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1 Review

$_2$ Bile acid signaling and liver regeneration $\stackrel{ riangle}{\sim}$

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ABSTRACT

The liver is able to regenerate itself in response to partial hepatectomy or liver injury. This is accomplished by a 18 complex network of different cell types and signals both inside and outside the liver. Bile acids (BAs) are recently 19 identified as liver-specific metabolic signals and promote liver regeneration by activating their receptors: 20 Farnesoid X Receptor (FXR) and G-protein-coupled BA receptor 1 (GPBAR1, or TGR5). FXR is a member of the nu- 21 clear hormone receptor superfamily of ligand-activated transcription factors. FXR promotes liver regeneration 22 after 70% partial hepatectomy (PHx) or liver injury. Moreover, activation of FXR is able to alleviate age-related 23 liver regeneration defects. Both liver- and intestine-FXR are activated by BAs after liver resection or injury and 24 promote liver regeneration. TGR5 regulates BA hydrophobicity and stimulates BA excretion in urine 26 activated during liver regeneration. BA signaling thus represents a novel metabolic pathway during liver regeneration. 27 This article is part of a Special Issue entitled: Nuclear receptors in animal development. 28

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34 1. Introduction

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Liver is one of the few organs that can powerfully regenerate itself 35in response to partial ablation or liver injury. Liver regeneration has 36 been widely studied as a paradigm for regenerative organ regrowth 37 since the introduction of a rodent partial hepatectomy (PHx) 38 model in 1931 [1]. Unlike a typically anatomic regeneration, regen-39 eration of the liver is a compensatory hyperplasia of the remaining 40 tissues and is driven by the functional deficit of the organism. Liver 41 regeneration consists of several well orchestrated phases, with 42 43 rapid induction of proliferating factors activating the quiescent hepatocytes and priming their subsequent progression through the 44 cell cycle, followed by re-establishment of normal liver size and 45renewed quiescence [2-4]. Growth factors and cytokines are the 46 47 important early signals to induce the expression of downstream target genes via activation of several key transcription factors [5]. 48 In addition to growth factors and cytokines, metabolic signals are 49 50considered as the third major signals during liver regeneration, which is however relatively less studied [6]. Recently, BAs were iden-51 tified as key metabolic signals during liver regeneration and their 5253roles in promoting liver regeneration have received more and more 54attention [7,8]. In this review, the recent advance of BA signaling in 55liver regeneration will be summarized and discussed.

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2. Metabolic signals and liver regeneration

Liver regeneration is an adaptive regrowth response induced by 57 specific stimuli and the subsequently sequential changes in gene ex-58 pression and morphologic reconstruction. It is generally accepted that 59 the remaining hepatocytes are the major cell types that replicate to re-60 generate liver in the models of 70% PHx or liver injury. Only in some 61 special injury models, activation and replication of liver progenitors 62 are observed when the hepatocytes fail to replicate normally. In addi-63 tion to hepatocytes, other cell types are also actively involved in liver re-64 generation or repair. Recently, several excellent reviews also highlight 65 the roles of liver stellate cells, liver sinusoidal endothelial cells and 66 liver stem/progenitor cells in liver regeneration and repair [9–12]. 67

Liver regeneration includes a highly complex network of signal 68 transductions. The essential circuitry required for this process is defined 69 mainly by three major networks: cytokine, growth factor and metabolic 70 signaling [13]. These three networks subsequently activate specific 71 genes and signaling pathways that are essential for liver regeneration. 72 Compared to the cytokine and growth factor networks, little is known 73 about the roles of metabolic signals in liver regeneration. The identification of several nuclear receptors as receptors for liver metabolites 75 provides a novel insight into the roles of metabolic signals in liver regeneration. Among them, the Farnesoid X Receptor (FXR, NR1H4) is identified as a primary BA receptor [14,15]. FXR belongs to a sub-cluster of 78 metabolic nuclear receptors that also includes Vitamin D Receptor 79 (VDR, NR111), Constitutive Androstane Receptor (CAR, NR113), 80 Pregnane X Receptor (PXR, NR1H2) and Liver X Receptor alpha and 81 beta (LXR α , NR1H3; LXR β , NR1H2). Peroxisome proliferator-activated 82

