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A dual functional fluorescent probe for glioma imaging mediated by BBB penetration and glioma cell targeting

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ABSTRACT

Glioma is a huge threat for human being because it was hard to be completely removed owing to both the infiltrating growth of glioma cells and integrity of blood brain barrier. Thus effectively imaging the glioma cells may pave a way for surgical removing of glioma. In this study, a fluorescent probe, Cy3, was anchored onto the terminal of AS1411, a glioma cell targeting aptamer, and then TGN, a BBB targeting peptide, was conjugated with Cy3-AS1411 through a PEG linker. The production, named AsT, was characterized by gel electrophoresis, ^{1}H NMR and FTIR. In vitro cellular uptake and glioma spheroid uptake demonstrated the AsT could not only be uptaken by both glioma and endothelial cells, but also penetrate through endothelial cell monolayer and uptake by glioma spheroids. In vivo, AsT could effectively target to glioma with high intensity. In conclusion, AsT could be used as an effective glioma imaging probe.

1. Introduction

Brain tumor is an increasingly serious threat for human being. Among all original brain tumors, glioma accounts for 80% and is characterized by high mortality [1]. Due to the infiltration growth of glioma cells, completely removing glioma cells by surgery is an impossible mission and the reserved glioma cells often, if not absolutely, lead to recurrence. Although surgery resection followed with chemotherapy/radiotherapy may expand the median survival time, it still is restricted at approximately 15 months [2]. Thus accurate imaging of glioma cells is urgent for complete removing of glioma cells by surgery, which may decrease the recurrence rate and further improve life span of glioma bearing patients.

At present, fluorescent imaging-aid surgical resection of tumor cells has been come into reality in peripheral tumor [3]. However, these imaging probes could not be used for glioma cells imaging because of the infiltrated growth of glioma cells and the integrity of blood brain barrier (BBB) in the infiltrated area which restricted the distribution of these probes from blood to brain and brain glioma [4]. To conquer this barrier and specific target to glioma cells, probes must be recognized by both BBB and glioma cells. Fortunately, there are many receptors that overexpressed on BBB or glioma cells, which ligands were often used in glioma targeting delivery. TGN is a peptide that selected from a library by phage display, which showed high BBB targeting efficiency and has been used for brain targeting delivery with well outcomes [5,6]. Thus we chose TGN as part of our probe to enable the probe penetrating through BBB. Beside, nucleolin is overexpressed on glioma cells [7]. Its corresponding ligand, AS1411, is a G-riched aptamer and has been utilized as targeting ligand to mediate nanoparticles target to glioma [8,9]. Taking the advantages of TGN and AS1411 together, we conjugated TGN and AS1411 for glioma imaging.

In this study, TGN and AS1411 were conjugated through a dual functional PEG linker. To track the probe, shorted as AsT, Cy3 was anchored onto the 3′ end of AS1411. Several experiments, including cellular uptake, tumor spheroids uptake, ex vivo imaging and slices distribution, were carried out to evaluate the glioma targeting effect of AsT.

2. Materials and methods

2.1. Materials

AS1411, Cy3-AS1411 and TGN were synthesized by Sangon (Shanghai, China). Maleimide-polyethyleneglycol-N-hydroxysuccinimide (MAL-PEG-NHS) was purchased from Seebio Biotech, Inc. (Shanghai, China). C6, U87, BG2, A549, HUVEC and Raw246.7 cell lines were purchased from the Institute of Biochemistry and

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Cell Biology, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences (Shanghai, China). bEnd.3 cell line was purchased from American type culture collection (ATCC) (VA, USA). Dulbecco's Modified Eagle Medium (high glucose) cell culture medium (DMEM) and FBS were purchased from Life Technologies (NY, USA). Cell culturing dishes and plates were purchased from Wuxi NEST Biotechnology Co. Ltd. (Wuxi, China). Rabbit anti-nucleolin antibody was purchased from Abcam (Hong Kong, China). Alexafluor 488-conjugated donkey anti rabbit antibody was purchased from Jackson Immunoresearch Laboratories, Inc. (PA, USA). Western blot related reagents were pruchased from Beyotime (Haimen, China).

BALB/c mice (male, 4–5 weeks, 18–22 g) were obtained from Dashuo Bio technology Co., Ltd, (Chengdu, China) and maintained under standard housing conditions. All animal experiments were carried out in accordance with protocols evaluated and approved by the ethics committee of Sichuan University.

2.2. AsT synthesis and characterization

AS1411 was functioned with TGN utilizing Maleimide-PEG-NHS as a linker (Fig. 1). Ten OD of 5′ NH_2 modified As1411 was added with 254 μg of Maleimide-PEG-NHS in 1 mL pH 8.0 PBS and stirred 3 h for conjugation. After changed the pH of PBS from 8.0 to 7.0, the mixture was added with 66 μg TGN and stirred another 24 h for the TGN conjugation. Cy3 modified AsT (Cy3-AsT) was prepared as above except 5′ NH_2 modified As1411 changed to 5′ NH_2 and 3′ Cy3 modified As1411. To character the conjugation, production was applied to ^1H NMR using a400 MHz spectrometer (Varian, CA, USA). Fourier transform infrared spectroscopy (FTIR) was also used to characterize the AsT. Additionally, 0.5% agarose gel electrophoresis was used to further determine the conjugation of AS1411 with TGN.

