



Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl

Synthesis, binding affinity, radiolabeling, and microPET evaluation of 4-(2-substituted-4-substituted)-8-(dialkylamino)-6-methyl-1-substituted-3,4-dihydropyrido[2,3-*b*]pyrazin-2(1*H*)-ones as ligands for brain corticotropin-releasing factor type-1 (CRF₁) receptors



Jeffrey S. Stehouwer^{a,*}, Chase H. Bourke^b, Michael J. Owens^b, Ronald J. Voll^a, Clinton D. Kilts^c, Mark M. Goodman^{a,b}

^a Center for Systems Imaging, Department of Radiology and Imaging Sciences, Emory University, WWHC 209, 1841 Clifton Rd NE, Atlanta, GA 30329, USA

^b Department of Psychiatry and Behavioral Sciences, Emory University, Atlanta, GA, USA

^c Psychiatric Research Institute, University of Arkansas for Medical Sciences, Little Rock, AR, USA

ARTICLE INFO

Article history:

Received 13 August 2015

Revised 1 October 2015

Accepted 5 October 2015

Available online 8 October 2015

Keywords:

Corticotropin-releasing factor

CRF-1 receptor

Fluorine-18

PET imaging

Positron emission tomography

ABSTRACT

Compounds **1–14** were synthesized in a search for high-affinity CRF₁ receptor ligands that could be radiolabeled with ¹¹C or ¹⁸F for use as positron emission tomography (PET) radiotracers. Derivatives of **2** were developed which contained amide *N*-fluoroalkyl substituents. Compounds [¹⁸F]**24** and [¹⁸F]**25** were found to have appropriate lipophilicities of log *P*_{7,4} = 2.2 but microPET imaging with [¹⁸F]**25** demonstrated limited brain uptake.

© 2015 Elsevier Ltd. All rights reserved.

The neuropeptide corticotropin-releasing factor (CRF)^{1,3} coordinates the neuroendocrine response to stress through the hypothalamic–pituitary–adrenal axis by acting on the CRF Type-1 (CRF₁) receptor.⁴ Dysregulation of brain CRF signaling has been proposed to be involved in stress, anxiety, depression, and addiction; and brain regional alterations in CRF₁ density have been detected in depressed patients and victims of suicide.^{5–8} This evidence has resulted in a more than 20-year search for small molecule, brain bioavailable CRF₁ antagonists that could be used as therapeutics for treating CRF₁-related disorders.^{9–11} Therapeutic medication development can be uniquely aided by the availability of validated positron emission tomography (PET) and single-photon emission computed tomography (SPECT) radiotracers that can be used to both support target occupancy measurements of candidate

therapeutics and to explore mechanisms of pathophysiology and treatment response by measurement of receptor density and expression levels.^{12–15} Numerous attempts to develop a viable brain CRF₁ receptor PET radiotracer have been reported over the years but no candidates to date have been successful due to problems such as high lipophilicity and inability to cross the blood–brain barrier (BBB), rapid metabolism, or insufficient specific receptor binding.^{16–19} As part of an effort to develop a viable brain CRF₁ receptor PET tracer we have been exploring various structural motifs²⁰ and report here our results with compounds **1–15** (Scheme 1), and derivatives thereof, which are based on a previous report that compounds **2**, **6**, and **9** displayed high binding affinity (IC₅₀ = 0.70, 0.49, and 0.92 nM, respectively) at the rat brain CRF₁ receptor.²¹ We anticipated that the reported high binding affinity of **2**, **6**, and **9** would enable radiotracer derivatives of this structural class to achieve specific binding at CRF₁ and that the presence of the carbonyl group would enhance water solubility, thereby reducing log *P* and increasing BBB passage.^{22–25} Furthermore, the synthetic route for this class of compounds²¹ (Scheme S1, Supplementary material) allows the aromatic core and the *N,N*-dialkyl group to be held constant while the

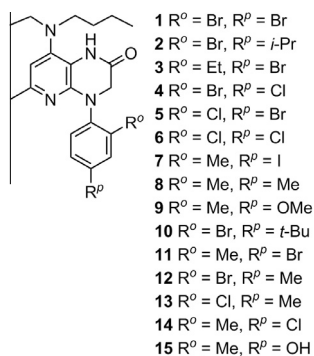
Abbreviations: CRF, corticotropin-releasing factor; CRF₁, CRF type-1 receptor; PET, positron emission tomography; SPECT, single-photon emission computed tomography; BBB, blood–brain barrier; DMA, dimethylacetamide; SUV, standard uptake value; TACs, time–activity curves; CPCU, chemical processing control unit; rcy, radiochemical yield; EOB, end-of-bombardment.

