-HepG2 cells. The uptake of FA-SeNPs was then 151 measured by using Spectra Max M5 Microplate reader (Bio-Tek).¹⁴ 152 Download English Version:

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