2.3. Nucleolin expression

C6, HUVEC, bEnd.3, BG2, Raw246.7, U87 and A549 cells were lysed using RPMI buffer containing 1 mM PMSF. The protein concentration was determined using the BCA method. Equivalent amounts of protein were boiled for 5 min in loading buffer and then separated by 10% SDS polyacrylamide gel electrophoresis. Proteins were transferred to nitrocellulose membranes. The membranes were blocked for 1 h in TBS containing 4% low-fat milk. Membranes were then probed with specific antibodies recognizing the target proteins, and the proteins were visualized using an ECL reagent.

2.4. Cell uptake of AsT

C6 glioma cells and bEnd.3 cells were seeded into 24-well plates at the density of 10^4 per 400 μL/well. One day later, 1 OD/mL Cy3-AsT or Cy3-As1411 was added to cells and incubated for 1 h. After washed by ice-cold PBS for three times, the fluorescence intensity was observed by fluorescent microscopy (Leica, Germany).

2.5. Three dimension tumor spheroid uptake

To prepare three diamensions tumor spheroids, C6 cells were seeded at a density of 2×10^3 cells/400 μL per well in 48-well plates coated by 150 μL 2% low melting-temperature agarose [10]. Seven days after cell seeded, tumor spheroids were added with Cy3-AsT or Cy3-AS1441 at a concentration of 1 OD/mL. After 4 or 8 h incubation, spheroids were rinsed with ice-cold PBS for three times and fixed by 4% paraformaldehyde for 30 min. Then the spheroids were transferred to glass slides and fluorescent intensity was observed by confocal microscopy (SP5, Leica, Germany).

To determine the dual targeting effect of AsT, bEnd.3 cells were seeded in 24-well Transwell membrane at a density of 2×10^4 cells/200 μL per well. Transendothelial electrical resistance (TEER) was detected by an epithelial voltohmmeter (Millicel-RES, Millipore, USA) to evaluate the cell monolayer integrity. Monolayers with TEER over 200 Ω/cm^2 were used for further experiment. Three dimensions tumor spheroids were prepared as above. Six days later, bEnd.3 monolayers contained Transwell were transferred to the 48-well plate that cultured C6 spheroids and co-cultured for another one day. One OD/mL of Cy3-AsT or Cy3-AS1411 were added into the Transwell and incubated for 8 h. Then the spheroids were rinsed and fixed. The fluorescent intensity was observed by confocal microscopy (SP5, Leica, Germany).

2.6. Ex vivo imaging and tissue distribution

Glioma bearing mice were established as described previously [11]. Mice were anesthetized and 5 μL suspension containing 5×10^5 C6 cells was injected into the right corpus striatum of the mice. Ten days later, 300 OD/kg of Cy3-AsT or Cy3-AS1411 were i.v. injected into the glioma bearing mice. Two hours later, mice were sacrificed and perfused. Then the tissues were sampled and the distribution of fluorescence was observed by an IVIS spectrum in vivo imaging system (Caliper, MA, USA). After fixed with 4% paraformaldehyde overnight, the tissues were dehydrated for preparing frozen section with 10 μm thicknesses. Nucleon was

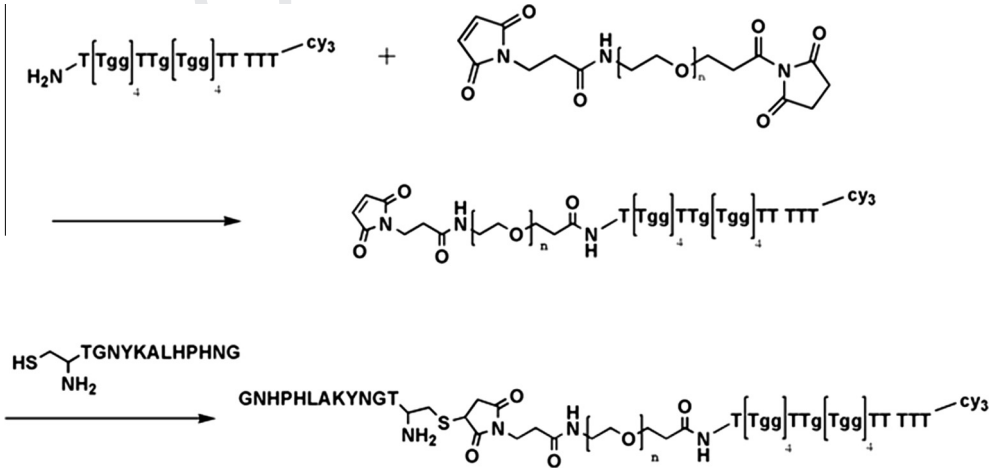

Fig. 1. Synthesis route of AsT.