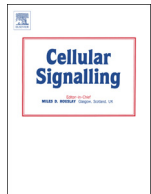




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New advances of DNA methylation in liver fibrosis, with special emphasis on the crosstalk between microRNAs and DNA methylation machinery[☆]

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ABSTRACT

Epigenetics refers to the study of heritable changes in the pattern of gene expression that is controlled by a mechanism specifically not due to changes the primary DNA sequence. Well-known epigenetic mechanisms include DNA methylation, post-translational histone modifications and RNA-based mechanisms including those controlled by small non-coding RNAs (miRNAs). Recent studies have shown that epigenetic modifications orchestrate the hepatic stellate cell (HSC) activation and liver fibrosis. In this review we focus on the aberrant methylation of CpG island promoters of select genes is the prominent epigenetic mechanism to effectively silence gene transcription facilitating HSC activation and liver fibrosis. Furthermore, we also discuss epigenetic dysregulation of tumor-suppressor miRNA genes by promoter DNA methylation and the interaction of DNA methylation with miRNAs involved in the regulation of HSC activation and liver fibrosis. Recent advances in epigenetics alterations in the pathogenesis of liver fibrosis and their possible use as new therapeutic targets and biomarkers.

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Abbreviations: miRNAs, small non-coding RNAs; HSC, hepatic stellate cell; ECM, extracellular matrix; α -SMA, α -smooth muscle actin; TGF- β , transforming growth factor- β ; PDGF, platelet-derived growth factor; FGF, fibroblast growth factor; ICAM-1, intercellular adhesion molecule-1; MCP-1, monocyte chemoattractant protein-1; DNMTs, DNA methyltransferases; MBD, methyl-CpG-binding domain; 5-azadC, 5-aza-2'-deoxycytidine; PTEN, Phosphatase and tensin homologue; PI3K, Phosphatidylinositol-3-kinase; RASAL1, Ras GTPase activating-like protein 1; PTCH1, patched1; Gli1, glioma-associated oncogene homolog 1; PPAR γ , Transcription factor peroxisome proliferator-activated receptor- γ ; NF κ B, Nuclear factor κ B; TET, Ten-Eleven- Translocation; TDG, thymine DNA glycosylase; BER, base excision repair; pri-miRNAs, primary miRNA transcripts; miRNPs, miRNA-protein complexes; UTR, untranslated region; RBPs, RNA-binding proteins; TS-miRNA, tumor-suppressive miRNA; TSG, tumor-suppressor gene; GSTP1, glutathione S-transferase pi 1; CDH1, E-cadherin 1; BDNF, brain-derived neurotrophic factor; IUGR, Intrauterine growth restriction.

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56 1. Introduction

57 Liver fibrosis results from persistent liver injury, including viral
58 hepatitis, alcohol abuse, metabolic diseases, autoimmune diseases,
59 and cholestatic liver diseases [1]. During fibrosis progression, inflam-
60 mation and liver injury trigger complex cellular events that result in
61 collagen deposition and the disruption of the normal liver architec-
62 ture [2]. Over the last two decades, sinusoidal resident hepatic stellate
63 cells (HSCs) have been commonly recognized as the major source of
64 extracellular matrix (ECM). In the normal liver, HSCs are quiescent,
65 vitamin A-storing adipogenic cells. However, following a fibrogenic
66 stimulus, HSCs undergo a complex activation process associated
67 with morphological changes from a quiescent vitamin A-storing cell
68 to that of an activated myofibroblast-like cell [3,4]. HSC activation is
69 also associated with a dramatic increase in the synthesis and deposi-
70 tion of ECM components, marked upregulation of α -smooth muscle
71 actin (α -SMA), collagen, tissue inhibitors of metalloproteinases
72 (TIMP1) and desmin, production of profibrogenic cytokines/growth
73 factors such as transforming growth factor- β (TGF- β), platelet-derived
74 growth factor (PDGF) and fibroblast growth factor (FGF), as well as
75 pro-inflammatory molecules including interleukin (IL)-6, intercellular
76 adhesion molecule-1 (ICAM-1) and monocyte chemoattractant protein-1
77 (MCP-1) [5–7].

78 Because HSC activation and liver fibrosis are orchestrated by the
79 same signals, for example by growth factors such as TGF- β , the
80 molecular mechanisms which exert global control of HSC activation
81 and liver fibrosis incompletely understood. Recent works from our
82 group and from others implicated that epigenetic modifications play
83 an important role in determining HSC activation and liver fibrosis
84 (Fig. 1). Here we review insights into the role of epigenetics in HSC
85 activation and liver fibrosis.

