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The synthesis and BK channel-opening activity of *N*-acylaminoalkyloxime derivatives of dehydroabietic acid



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ABSTRACT

A series of *N*-acylaminoalkyloxime derivatives of dehydroabietic acid were synthesized and evaluated for BK channel-opening activities in an assay system of CHO-K1 cells expressing hBK α channels. The structure-activity relationship study revealed that a non-covalent interaction between the S atom of the 2-thiophene and the carbonyl O atom may contribute to conformation restriction for interaction with the ion channel. This research could guide the design and synthesis of novel abietane-based BK channel opener.

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Large-conductance calcium-activated K⁺ channels (also called maxi-K or BK channels) characteristically respond to two distinct physiological stimuli, that is, changes in membrane voltage and in cytosolic Ca²⁺ concentration, and may couple with other ion channels (such as Ca²⁺ ion channels,^{1,2} chloride channel,³ TRPC channels⁴) to serve as a negative feedback pathway controlling ionic homeostasis, cell excitability, and neuron activity.⁵ BK channels consist of channel-forming α -subunits and accessory β -subunits (β_1 – β_4) arranged in tetramers,⁶ having a voltage sensor and pore as the membrane-spanning domain and having a cytosolic domain containing metal-binding sites. Recently published studies on electron cryomicroscopy (cryo-EM)⁷ and X-ray crystallographic structure analysis⁸ of the BK channel have provided the first glimpse into the assembly of the quaternary structure of this massive channel protein, corroborating the close interactions among these domains during channel gating that were suggested by the previous functional studies.⁹ Recent cloning studies have revealed the presence of multiple splice variants of α -subunits^{10–12} and multiple subtypes of β -subunits (β_1 , β_2/β_3 and β_4).¹³ Thus, there is a large diversity of BK channels, which may be specific to tissues and organs. The BK channels are expressed in a number of organ systems, such as smooth muscle cells, skeletal muscle cells, neuronal

cells, and secretory epithelial cells,¹⁰ and they have important physiological roles in modulating muscle contraction or neurotransmitter release and hormone secretion.¹⁴

The physiological role and widespread distribution of BK channels suggest that agents that open these channels could have profound impacts on diseases such as ischemic stroke, epilepsy, asthma, and bladder overactivity.¹⁵ During the past few years, various classes of BK channel openers as well as their chemistry and pharmacology have been described.^{16–19} Well-characterized BK channel openers not only are expected to have therapeutic potential, but also should be of assistance in understanding the function, structure and role of BK channels.

Our previous study showed that the dehydroabietic acid (DHAA, **1**, Fig. 1) structure provides a template for chemical modulators of BK channels.²⁰ By introducing an oxime ether chain in position 7 of the dehydroabietane skeleton, we obtained compounds such as CYM04 with BK channel-opening activity comparable to that of NS1619 and its mechanism of action has been studied.^{21–23} While several interaction models²³ of chemical openers with BK channels have been proposed, binding site and binding mode of chemical openers are still unrevealed. Most probable binding sites may be present in the transmembrane helices (TM) constituting the channel pore (such as TM5 (S5) and TM6 (S6)),²⁴ however, the intracellular domain such as S6-RCK1 linker has been proposed as a binding site of CYM04.²³ In this context, shape and contiguity of hydrophobic moiety of the chemical BK channel openers are

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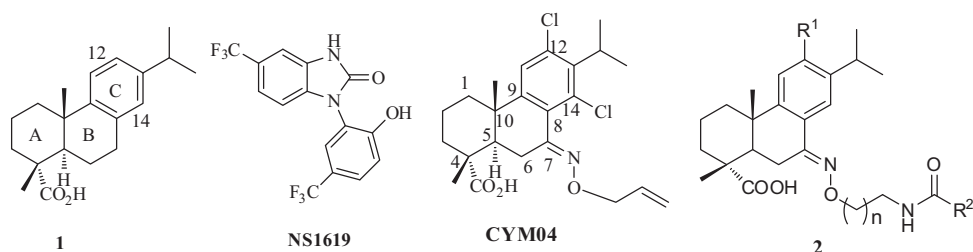
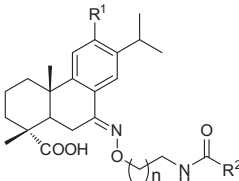
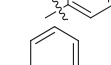
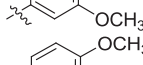
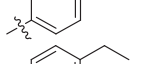
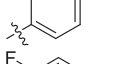
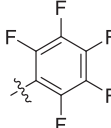
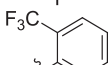
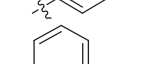
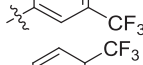
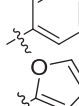


Figure 1. The structures of dehydroabietic acid (**1**), NS 1619, CYM04 and **2**.

Table 1

Structure and BK α -opening properties of dehydroabietate derivatives

				
Compound	R ¹	n	R ²	Ionic current in the presence of test compound (30 μ M) as % of control current (n = 8)
Buffer	—	—	—	104.0 $\pm$ 2.6
NS1619	—	—	—	267.5 $\pm$ 45.0
CYM04	—	—	—	287.5 $\pm$ 65.8
7a	Br	1	—	98.6 $\pm$ 15.0
9a	Br	1	CH ₃	139.5 $\pm$ 26.3
9b	Br	1		141.8 $\pm$ 17.8
9c	Br	1		146.4 $\pm$ 13.8
9d	Br	1		148.8 $\pm$ 32.4
9e	Br	1		121.4 $\pm$ 19.6
9f	Br	1		101.2 $\pm$ 4.7
9g	Br	1		90.4 $\pm$ 3.6
9h	Br	1		102.4 $\pm$ 11.0
9i	Br	1		68.0 $\pm$ 3.6
9j	Br	1		118.0 $\pm$ 40.0
9k	Br	1		136.4 $\pm$ 33.3
9l	Br	1		123.0 $\pm$ 13.8
9m	Br	1		112.7 $\pm$ 5.0
9n	Br	1		107.0 $\pm$ 7.9
9o	Br	1		151.3 $\pm$ 20.4

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