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# <sup>3</sup> Digital pathology and image analysis in tissue biomarker research

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## ABSTRACT

Digital pathology and the adoption of image analysis have grown rapidly in the last few years. This is largely due to the implementation of whole slide scanning, advances in software and computer processing capacity and the increasing importance of tissue-based research for biomarker discovery and stratified medicine. This review sets out the key application areas for digital pathology and image analysis, with a particular focus on research and biomarker discovery. A variety of image analysis applications are reviewed including nuclear morphometry and tissue architecture analysis, but with emphasis on immunohistochemistry and fluorescence analysis of tissue biomarkers. Digital pathology and image analysis have important roles across the drug/companion diagnostic development pipeline including biobanking, molecular pathology, tissue microarray analysis, molecular profiling of tissue and these important developments are reviewed. Underpinning all of these important developments is the need for high quality tissue samples and the impact of pre-analytical variables on tissue research is discussed. This requirement is combined with practical advice on setting up and running a digital pathology laboratory. Finally, we discuss the need to integrate digital image analysis data with epidemiological, clinical and genomic data in order to fully understand the relationship between genotype and phenotype and to drive discovery and the delivery of personalized medicine.

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## 44 1. Introduction

While it is a common misconception that digital pathology and 45 image analysis is new, research on the use of computers and soft-46 47 ware for analyzing and measuring cells or tissues in pathology date as far back as the 1960's and 70's [1-4]. That's over 40 years ago! 48 Clearly, the hardware and software systems then were limited in 49 their capability by comparison to today - but those studies were 50 the first to demonstrate the value that computer-based imaging, 51 52 cellular measurement and quantitation could play in pathological 53 diagnosis and discovery.

54 As computer hardware advanced rapidly in the 1980's and 1990's, there was considerable promise that image analysis would 55 be embraced as part of routine diagnosis in pathology. Some even 56 57 posited that the technology would ultimately replace human pathologists. There was enormous investment in automated cytol-58 59 ogy screening based on IA, with the promise that this could be used 60 to reduce cytology workload and improve diagnostic performance 61 across laboratories worldwide. Clearly this did not happen on the

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http://dx.doi.org/10.1016/j.ymeth.2014.06.015 1046-2023/© 2014 Elsevier Inc. All rights reserved. scale predicted and even the most state of the art IA systems failed to significantly change practice in pathology. So the initial enthusiasm for digital IA technology in pathology waivered with the focus shifting to molecular pathology and the promise of diagnostic classification of tissue samples without the need for morphology. Three principle factors changed that: (1) the recognition that molecular pathology still relies heavily on tissue interpretation (2) the drive for targeted therapies based on the presence or absence of tissue-based markers and (3) digital scanning and whole slide imaging (WSI) of entire glass slides in pathology.

The last factor has been hugely instrumental in the recent upsurge in the adoption of image analysis again in both the research and diagnostic sector. Whole slide imaging (WSI), and associated viewing software, allows entire slides to be digitally scanned at high resolution, reviewed by an experienced morphologist, regions selected and image analysis applied to measure specific features. This potentially circumvents the need to use traditional microscopy, manual selection, restricted image capture using a CCD camera, transfer to an image analysis package and subsequent measurement of specific features. WSI can bring these processes together, making image analysis much more practicable and easy to adopt, while facilitating integration into existing workflows in both research and primary diagnostic laboratories.

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85 Industrial image analysis systems have grown dramatically in 86 recent years. This is likely to continue as the applications in discov-87 ery, preclinical and clinical research continue to demand quantita-88 tive methods, and as new diagnostic tools are translated into 89 diagnostic practice.

This article aims to provide readers with a rapid overview of the 90 91 current status of digital pathology and image analysis in biomarker 92 research and diagnostic practice, including practical advice on 93 adopting and developing these technologies.

#### 2. Whole slide scanning and digital slides 94

#### 2.1. Whole slide scanners 95

96 While the digital capture of individual images is still utilized 97 widely in the research and tissue diagnostic community, whole 98 slide scanning (WSI) is by far the most rapidly expanding means 99 of digital image capture in pathology. WSI allows the digital 100 capture of the entire tissue sample at high resolution and with 101 appropriate software allows the viewing of the slide at any position 102 and at any magnification. In this way it replicates what is achiev-103 able with standard microscopy, but provides a range of additional 104 advantages - including facilitating image analysis.

Over recent years WSI instrumentation has become more widely accepted and affordable in pathology research laboratories and in primary diagnostic laboratories. However, given the pace of development, there are likely to be further systems available from new providers as the market continues to expand.

110 Most of the systems rely on two variants of image capture (1) 111 line scanning and (2) tile scanning, both of which generate multi-112 ple high resolution images (in the form of lines or tiles) that are 113 subsequently aligned or stitched together to create a complete, composite image of the original whole tissue section. Collecting 114 115 image data by either method is achieved by passing the slide 116 underneath the objective using a carefully controlled motorized 117 scanning stage or objective assembly. The image data is rapidly 118 recorded as the slide is traversed and image data stitched together 119 in real time.

