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## Effects of intraluteal implants of prostaglandin E1 or E2 on angiogenic growth factors in luteal tissue of Angus and Brahman cows



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### ABSTRACT

Previously, it was reported that intraluteal implants containing prostaglandin E<sub>1</sub> or E<sub>2</sub> (PGE<sub>1</sub> and PGE<sub>2</sub>) in Angus or Brahman cows prevented luteolysis by preventing loss of mRNA expression for luteal LH receptors and luteal unoccupied and occupied LH receptors. In addition, intraluteal implants containing PGE<sub>1</sub> or PGE<sub>2</sub> upregulated mRNA expression for FP prostanoid receptors and downregulated mRNA expression for EP<sub>2</sub> and EP<sub>4</sub> prostanoid receptors. Luteal weight during the estrous cycle of Brahman cows was reported to be lesser than that of Angus cows but not during pregnancy. The objective of this experiment was to determine whether intraluteal implants containing PGE<sub>1</sub> or PGE<sub>2</sub> alter vascular endothelial growth factor (VEGF), fibroblast growth factor-2 (FGF-2), angiopoietin-1 (ANG-1), and angiopoietin-2 (ANG-2) protein in Brahman or Angus cows. On Day 13 of the estrous cycle, Angus cows received no intraluteal implant and corpora lutea were retrieved, or Angus and Brahman cows received intraluteal silastic implants containing vehicle, PGE<sub>1</sub>, or PGE<sub>2</sub> on Day 13 and corpora lutea were retrieved on Day 19. Corpora lutea slices were analyzed for VEGF, FGF-2, ANG-1, and ANG-2 angiogenic proteins via Western blot. Day-13 Angus cow luteal tissue served as preluteolytic controls. Data for VEGF were not affected ( $P > 0.05$ ) by day, breed, or treatment. PGE<sub>1</sub> or PGE<sub>2</sub> increased ( $P < 0.05$ ) FGF-2 in luteal tissue of Angus cows compared with Day-13 and Day-19 Angus controls but decreased ( $P < 0.05$ ) FGF-2 in luteal tissue of Brahman cows when compared with Day-13 or Day-19 Angus controls. There was no effect ( $P > 0.05$ ) of PGE<sub>1</sub> or PGE<sub>2</sub> on ANG-1 in Angus luteal tissue when compared with Day-13 or Day-19 controls, but ANG-1 was decreased ( $P < 0.05$ ) by PGE<sub>1</sub> or PGE<sub>2</sub> in Brahman cows when compared with Day-19 Brahman controls. ANG-2 was increased ( $P < 0.05$ ) on Day 19 in Angus Vehicle controls when compared with Day-13 Angus controls, which was prevented ( $P < 0.05$ ) by PGE<sub>1</sub> but not by PGE<sub>2</sub> in Angus cows. There was no effect ( $P > 0.05$ ) of PGE<sub>1</sub>

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or PGE<sub>2</sub> on ANG-2 in Brahman cows. PGE<sub>1</sub> or PGE<sub>2</sub> may alter cow luteal FGF-2, ANG-1, or ANG-2 but not VEGF to prevent luteolysis; however, species or breed differences may exist.

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## 1. Introduction

The corpus luteum is the source of progesterone during the estrous cycle [1,2]. Luteal weights are similar from the middle of the estrous cycle and throughout pregnancy in *Bos taurus* cows [3,4]. Concentrations of circulating progesterone in *Bos taurus* cows increase twofold from Day 12 to Day 18 after breeding and do not change from Day 20 to Day 260 of pregnancy [4]. However, luteal weights are lighter in *Bos indicus* cows on Day 15 of the estrous cycle than in *Bos taurus* cows, but luteal weights are similar during mid to late pregnancy in *Bos indicus* and *Bos taurus* cows. Luteal secretion of progesterone and prostaglandin E per unit mass *in vitro* on Day 15 of the estrous cycle and Day 200 of pregnancy are similar in *Bos indicus* and *Bos taurus* cows [5,6,9]. The placentas of cows do not secrete progesterone when the corpus luteum is functional, which is important to maintain luteal function throughout pregnancy [10].

Loss of progesterone secretion at the end of the estrous cycle in ewes or cows is *via* uterine PGF<sub>2 $\alpha$</sub>  secretion [1,2]. In a single sample of uterine venous blood of cows on Day 18 of pregnancy, PGF<sub>2 $\alpha$</sub>  is decreased but not in ovarian arterial or venous blood or in luteal tissue [7]. In the ewe, PGF<sub>2 $\alpha$</sub>  in the endometrium, uterine or ovarian venous blood, luteal tissue, binding of PGF<sub>2 $\alpha$</sub>  to luteal membranes, or transport of PGF<sub>2 $\alpha$</sub>  from the uterine vein to the adjacent ovarian artery of the luteal-containing ovary are not decreased to explain prevention of luteolysis during early pregnancy [1,2,8]. The embryo–endometrial interaction provides resistance to PGF<sub>2 $\alpha$</sub> -induced luteolysis during early pregnancy [2]. This resistance to PGF<sub>2 $\alpha$</sub>  is due to increases in endometrial PGE<sub>1</sub> and PGE<sub>2</sub> and PGE (PGE<sub>1</sub> + PGE<sub>2</sub>) during early pregnancy [1,2].

