



Mini-review

Potential molecular, cellular and microenvironmental mechanism of sorafenib resistance in hepatocellular carcinoma

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ARTICLE INFO

Article history:

Received 4 March 2015

Received in revised form 23 June 2015

Accepted 25 June 2015

Keywords:

Advanced hepatocellular carcinoma (HCC)

Epithelial–mesenchymal transitions (EMT)

and mesenchymal–epithelial transitions

(MET)

Cancer stem cells (CSCs)

Microenvironment

Acquired resistance

ABSTRACT

Sorafenib, an orally-available kinase inhibitor, is the only standard clinical treatment against advanced hepatocellular carcinoma. However, development of resistance to sorafenib has raised concern in recent years due to the high-level heterogeneity of individual response to sorafenib treatment. The resistance mechanism underlying the impaired sensitivity to sorafenib is still elusive though some researchers have made great efforts. Here, we provide a systemic insight into the potential molecular, cellular and microenvironmental mechanism of sorafenib resistance in hepatocellular carcinoma depending on abundant previous studies and reports.

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Introduction

Hepatocellular carcinoma (HCC) is one of the most common primary malignant tumors and the largest cancer-related deaths ranking only second to lung cancer, with a fairly high and increasing incidence, frequently relapse and dismal prognosis [1,2]. Despite the remarkable progress in the prevention, detection, and treatment of cancer over the last five decades, no adequate therapy remains sufficiently effective due to late stage diagnosis and inadequate clinical strategies for inhibiting metastasis and promoting apoptosis [3–6]. Furthermore, multi-drug resistance of cancer cells has been implicated as a major challenge considering the irreplaceable role of chemotherapeutic interventions in anti-cancer treatment [7]. The resistance was postulated to associate with elevated expression of drug efflux transporters, changes in drug kinetics, amplification of drug targets or tumor heterogeneity comprising of genetic variation, the microenvironment, and cell plasticity [8]. For patients with HCC diagnosed at advanced stage, sorafenib is the only choice of systemic therapy when potentially curative treatment, such as resection and liver transplantation, may be merely applicable for patients diagnosed at early stage [9]. Recently, the unstable efficacy of sorafenib has raised concern of more and more researchers

and ‘sorafenib resistance’ has become a hot term used to describe the impaired efficacy of sorafenib, especially for patients with advanced HCC.

Sorafenib, as a multikinase inhibitor, suppresses tumor angiogenesis and proliferation by inhibiting serine/threonine kinases, as well as receptor tyrosine kinases. Intracellular Raf serine/threonine kinase isoforms inhibited by sorafenib include Raf-1 (or C-Raf), wild-type B-Raf and mutant B-Raf. Receptor tyrosine kinases inhibited by sorafenib include vascular endothelial growth factor receptor (VEGFR)-1, VEGFR-2, VEGFR-3, platelet-derived growth factor receptor (PDGFR)-b, c-KIT, FMS-like tyrosine kinase 3 (FLT-3) and RET [10]. Among these kinases, RAF and VEGFR are presumed to be essential for the anti-proliferative effects evoked by sorafenib. RAF kinases are present at the level of the cancer cells, while the VEGFR is present on the surface of endothelial cells [11]. Inhibition of the VEGFR accounts, at least in part, for the antiangiogenic effects of sorafenib. In majority of patients with advanced HCC, the RAF–MEK–ERK cascade is often activated by autocrine and paracrine loops, involving for example the production of amphiregulin, an agonist of the Epidermal Growth Factor Receptor (EGFR) [12]. The analysis of vitro experiments reported that there was a strong correlation between the inhibition of the RAF–MEK–ERK cascade and the anti-clonogenesis effect of sorafenib [13]. In addition, cytotoxic effect of sorafenib is also a crucial role in antitumor treatment. Generally, apoptosis is the major form of cytotoxicity and it is required for tumor regression and sustained clinical remissions [14]. The pro-apoptotic effect of sorafenib in HCC cells had also been studied extensively. In 2008, the SHARP (Sorafenib HCC Assessment Randomised Protocol)

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1 trials showed an improved overall survival in Child–Pugh class A
2 patients with advanced HCC upon treatment with the antiangiogenic
3 and antiproliferative agent sorafenib. In addition, Oriental con-
4 firmed the efficacy and safety of sorafenib again in treatment of
5 advanced hepatocellular carcinoma by another multicenter, ran-
6 domized, double-blind, placebo-controlled phase III clinical trials.