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receptors (PPARs) are also important metabolic nuclear receptors. All 83 these receptors bind to DNA either as a monomer or as a heterodimer 84 85 with a common partner for nuclear receptors, Retinoid X Receptor 86 (RXR, NR2B1) to regulate the expression of various genes involved in BA, lipid, glucose, and drug metabolism [16]. Interestingly, their roles 87 in liver regeneration are also under active investigation. For example, 88 upon PHx, liver regeneration is impaired in mice lacking RXR α in hepa-89 tocytes [17]. LXR may suppress liver regeneration after PHx through 90 91 regulating the cholesterol levels in the liver [18]. CAR activation strongly 92 induces hepatomegaly and may contribute to normal liver regeneration 93 after 70% PHx [7,19]. Dai et al. indicated that PXR is required for normal progression of liver regeneration by modulating lipid homeostasis and 94regulating hepatocyte proliferation [20]. In contrast, PPAR γ acts as a 9596 negative regulator of hepatocyte proliferation and may be responsible for the inhibition of liver growth in the late phase of liver regeneration 97 [21]. There is a detailed summary on nuclear receptors in liver regener-98 99 ation recently [22].

Liver is a major organ for metabolism. Therefore, there is an im-100 mense metabolic demand during liver regeneration. The requirement 101 of metabolic signals for liver regeneration has been known for a long 102time. However, their direct effect on liver regeneration is still unclear. 103 Among different metabolic signals, BAs are attractive signals for liver re-104 105 generation because the levels of BAs are tightly regulated. BAs are in-106 trinsically toxic and cause liver injury if the levels are not controlled properly. As such, liver resection or injury will generate a BA overload 107in the liver, which is a potential driving force for liver regeneration [7,8]. 108

1093. BA signaling and liver regeneration

BAs are liver-specific metabolites. They are end products from cho-110 lesterol catabolism and are important for nutrition absorption in the in-111 112 testine, which include cholesterol, lipids and fat-soluble vitamins. BAs are synthesized in the liver and stored in the gall bladder. They are se-113114 creted into the intestine when a meal is ingested, but 95% BAs are reabsorbed and transported back to the liver through the portal vein. 115This system is known as enterohepatic circulation. Hepatic BAs com-116 prise less than 5% of the total BA pool and PHx increases bile influx, 117 118 which rapidly generates a BA overload in the liver. Consistently with this, there is a sharp repression of Cyp7a1 gene expression after 70% 119 PHx or liver injury [23]. Cyp7a1 is the rate-limiting enzyme required 120for the BA production from cholesterol catabolism. The importance for 121 122 a stringent control of BA levels is illustrated by a delicate regulation of Cyp7a1 expression. There are many factors and pathways that can reg-123 ulate the expression of Cyp7a1 gene. A negative feedback loop is identi-124 125fied to regulate BA levels, in which high levels of BA activate FXR to increase the mRNA levels of SHP, which is a negative regulator of 126127Cyp7a1 gene expression. Moreover, additional regulators of Cyp7a1 expression are identified, including cytokines, growth factors and nuclear 128receptors [24-30]. During liver regeneration, in addition to FXR-SHP 129axis, hepatocyte growth factor and JNK pathways are also involved in 130suppressing Cyp7a1 expression during the acute phases of liver regen-131 132eration [23].

The strong suppression of BA synthesis during liver regeneration in-133dicates a BA overload stress in the liver. This also suggests that BAs may 134participate in the liver regeneration. Indeed, interruption of normal 135enterohepatic biliary circulation has been previously known to inhibit 136137 liver regeneration [31,32]. Moreover, there is also some direct evidence that BAs are able to stimulate hepatocyte proliferation [33–35]. In a 138 study of 70% PHx mouse model, supplementation with a low dose of 139BAs promotes liver regeneration, while reduction of BA levels by a BA-140 binding resin delays liver regeneration [7]. Defective BA signaling that 141 causes delayed liver regeneration is also demonstrated in other animal 142models. In the absence of MRP3, a BA transporter, liver regeneration is 143 delayed in mice due to lower BA concentration in the portal blood 144 [36]. Similarly, deletion of Cyp27, an enzyme required for normal BA 145146 production and metabolism, results in lower BA pool and defective liver regeneration in mice [37]. In FXR -/- mice, the effect of BAs on 04 promoting liver regeneration is lost [7]. Similarly, in MRP3 -/- and 148 Cyp27 - / - mice, the delayed liver regeneration is due to impaired 149 FXR activation, suggesting that FXR is the key player to mediate BA ef- 150 fect on liver regeneration. In conclusion, the identification of a novel 151 role of FXR in liver regeneration is a key to understand the molecular 152 mechanism by which BAs affect liver regeneration. 153

