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Isolation and cytotoxicity evaluation of taxanes from the barks of *Taxus wallichiana* var. *mairei*



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ABSTRACT

Fifteen taxanes (**1–15**) including a new taxane glucoside, 7 β ,9 α ,10 β -triacetoxyl-13 α -hydroxy-5 α -O-(β -D-glucopyranosyl)taxa-4(20),11-diene (**1**), were isolated from the barks of *Taxus wallichiana* var. *mairei*. Compounds **1–15** representing three sub-types of 6/8/6-taxane were evaluated in vitro for anti-proliferative activity against a panel of parental and drug-resistant cancer cells. Potent compounds were found while several exhibited selective cytotoxicity. Especially, **3**, **8**, and **10** showed selective inhibition to breast carcinoma cell line MCF-7, while **13** selectively inhibited taxol resistant human ovarian carcinoma cell line A2780/TAX (IC₅₀ = 0.19 μ M), being more potent than the clinical drugs taxol (IC₅₀ = 4.4 μ M) and docetaxol (IC₅₀ = 0.42 μ M), and less cytotoxic to mouse embryonic fibroblast cell line NIH-3T3, a cell line close to normal cell line. The possible P-glycoprotein evasion mechanism of **13** against A2780/TAX and the preliminary structure–activity relationships (SARs) of this group of compounds were also discussed.

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The discovery of paclitaxel (taxol) as a potent anticancer drug, initially isolated from *Taxus brevifolia*, has spurred several groups all over the world to conduct research work on other *Taxus* species, to isolate potentially more effective paclitaxel derivatives or as starting materials for semisynthesis. So far more than 550 taxanes have been identified, and new taxanes continue to be isolated from the needles, bark, stem, and roots of *Taxus* species.¹ Although taxol and its derivatives are widely used in clinical applications, the chemotherapy often fails due to its acquired-drug resistance, neuro-cytotoxicity, and low oral bioavailability. Thus, there is continuously demanding of new information facilitating the novel taxane-based drug design and discovery. However, previous pharmaceutical study on taxanes mainly focused on the modification of the taxol prototype with a 6/8/6-ring system,^{2–7} and systematically evaluation of the anticancer properties of taxanes with different sub-types remains scarce.

Taxus wallichiana var. *mairei* (Lemée & H. Léveillé) L. K. Fu & Nan Li, also known as *Taxus chinensis* (Pilger) Rehder var. *mairei* (Lemée & H. Léveillé) W.C. Cheng & L.K. Fu, an evergreen tree that is only distributed in China.⁸ Previous phytochemical studies on this plant

have led to the isolation a number of taxanes, some of which showed significant cytotoxicity.^{9–14} In our screening program aimed at the discovery of structurally unique and biologically significant taxanes from *Taxus* plants, a new taxane glucoside, 7 β ,9 α ,10 β -triacetoxyl-13 α -hydroxy-5 α -O-(β -D-glucopyranosyl)taxa-4(20),11-diene (**1**), and 14 known analogues (Fig. 1), 2-deacetoxytaxinine J¹⁵ (**2**), 2-deacetoxyaustrospicatin¹⁶ (**3**), cephalomannine¹⁷ (**4**), 10-deacetyl-7-*epi*-taxol¹⁸ (**5**), taxol¹⁹ (**6**), 7-xylosyl-10-deacetyl taxol²⁰ (**7**), 7- β -xylosyl-cephalomannine²⁰ (**8**), 7- β -xylosyl-taxol²⁰ (**9**), 7- β -xylosyl-10-deacetyl-cephalomannine²⁰ (**10**), 7-xylosyl-10-deacetyl taxol C²⁰ (**11**), taxinine M²¹ (**12**), taxacin²² (**13**), 10-deacetylbaicatin III²³ (**14**), and baicatin III^{17,20} (**15**) were isolated from the barks of *T. wallichiana* var. *mairei*.

Subsequent cytotoxic screening against a panel of cancer cell lines revealed that some of the 6/8/6-taxanes with different sub-types exhibited selective inhibition to some of the cell lines, such as the MCF-7 and taxol-resistant A2780/TAX. Herein, details of the isolation, structural elucidation, cytotoxic activity as well as a preliminary structure–activity relationship (SAR) of these compounds are described.