7 However, the promising systemic chemotherapeutic agent
8 sorafenib only demonstrated relatively limited benefits rather than
9 eradicated the microscopic residual and cured the patients with ad-
10 vanced HCC. Sorafenib is beneficial in only around 30% of patients,
11 and acquired resistance often develops within 6 months [15,16], sug-
12 gesting the existence of primary and acquired sorafenib resistance
13 in hepatocellular carcinoma cells. So far, some researchers have in-
14 vestigated the mechanism underlying resistance to sorafenib from
15 molecular and clinical points of view. Huang et al. have reported
16 that overexpression of both α B-Crystallin and 14-3-3 ζ decreased
17 hepatoma cells sensitization to sorafenib in clinical by inducing
18 epithelial–mesenchymal transition (EMT) in HCC cells [17]. Chiou
19 et al. presented evidence that glucose–regulated protein 78 (GRP78)
20 is a positive modifier for sorafenib resistance acquisition in HCC and
21 GRP78 knockdown enhanced the efficacy of sorafenib-mediated cell
22 death [18]. Chen et al. also conducted experiments to investigate
23 sorafenib resistance and the results showed that long-term expo-
24 sure to sorafenib activated the phosphatidylinositol 3-kinase
25 (PI3K)/Akt signaling pathway and mediate acquired resistance to
26 sorafenib in HCC cell lines [19]. All the insights into the molecu-
27 lar and clinical changes involved in sorafenib treatment provided a
28 greater understanding of the underlying mechanism of primary and
29 acquired resistance to sorafenib. However, a systemic and compre-
30 hensive analysis about the mechanism of acquiring resistance to
31 sorafenib was still elusive [20]. Here, in order to clarify the drug re-
32 sistance mechanism clearly, we reviewed studies on sorafenib
33 resistance in liver cancer from different angles, including molecu-
34 lar markers, signaling pathways, drug resistance principles, regulation
35 systems, therapeutic implications and recent approaches. Increas-
36 ing evidence suggested that deeper insight into the formation of
37 sorafenib resistance would shed light on the specific drug resis-
38 tance mechanism and might lead to identification of potential clinical
39 biomarkers for prognosis evaluation and targets for new therapeu-
40 tic strategies. Therefore, in the present review, by synthesizing the
41 retrospective studies and reports, we provided potential molecu-
42 lar, cellular and microenvironmental mechanism underlying impaired
43 sensitivity to sorafenib in hepatocellular carcinoma, which had pro-
44 found effect on inhibiting tumor progression, evaluating patient
45 survival and predicting sorafenib treatment response.

47 Epithelial–mesenchymal transitions (EMT)

48 Epithelial–mesenchymal transitions (EMT) are mainly marked
49 by the loss of cell–cell interactions and of epithelial apico-basal po-
50 larity with the concomitant acquisition of mesenchymal markers
51 and enhanced migratory behavior [21,22]. As is well known, EMT
52 is of paramount relevance for various developmental processes, in-
53 cluding gastrulation, neural crest formation and carcinoma
54 progression. In the last decades, EMT has been extensively studied
55 and validated in the progression of various carcinomas such as HCC
56 [23,24]. A large body of researches has claimed the important role
57 of EMT in facilitating tumor development and progression by driving
58 metastasis, through the acquisition of enhanced migratory and in-
59 vasive potential [25–30]. Metastasis, induced by EMT, deconvoluted
60 into several steps including intravasation, circulation, margin-
61 ation, extravasation and colonization, is a crucial cause of deaths
62 in cancer therapy [31,32]. It had been reported that intrahepatic me-
63 tastases were observed in about 30% of cases after surgical removal
64 of small HCC nodules and in 80% of HCC autopsy cases [33]. Via EMT,
65 epithelial cells dissolve intercellular connections and acquire
66