In cows, *Bos taurus* and *Bos indicus* luteal tissue during the estrous cycle or pregnancy secretes much PGE<sub>1</sub> or PGE<sub>2</sub> *in vitro*, which increases with time in culture [6,9]. PGE<sub>1</sub> or PGE<sub>2</sub> stimulates bovine luteal progesterone secretion *via* cAMP *in vitro* [11]. In cows, PGE<sub>1</sub> given intramuscularly increased circulating progesterone for the duration of the 72-hour sampling period [12]. In Angus and Brahman cows, intraluteal silastic implants containing PGE<sub>1</sub> or PGE<sub>2</sub> on Day 13 to Day 19 prevented the decline in luteal weight, circulating progesterone, luteal mRNA for LH receptors, and unoccupied and occupied receptors for LH in Angus or Brahman cows [13]. However, the FP prostanoid receptor was decreased on Day 19 in both Angus and Brahman vehicle controls, whereas PGE<sub>1</sub> or PGE<sub>2</sub> on Day 19 in these same cows increased mRNA for the prostanoid FP and EP3B receptors and decreased mRNA for prostanoid EP2 and EP4 receptors [14]. An EP3 prostanoid receptor agonist *in vivo* is as effective as PGF<sub>2 $\alpha$</sub>  in causing luteolysis [15]. Data in ewes also support PGE<sub>1</sub> or PGE<sub>2</sub> as antiluteolysins to prevent luteolysis. Concentrations of PGE (PGE<sub>1</sub> + PGE<sub>2</sub>) are increased in uterine venous blood during early pregnancy [1,2]. Chronic intra-uterine infusions of PGE<sub>1</sub> or PGE<sub>2</sub> adjacent to the corpus

luteum-containing ovary prevent a spontaneous or induced luteolysis [1,2]. The common antiluteolytic mechanism in the cow or ewe is that both PGE<sub>1</sub> or PGE<sub>2</sub> prevent loss of luteal mRNA for LH receptors and prevent loss of luteal unoccupied and occupied receptors for LH [13,16].

The angiogenic factors VEGF, FGF-2, ANG-1, and ANG-2 have been associated with growth or demise of luteal tissue potentially through alterations in vascularity [17–22]. VEGF was higher in developing corpora lutea than at mid-or-late luteal stages of the estrous cycle; FGF-2 was elevated throughout the estrous cycle and has been reported to be involved in both maintenance and demise of the corpus luteum, whereas ANG-1 has been reported to facilitate luteal vascular growth and ANG-2 was reported to increase in luteal tissue during luteal regression [17–22]. Studies on changes in luteal angiogenic growth factors have used primarily PGF<sub>2 $\alpha$</sub> -induced luteolysis. The decline in progesterone in jugular venous blood of cows decreases to less than 1 ng/mL within 4 hours after lutectomy, which is similar to an exogenous dose of PGF<sub>2 $\alpha$</sub> -induced luteolysis at midcycle [23]. This indicates that PGF<sub>2 $\alpha$</sub> -induced luteolysis is a pharmacologic effect, because progesterone decreases slowly over 3 to 4 days during a natural luteolysis [24]. Furthermore, synchronization of estrus in cows alters the response to agonists by bovine luteal or endometrial tissue *in vitro* [25,26].

The objective of this experiment was to determine whether intraluteal implants containing PGE<sub>1</sub> or PGE<sub>2</sub> affect the luteal angiogenic factors VEGF, FGF-2, ANG-1, and ANG-2 in cows when estrus is not synchronized.

## 2. Materials and methods

### 2.1. Cows and surgery

This experiment was approved by Texas A&M University Animal Care and Use Committee (TAMU AUC 2008–129). Cows were checked twice daily (05:00 and 17:00 hours) for estrus during September to November. Sixteen non-lactating multiparous Angus cows and 12 nonlactating multiparous Brahman cows aged from 5 to 8 years were maintained on pasture and feed, and water was withdrawn 12 hours before surgery. Before surgery, the flank of the cow was clipped, washed with soap and water, scrubbed with Betadine (Purdue Frederick, Stamford, CT, USA), and Lidocaine (Hospira Inc., Lake Forest, IL, USA) was administered subcutaneously into the flank as a local anesthesia. Ten milliliters of penicillin was given around the incision and intramuscularly (Bimeda Inc., Le Sueur, MN) after surgery [13,14].

### 2.2. Intraluteal implants and collection of samples

One-half of Dow 382 silastic elastomer (Dow Corning Chemical Co, Midland, MI) was mixed in a sterile tube with

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