The air-dried barks of *T. wallichiana* var. *mairei* was extracted with refluxing MeOH to yield an extract, which was partitioned into EtOAc-soluble and EtOAc-insoluble fractions. After a series of column chromatography (CC) steps over silica gel and reversed-phase silica gel, followed by preparative TLC, the EtOAc-soluble

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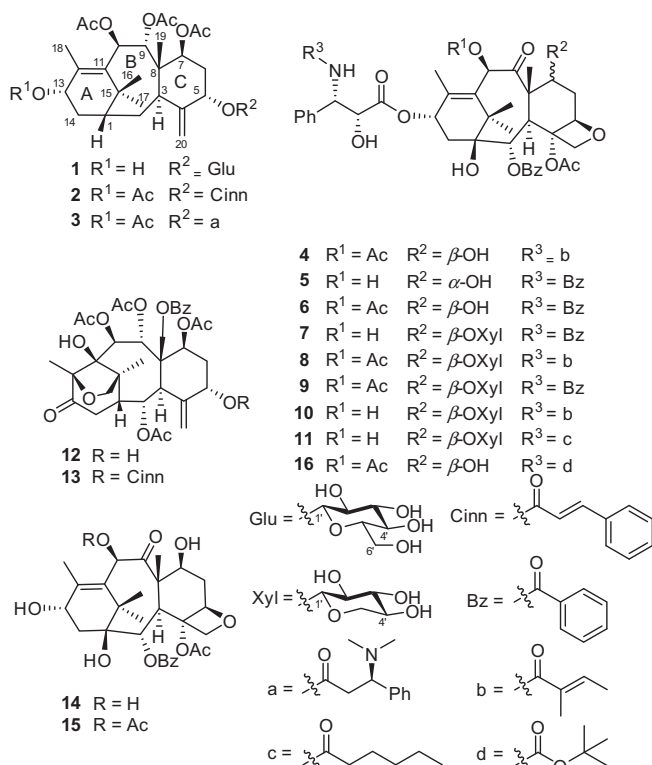


Figure 1. Structures of compounds **1–15** and clinical drug docetaxol (**16**).

fraction afforded compounds **1–15** (Supplementary data, experimental section).

Compound **1**²⁴ was isolated as a colorless oil. The molecular formula of C₃₂H₄₈O₁₃ was determined by the HRESIMS ion at *m/z* 663.2959 [M+Na]⁺ (calcd for 663.2987) and ¹³C NMR data. The 1D NMR spectra (Table 1) resolved 32 carbon resonances attributable to four methyls (δ_H 0.81, 1.01, 1.56, and 2.29; δ_C 13.7, 16.1, 27.7, and 31.5), three acetoxys (δ_H 1.93, 2.01, and 2.03; δ_C 21.0, 21.3, 21.4, 169.4, 170.2, and 170.7), a pair of exoethylene protons [δ_H 4.86 (1H, s) and 5.16 (1H, s)], and a glucose moiety [δ_H 4.26 (1H, d, *J* = 7.7 Hz); δ_C 63.0 (CH₂), 71.6 (CH), 74.2 (CH), 77.4 (CH), 77.5 (CH) and 104.8 (CH)]. The aforementioned data were similar to those of the known taxane taxezopidine H²⁵ except that the cin-

Table 1

¹H and ¹³C NMR data of **1** in acetone-d₆^a

No.	δ _H	δ _C	No.	δ _H	δ _C
1	1.80 (1H, m)	41.4	16	1.01 (3H, s)	31.5
2	1.81 (1H, m)	28.1	17	1.56 (3H, s)	27.7
3	2.96 (1H, d 4.7)	37.9	18	2.29 (3H, s)	16.1
4		150.7	19	0.81 (3H, s)	13.7
5	4.23 (1H, dd 3.7, 2.0)	81.9	20	4.86 (1H, s)	112.9
				5.16 (1H, s)	
6	1.75 (1H, m)	36.9	7-OAc	2.01 (3H, s)	21.4, 170.2
7	5.60 (1H, dd 11.5, 5.3)	71.2	9-OAc	2.03 (3H, s)	21.3, 170.7
8		47.0	10-OAc	1.93 (3H, s)	21.0, 169.4
9	5.84 (1H, d 11.1)	77.5	1'	4.26 (1H, d 7.7)	104.8
10	6.20 (1H, d 11.1)	73.3	2'	3.21 (1H, m)	74.2
11		133.7	3'	3.36 (1H, m)	77.4
12		142.7	4'	3.33 (1H, m)	71.6
13	4.71 (1H, m)	68.5	5'	3.28 (1H, m)	77.5
14	1.19 (1H, m),	35.0	6'	3.63 (1H, m)	63.0
	2.63 (1H, m)			3.81 (1H, m)	
15		39.9			

^a Data were recorded at 400 MHz for ¹H NMR and 100 MHz for ¹³C NMR. (*J* in Hz, δ in ppm).