* Corresponding author.

E-mail address: jstehou@emory.edu (J.S. Stehouwer).

<http://dx.doi.org/10.1016/j.bmcl.2015.10.010>

0960-894X/© 2015 Elsevier Ltd. All rights reserved.



Scheme 1.

2,4-substituted pendant aryl ring is varied to produce a library of high-affinity candidates that are amenable to radiolabeling with ¹¹C or ¹⁸F.

The binding affinities of compounds **1–14** at the CRF₁ and CRF₂ receptors were determined by in vitro competition binding assays at 23 °C using HEK293T cells transfected with the human CRF₁ or CRF₂ receptor (Table 1). Compounds **1–14** all bound to the CRF₁ receptor with high affinity ($K_i = \sim 1.3$ – 5.4 nM). Thus, although these compounds have various combinations of substituents on the pendant aryl ring, they all have binding affinities within a narrow range that differs by ~ 4 nM. The difference in affinities of **2**, **6**, and **9** between our results and the previously reported results²¹ is presumably due to the difference between human and rat CRF₁ receptors as well as the fact that we are reporting K_i values and the previous work reported IC₅₀ values. Sauvagine and R121919²⁶ were used as positive controls in our assays: R121919 was found to have an affinity of $K_i = 2.6 \pm 0.4$ nM for the CRF₁ receptor which is comparable to the previously reported values of $K_i = 3.5$ nM²⁶ and $K_i = 3.0 \pm 0.16$ nM;²⁷ and sauvagine was found to have an affinity of $K_i = 0.8 \pm 0.1$ nM which is in agreement with the previously reported value of $K_i = 0.8$ – 1 nM.²⁸ The competing radioligand for these studies, [¹²⁵I]-Tyr⁰-sauvagine, has a reported affinity of 0.2–0.4 nM for the CRF₁ receptor.²⁸ Compounds **1–3**, **6**, and **8–12**

Table 1

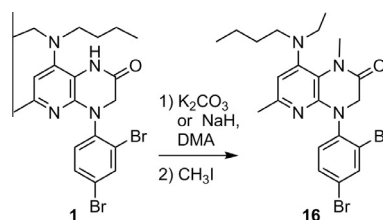
Results of in vitro competition binding assays at 23 °C using transfected human CRF₁ and CRF₂ receptors in HEK293T cells

Compd	R ^O	R ^P	CRF ₁		CRF ₂	
			K_i (nM) \pm SEM ^a	<i>n</i> =	% Displacement ^b	<i>n</i> =
1	Br	Br	1.3 \pm 0.3	3	9.4 \pm 2.3	2
2	Br	<i>i</i> -Pr	2.4 \pm 0.2	3	0.7 \pm 3.3	2
3	Et	Br	2.5 \pm 0.7	6	2.1 \pm 2.5	2
4	Br	Cl	2.5 \pm 0.2	3	N/D	N/A
5	Cl	Br	2.7 \pm 0.4	3	N/D	N/A
6	Cl	Cl	3.4 \pm 0.4	3	0.9 \pm 3.4	2
7	Me	I	3.4 \pm 0.6	3	N/D	N/A
8	Me	Me	3.9 \pm 1.2	3	6.8 \pm 1.4	2
9	Me	OMe	4.0 \pm 0.6	3	–2.3 \pm 3.2	2
10	Br	<i>t</i> -Bu	4.2 \pm 2.2	3	–3.8 \pm 0.5	2
11	Me	Br	4.4 \pm 0.9	3	5.9 \pm 0.7	2
12	Br	Me	4.4 \pm 1.1	3	5.5 \pm 4.9	2
13	Cl	Me	4.4 \pm 0.4	3	N/D	N/A
14	Me	Cl	5.4 \pm 0.9	3	N/D	N/A
R121919	N/A	N/A	2.6 \pm 0.4	9	23.8 \pm 0.3	2
Sauvagine	N/A	N/A	0.8 \pm 0.1	9	100.0 \pm 0.0	2
Antisauvagine-30	N/A	N/A	N/D	N/A	99.0 \pm 0.6	2

N/D = not determined. N/A = not applicable.