mesenchymal properties and metastasize to the native or distant
67 sites. Once cancer cells seed the metastatic site, a mesenchymal to
68 epithelial transition (MET) occurs, inducing colonization and growth
69 of the metastatic foci with re-expression of cell adhesion mole-
70 cule E-cadherin, facilitating tumor cells to seed in the metastatic
71 sites [34–40]. E-cadherin re-expression accompanied by a partial
72 MET in the metastatic sites increases post-extravasation survival of
73 the cancer cells and resistance to multi-drugs [35,37]. Addition-
74 ally, further study found that a variety of biological molecules such
75 as HIF, TGF and miRNAs involved in the process of MET through dif-
76 ferent signaling pathways respectively accelerated the formation of
77 distant tumor metastasis [41–43]. In epithelial cancers, EMT re-
78 sulted in the loss of E-cadherin and in turn, tumor cells attain
79 enhanced migratory and invasive potential. E-cadherin, whose down-
80 regulated expression is a main biomarker for activation of EMT,
81 shows reverse relationship with drug resistance [44]. By undergo-
82 ing EMT, the tumor cells acquired resistance to a number of chemo-
83 and radiotherapies [34,45–48]. The relationship between epithelial–
84 mesenchymal transition (EMT) and drug resistance was first
85 described in connection with cancer stem cells by Mani et al., who
86 inferred that blocking or reversing EMT might cause chemoresistant
87 cells to revert to chemosensitive cells [49]. In addition, in breast
88 cancer therapy, it was reported that epithelial cells acquire some
89 cancer stem cells properties via EMT, such as anti-apoptosis and drug
90 resistance [50]. In the process of EMT in tumor cells, some biolog-
91 ical molecules, such as TGF β , Slug and FOXC2, play very crucial role
92 in explaining the sensitivity of tumor cells to chemotherapy drugs.
93 Activation of TGF β or FOXC2 and overexpression of Slug impaired
94 the sensitivity of tumor tissues to drug [49,51,52]. The drug resis-
95 tance was also reported in correlation with EMT in HCC [17–19].
96 Various signaling pathways have been involved in regulation of EMT
97 program [27], indicating that these signaling pathways and mole-
98 cules involved are potentially associated with drug resistance. The
99 Raf/MEK/ERK pathway represents a dominant signaling network pro-
100 moting proliferation and metastasis and is the main jamming target
101 pathway of sorafenib [53–56]. The genetic and molecular network
102 involved in the signal pathway is very complicated. In fact, some
103 studies have demonstrated that treatment with Raf kinase inhibi-
104 tors can even paradoxically induce ERK cascade signaling by
105 promoting dimerization of Raf family members [57,58]. The
106 biomarkers for sorafenib efficacy, such as Plasma c-KIT, hepato-
107 cyte growth factor and angiopoietin-2, are likely to be downregulated
108 in the sorafenib sensitive cell lines while the phospho-Akt and p85
109 (a regulatory subunit of PI3K) are upregulated in sorafenib resis-
110 tant cell lines [19,59,60]. Kunnimalaiyaan et al. hypothesized that
111 Notch signaling pathway is a potential regulator of EMT program
112 in HCC and NICD3 protein increased E-cadherin and activated tran-
113 scription factors Snail, Slug, and MMPs and induced EMT [27], which
114 might lead to development of sorafenib resistance. Acquired resis-
115 tance to sorafenib in HCC might also be mediated by the activation
116 of phosphatidylinositol 3-Kinase/Akt signaling pathway [19,61].
117 Overexpression of both α B-Crystallin (Cryab) and 14-3-3 ζ can induce
118 epithelial–mesenchymal transition (EMT) in HCC cells through ac-
119 tivation of the extracellular-regulated protein kinase (ERK) cascade,
120 a crucial up-stream factor of ERK1/2/Fra-1/sluc signaling pathway,
121 which can counteract the effect of sorafenib [62–67]. Therefore, it
122 is postulated that sorafenib resistance is closely correlated with the
123 dynamic changes of numerous molecules, especially their up- or
124 down-regulated expression, involved in the mutual transition
125 between epithelial and mesenchymal cell phenotypes and various
126 signaling pathways. Particularly, epithelial cells are more suscep-
127 tible to a Raf kinase inhibitor, sorafenib, whereas mesenchymal cells
128 showed significant resistance [68,69]. In addition, PI3K/Akt showed
129 higher activity and integrin-linked kinase (ILK) exhibit in the mes-
130 enchymal cells as compared to epithelial cells [70,71]. Many
131 pathways and molecules involved in these pathways play a crucial
132

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