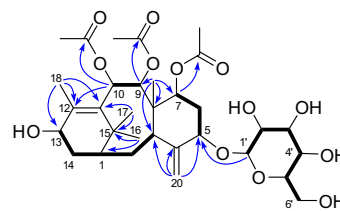


Figure 2. Key HMBC (H→C) and ¹H–¹H COSY (—) correlations of **1**.

namoyl unit in taxezopidine H was replaced by a glucose moiety in **1**. This was supported by the downfield-shifted C-5 signal (δ_C 81.9) in **1** due to the glycosylation shift effect (C-5 at δ_C 75.2 in taxezopidine H). The *D*-configuration of the glucosyl unit was determined by HPLC analysis.²⁶ The gross structure of **1** was further confirmed by detailed 2D NMR analysis including HMBC and COSY spectra (Fig. 2).

The relative configuration of **1** was determined by analysis of its ¹H–¹H coupling constant and NOESY data. The fusion patterns of A/B/C ring system and the orientations of the substituents (acetyl and hydroxyl groups) on rings A and B were assigned as the same as those of taxezopidine H by comparison of their 1D NMR data. The NOESY correlations of H-7/H-3 and CH₃-19/H-6β indicated that H-7, H-3, CH₃-19, and H-6β adopted the axial bonds of the quasi chair-conformational ring C. The small coupling constant of H-5 [δ_H 4.23 (dd, *J* = 3.7 and 2.0 Hz)] indicated that H-5 adopted equatorial bond and was in β-orientation. Thus, compound **1** was assigned as 7β,9α,10β-triacetoxy-13α-hydroxy-5α-*O*-(β-*D*-glucopyranosyl)taxa-4(20),11-diene.

The cytotoxicity of compounds **1–5** and **7–15** were evaluated by sulforhodamine B (SRB) assay with docetaxol (**16**) and taxol (**6**) used as positive controls (Supplementary data, experimental section). Ten human cancer cell lines including human ovarian carcinoma cell A2780 with the taxol resistant cell A2780/TAX, human ileocecum carcinoma cell HCT-8 with the vincristine resistant cell HCT-8/VCT, breast carcinoma cell line MCF-7 with the doxorubicin resistant cell MCF-7/DOX, human lung carcinoma cell line A549 with the *cis*-platinum resistant cell A549/CDDP, colon carcinoma cell SW480, and hepatoma carcinoma cell line HepG2 were chosen for this biological activity assay. The experiments were conducted in three independent replicates.

The bioassay results (Table 2) showed that most of the taxanes exhibited moderate to good inhibitory activity (IC₅₀ <10 μM) against some of the cell lines, and compounds **4** and **5** sharing the most structural features with taxol showed broad inhibitory effects against most of the cell lines, with the activities comparable to those of the positive control (most IC_{50s} <0.1 μM). Compared to **4–6** and **16**, the introduction of an *O*-β-xylose in R² (**7–11**) or loss of the C-13 ester side chain (**14–15**) greatly decrease their activities, which is consistent with the reported SAR of taxol analogues.²⁷

Interestingly, despite of the general low activity, the *O*-β-xylose bearing compounds **7–11** reserved their anti-proliferative activity and showed selective cytotoxicity towards certain cancer cell lines. Compounds **8** and **10** selectively inhibited MCF-7 with IC₅₀ values at 0.029 and 0.14 μM, respectively. Further analysis showed *N*-butanoylphenylisoserine substitution at R³ in **8** and **10** led to generally lower cytotoxicity to most of cell line but more selective and cytotoxic to MCF-7, as compared to those with phenyl substitution in **7** and **9**. Meanwhile, **11** selectively inhibited drug resistant cell lines A549/CDDP (IC₅₀ = 0.64 μM) and A2780 (IC₅₀ = 1.5 μM), respectively.

Compounds **1–3**, **12**, and **13**, which possess Δ^{4,20} terminal double bonds, showed greatly decreased activities to most of cancer cell lines. However, selective cytotoxicity was also observed within

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