^a Versus [¹²⁵I]-Tyr⁰-sauvagine.²⁸

^b Mean% displacement of radiotracer ([¹²⁵I]antisauvagine-30)²⁹ specific binding at CRF₂ receptors by 1 μ M of competing ligand.



Scheme 2.

Table 2

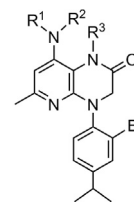
Results of in vitro competition binding assays at 23 °C using transfected human CRF₁ receptor in HEK293T cells

Compd	R ^O	R ^P	R ¹	R ²	R ³	K_i (nM) \pm SEM ^a	<i>n</i> =
16	Br	Br	Bu	Et	Me	2.4 \pm 0.6	2
17	Br	<i>i</i> -Pr	Me	Me	H	180 \pm 5	2
18	Br	<i>i</i> -Pr	Me	Et	H	16.4 \pm 1.6	3
20	Br	<i>i</i> -Pr	Pr	Pr	H	3.1 \pm 0.3	3
21	Br	<i>i</i> -Pr	Me	Me	CH ₂ CH ₂ F	59 \pm 9	3
22	Br	<i>i</i> -Pr	Et	Et	CH ₂ CH ₂ F	7.3 \pm 0.7	3
23	Br	<i>i</i> -Pr	Me	Me	(<i>E</i>)-CH ₂ CH=CHCH ₂ F	14.2 \pm 2.9	3
24	Br	<i>i</i> -Pr	Me	Me	CH ₂ CH ₂ CH ₂ CH ₂ F	21.6 \pm 2.8	3
25	Br	<i>i</i> -Pr	Me	Et	CH ₂ CH ₂ CH ₂ CH ₂ F	13.9 \pm 1.9	3

^a Versus [¹²⁵I]-Tyr⁰-sauvagine.²⁸

did not display binding affinity at the CRF₂ receptor (they were unable to significantly displace [¹²⁵I]antisauvagine-30),²⁹ thus demonstrating selectivity for the CRF₁ receptor and so further screening of compounds **4**, **5**, **7**, **13**, and **14** at the CRF₂ receptor was not performed.

Compound **1** was found to have the highest binding affinity for the CRF₁ receptor followed by compounds **2–5**, but these compounds do not have a position available for radiolabeling with ¹¹C or ¹⁸F. Thus, we shifted our focus to derivatives that could potentially be radiolabeled on the amide nitrogen atom. Compound **16** (Scheme 2) was prepared by amide *N*-methylation of **1** with CH₃I. This resulted in only a small decrease in CRF₁ binding affinity (Table 2) relative to **1** (Table 1). Compounds **17–20** (Scheme 3) were synthesized (Scheme S2, Supplementary material) to be used as intermediates in the synthesis of amide *N*-fluoroalkyl target compounds. The shorter *N,N*-dialkyl chains of **17–19** were expected to compensate for the increased lipophilicity that would result from adding alkyl groups to the amide nitrogen atom, while **20** was included to evaluate the effect on binding affinity of rearranging



- 17** R¹ = Me, R² = Me, R³ = H
18 R¹ = Me, R² = Et, R³ = H
19 R¹ = Et, R² = Et, R³ = H
20 R¹ = Pr, R² = Pr, R³ = H
21 R¹ = Me, R² = Me, R³ = CH₂CH₂F
22 R¹ = Et, R² = Et, R³ = CH₂CH₂F
23 R¹ = Me, R² = Me, R³ = (*E*)-CH₂CH=CHCH₂F
24 R¹ = Me, R² = Me, R³ = CH₂CH₂CH₂CH₂F
25 R¹ = Me, R² = Et, R³ = CH₂CH₂CH₂CH₂F
26 R¹ = Me, R² = Me, R³ = CH₂CH₂CH₂CH₂OTs
27 R¹ = Me, R² = Et, R³ = CH₂CH₂CH₂CH₂OTs

Scheme 3.

Download English Version:

<https://daneshyari.com/en/article/10590829>

Download Persian Version:

<https://daneshyari.com/article/10590829>

[Daneshyari.com](https://daneshyari.com)