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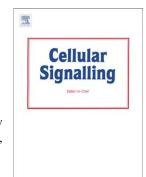
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Calmidazolium evokes high calcium fluctuations in *Plasmodium* falciparum

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ABSTRACT

Calcium and calmodulin (CaM) are important players in eukaryote cell signaling. In the present study, by using a knockin approach, we demonstrated the expression and localization of CaM in all erythrocytic stages of *Plasmodium falciparum*. Under extracellular Ca²⁺-free conditions, calmidazolium (CZ), a potent CaM inhibitor, promoted a transient cytosolic calcium ([Ca²⁺]_{cvt}) increase in isolated trophozoites, indicating that CZ mobilizes intracellular sources of calcium. In the same extracellular Ca²⁺-free conditions, the [Ca²⁺]_{cyt} rise elicited by CZ treatment was ~3.5 fold higher when the endoplasmic reticulum (ER) calcium store was previously depleted ruling out the mobilization of calcium from the ER by CZ. The effects of the Ca²⁺/H⁺ ionophore ionomycin (ION) and the Na⁺/H⁺ ionophore monensin (MON) suggest that the [Ca²⁺]_{cvt}-increasing effect of CZ is driven by the removal of Ca²⁺ from at least one Ca²⁺-CaM-related (CaMR) protein as well as by mobilization of Ca²⁺ from intracellular acidic calcium stores. Moreover, we showed that the mitochondrion participates in the sequestration of the cytosolic Ca²⁺ elicited by CZ. Finally, the modulation of membrane Ca²⁺ channels by CZ and thapsigargin (THG) was demonstrated. The opened channels were blocked by the unspecific calcium channel blocker Co²⁺ but not by 2-APB (capacitative calcium entry inhibitor) or nifedipine (L-type Ca²⁺ channel inhibitor). Taken together, the results suggested that one CaMR protein is an important modulator of calcium signaling and homeostasis during the *Plasmodium* intraerythrocytic cell cycle, working as a relevant intracellular Ca²⁺ reservoir in the parasite.

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