120 In most systems the magnification at which the slide is scanned 121 can be adjusted. This is commonly either at  $20 \times$  or  $40 \times$  magnifica-122 tion. Other select systems can scan under oil at  $63 \times$  to provide 123 higher resolution systems. 20× scans are sufficient for most standard H&E remote viewing applications although some institutions 124 125 prefer to scan at  $40 \times$  to ensure higher resolution. Fig. 1A shows a whole slide scan of a pancreatic cancer, scanned at 40× magnifica-126 127 tion where the image can be viewed at any magnification (Fig. 1B) 128 and where multiple slides can be viewed side by side for comparison 129 at any location or any magnification (Fig. 1C). Image analysis can 130 benefit from high resolution scans, particularly for applications that 131 involve nuclear detection and analysis. Applications such as in situ 132 hybridization (ISH) can be carried out at  $40 \times$  with fluorescence but may benefit from high magnification scans in order to resolve indi-133 vidual spots with chromogenic ISH. Haematology applications may 134 135 require 63× scanning (restricted to certain models of scanner) in 136 order to better resolve morphology and cell types. There is however 137 a storage premium to pay for high resolution scans.

Accurate focus across large areas of tissue during the scanning 138 process is essential. In most instrumentation, this is achieved by 139 mapping the topography variations that inherently exist across 140 141 even a very thin tissue section, and rapidly adjusting the focus as 142 the scan is being created [5]. The reliability of this process has 143 improved dramatically over recently years and most systems can 144 automatically scan large batches of slides with no human interven-145 tion at all.

Some WSI systems can also generate "multiplane" scans, which capture image data along the z-axis (Fig. 2) as numerous large

images in a stack. With appropriate viewing software, this provides the ability navigate images in the z-plane, creating a digital focus effect. This is particularly effective for cytology preparations, where the ability to focus is extremely important.

Finally, many scanning systems now offer fluorescence WSI. 152 This makes use of the benefits of fluorescence (see Section 4.6) 153 while providing full slide scans, digitally capturing all relevant data 154 for storage, remote review and image analysis. There are specific 155 challenges associated with fluorescence WSI, not least of which is 156 focus. Fluorescence images tend to contain less contextual back-157 ground information than brightfield images, and so provide less 158 data to support automated focus over large areas. However most 159 systems provide the ability to select defined regions of interest 160 for scanning, allowing large areas of slides to be successfully 161 scanned under fluorescence. 162

## 2.2. Image size and compression

Whole slide digital images are large. Scanning a typical tissue section of  $15 \times 20$  mm in size at  $20 \times$  viewing magnification  $(0.24 \,\mu\text{m per pixel})$  can generate images as large as 3.6 GB in size. Scanning at  $40 \times$  will generate images as large as 14.5 GB. These can be compressed to more manageable sizes (approx. 25:1 compression or more), reducing the file size without impacting on the visual quality of the image. Studies on the compression of images in digital pathology [6] have shown that extensive image compression can be applied without experts being able to visually perceive differences in image quality. Even images with high compression ratios can still be interpretable visually.

An important consideration, however, is how image compression can affect quantitative image analysis. Commercial systems 176 routinely apply different compression methods and levels as part 177 of their standard configuration and so variation from one instrument to the next could be detrimental. Basic studies have shown that densitometric measurement (which is used routinely for quantitative IHC image analysis) is more sensitive to compression that morphological measurement (e.g., nuclear size). Different compression methods offered by different vendors can have very 183 different effects on image analysis fidelity [7]. Kieran et al. [7] 184 showed that with some methods of compression, 5% of the nuclei 185 were segmented in error, with an error rate that steadily increased 186 as compressed image quality decreased. Care therefore needs to be 187 taken to assess the impact of compression artifacts on image anal-188 ysis, and the impact of compression needs to be validated for each 189 study depending on the features calculated. 190

## 2.3. Scanning speeds and automation

Most instruments can now scan slides in 1–3 min, some with the capability of automatically loading multiple slides without user intervention. Some of the larger scanning devices can accommodate in excess of 300 slides, making them ideal for high volume applications, including in busy clinical diagnostic laboratories or large scale tissue research facilities where large numbers of slides need to be scanned and archived daily. Smaller scanners are available, which can scan from 1 to 10 slides in a single action. These are ideal for specialist or incidental research requirements, for educational organizations that are scanning relatively small teaching collections, or for diagnostic labs that want to use digital pathology for infrequent second opinion or frozen section review.

## 2.4. Storage of digital slides

Given the size of digital slides and the numbers that are now 205 being routinely scanned in many research and diagnostic laboratories, storage represents a significant element of the investment

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