86 2. The pathogenesis of liver fibrosis

87 Liver fibrosis, irrespective of aetiology, is a dynamic and highly
88 integrated molecular, tissue and cellular process that leads to pro-
89 gressive accumulation of ECM components in an attempt to limit
90 hepatic damage in chronic liver diseases [8]. The terminal outcome
91 of liver fibrosis is liver cirrhosis, a condition characterized by distor-
92 tion of the normal architecture, septae and nodule formation, altered
93 blood flow, portal hypertension, hepatocellular carcinoma and ulti-
94 mately liver failure [9]. The hepatic stellate cell (HSC) is the main
95 fibrogenic cell type orchestrating the deposition of ECM in the injured
96 liver and it also has been identified as a primary effector in liver
97 inflammation [4].

98 HSCs are resident perisinusoidal cells in the subendothelial space
99 between hepatocytes and sinusoidal endothelial cells [10]. These cells
100 are strategically positioned to intimately interact with hepatocytes,
101 endothelial cells, and nerve endings through their numerous processes
102 extending across the space of Disse [11]. Under pathological conditions,
103 including injury, inflammation, hepatitis B virus (HBV) or hepatitis C
104 virus (HCV) infection, quiescent HSC have been reported to undergo a
105 particular process of activation which involves significant changes in
106 morphology and phenotypical responses observed in either human or
107 rat HSC when cultured on plastic substrate [12–14].

108 Several factors have been identified to promote HSC activation.
109 Damage to hepatocytes and Kupffer cell activation are still considered
110 the main effectors driving HSC activation [15,16]. Mediators released
111 from damaged hepatocytes, such as lipid peroxidation products,

intermediate metabolites of drugs or hepatotoxins, acetaldehyde 112
and 1-hydroxyethyl radical from alcohol metabolism as well as reactive 113
oxygen species (hydrogen peroxide, superoxide radical and others are 114
strong inducers of HSC activation [17]. Once activated by bacterial 115
products, Kupffer cells secrete a large number of pro-inflammatory 116
and fibrogenic mediators. Activation of HSC by macrophage-derived 117
TGF- β or insulin-like growth factor is an early feature of fibrogenesis 118
which promotes a switch in HSC gene expression to initiate matrix 119
remodeling [18]. 120

Advances of understanding gene regulation in HSCs has paralleled 121
the dramatic expansion of knowledge about both traditional mecha- 122
nisms of gene regulation, including transcription factor activity, local- 123
ization and modification, as well as epigenetic regulation of gene 124
expression by DNA methylation, histone modification and microRNA 125
interactions [19–24]. Elucidating the precise molecular mechanisms 126
underlying HSC activation and liver fibrosis is translating into fruitful 127
new therapeutic approaches. 128

3. Overview of DNA methylation 129

The methylation of the C5 position of the cytosine base with 130
S-adenosyl methionine as the methyl donor is found in approximately 131
70–80% of CpG dinucleotides in somatic mammalian cells and to some 132
extent in non-CpG sequences in embryonic stem cells [25,26]. DNA 133
methylation is currently the most widely studied form of epigenetic 134
programming. The methylation of cytosine residues within CpG 135
sequences is catalysed by DNA methyltransferases (DNMTs) [27]. In 136
mammals, five members of the DNMT family have been identified: 137
DNMT1, DNMT2, DNMT3A, DNMT3B, and DNMT3L. Among these pro- 138
teins, only DNMT1, DNMT3A, and DNMT3B exhibit methyltransferase 139
activity. DNMT3a and DNMT3b target unmethylated CpGs and therefore 140
are termed de novo methyltransferases, while DNMT1 maintains DNA 141
methylation during replication by copying the methylation pattern of 142
the parent DNA strand onto the newly synthesized strand [28,29]. 143

DNA methylation of the promoter regions is generally related to 144
transcriptional repression through different mechanisms, including 145
the inhibition of transcription factor binding and the recruitment 146
of methyl-CpG-binding domain (MBD) proteins and their associated 147
complexes [30]. So far, six methyl-CpG-binding proteins, including 148
MeCP2, MBD1, MBD2, MBD3, MBD4 and Kaiso, have been reported in 149
mammals [31]. MeCP2 is a member of a small family of methylated 150
DNA-binding domain proteins that was first described through its affin- 151
ity for DNA sequences containing methylated 5'-CpG-3' dinucleotides 152
[32]. The ability of MeCP2 to bind methylated DNA has been interpreted 153
in the context of transcriptional repression and silencing of specific 154
target genes. In addition, MeCP2 binds the corepressor mSin3A, which 155
is thought to recruit histone deacetylases, providing a mechanism for 156
the transcriptional repression of genes with methylated CpG sites 157
[33]. Interestingly, MeCP2 was shown to associate with the transcrip- 158
tional activator CREB1 at the promoter of somatostatin, a gene 159
upregulated in Mecp2 duplication mice, thereby suggesting a potential 160
activation mechanism [34]. 161

4. Methylated genes in liver fibrosis 162

Abnormal patterns of DNA methylation in liver fibrosis have been 163
recognized over the last few years and so far a number of aberrantly 164
hypermethylated genes have been discovered. These genes have 165
been found to be hypermethylated either by direct examination of 166

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