4. FXR and liver regeneration

FXR is highly expressed in the liver, intestine, and kidney where the 155 levels of BAs are relatively high [38]. FXR is the primary sensor of BAs 156 and both conjugated and unconjugated bile salts are able to activate 157 FXR at physiological concentrations [39,40]. FXR regulates BA homeo- 158 stasis by regulating genes involved in BA synthesis, secretion, transpor-159 tation, absorption, conjugation, and detoxification [41-45]. As expected, 160 FXR is also the BA receptor to mediate BA's effect on liver regeneration 161 [7]. FXR is shown to promote liver regeneration after 70% PHx and stim- 162 ulate liver repair after CCl₄-induced liver injury [46]. Interestingly, dif- 163 ferent from 70% PHx, there is massive cell death in liver injury model. 164 FXR is shown to have a dual role in promoting liver regeneration by 165 both stimulating hepatocyte proliferation and protecting the hepato- 166 cyte from death [46,47]. 167

In addition to metabolic genes, Foxm1b is identified as a FXR direct 168 target gene during liver regeneration [7,48]. Foxm1b is a key cell cycle 169 regulator essential for G1/S and G2/M progression [49,50]. Animal stud- 170 ies indicate that Foxm1b is a key transcription factor in liver regenera- 171 tion. Although liver can fully regenerate itself, aging dramatically 172 reduces this capacity of the liver. The delayed and reduced proliferative 173 response has been attributed to the decreased expression of some key 174 transcription factors, such as c-Myc and Foxm1b and to the failure of 175 aging-liver to diminish the age-specific C/EBP α -Brm-HDAC1 complex 176 after PHx [51–53]. The complex suppressed Foxm1b induction after 177 PHx through binding to Foxm1b promoter, which results in age- 178 related proliferation defects upon PHx or liver injury [54]. These studies 179 highlight Foxm1b as one of the key regulators in aging-liver regenera- 180 tion. Defective activation of FXR occurs in aged regenerating livers 181 [48], which may account for the insufficient Foxm1b induction. Com- 182 pared with young mice, aging mice did not have altered protein levels 183 of FXR and RXR. Therefore, aging may affect the levels of endogenous 184 FXR ligands such as BAs, which could result in defective activation of 185 FXR during liver regeneration. Interestingly, in pregnant mice, the loss 186 of FXR results in reduced liver growth, indicating a similar function of 187 FXR in mediating the liver growth during pregnancy [55]. 188

Sirtuin1 (SIRT1) that can modulate FXR activities also has an effect 189 on liver regeneration through modulating FXR activities during liver re- 190 generation [56]. The transgenic mice that overexpress SIRT1 showed in- 191 creased mortality, enhanced liver injury and impaired hepatocyte 192 proliferation after PHx. SIRT1 reduces FXR activities through persistent 193 deacetylation and lower FXR expression. In summary, FXR is a key re- 194 ceptor and transcription factor that specifically mediates the effect of 195 BA signaling to promote liver regeneration. 196

5. Intestine FXR and liver regeneration

FXR is highly expressed in both the liver and intestine. Both hepatic- 198 and intestine-FXR are involved in the regulation of BA homeostasis [57]. 199 One critical FXR target gene in the intestine is FGF15. Indeed, several re- 200 ports suggest that FGF15 secreted from the ileum has profound effects 201 on the suppression of Cyp7a1 gene expression and liver metabolism 202 through FGF receptor-mediated signaling pathways in the liver 203 [58–60]. Suppression of Cyp7a1 expression and decreased bile acid 204 synthesis are known to be beneficial for liver regeneration. Therefore, 205 FGF15 induction after liver damage may also contribute to the 206 normal liver regeneration. Indeed, there is a significantly delayed liver 207 regeneration and increased liver injury in intestine-specific FXR 208

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