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Short communication: Influence of storage and preservation on microbiological quality of silo ovine milk

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ABSTRACT

This study was designed to analyze the effects of the storage and preservation conditions on counts of mesophilic, thermoduric, psychotrophic, coliform, *Escherichia coli*, *Streptococcus agalactiae*, and *Staphylococcus aureus* organisms in silo ovine milk. A total of 910 analytical determinations were conducted from aliquots of 10 silo ovine milks. The conditions tested were unpreserved and azidiol-preserved milk stored at 4°C, and unpreserved milk stored at −20°C. Milk aged 2, 24, 48, 72, and 96 h post-collection for refrigerated aliquots, and 7, 15, and 30 d post-collection for frozen aliquots. The factors silo and storage conditions significantly contributed to variation of all microbiological variables, although milk age effect within storage was only significant for mesophilic, psychrotrophic, and coliform bacteria counts. In refrigerated raw milk, mesophile, psychrotroph, and coliform counts significantly increased over 96 h post-collection, whereas the other groups and bacteria species tested maintained their initial concentration. In all cases, azidiol preservation maintained the initial bacterial concentration in raw sheep milk under refrigeration throughout 96 h. Thus, azidiol was a suitable preservative for microbiological studies in sheep milk. Smallest counts were registered for frozen samples, particularly for coliforms, *E. coli*, *Strep. agalactiae* and *Staph. aureus*. Estimates of mesophilic, thermoduric and psychrotrophic organisms showed similar values on both azidiol-preserved and frozen milk samples. Coliforms and *E. coli* counts significantly decrease over time after freezing. Consequently, freezing at −20°C could also be appropriate for analysis of mesophilic, thermoduric, and psychrotrophic bacterial groups, but not for coliforms or mammary pathogens.

Key words: silo milk, bacterial culture, dairy sheep, milk microbiology

Short Communication

Silo and tank milk are both contaminated by bacteria from different sources, such as flora and pathogens present in beds, milking facilities, wash water, milking systems, udders, teats and teat canals, or mastitic milk. Some of these bacteria are pasteurization-resistant or are able to grow at low temperatures. These characteristics may hinder industrial dairy processing. Some of these species may also be pathogens for humans. Despite this, only aerobic mesophile count determination has been the target of various legal limits or quality payment schemes proposed by different countries. Thus, Regulation (EC) 853/2004 (European Commission, 2004) lays down mesophilic flora limit criterion for milk from other species than cows as ≤500,000 cfu/mL, when the final destination of milk does not include heat treatment; or 1,500,000 cfu/mL for heat-treated milk before processing. However, this policy makes no reference to other microbiological criteria, so no regulation exists on other bacterial standards of microbiological quality of sheep milk for many sheep milk-producing countries (e.g., Spain). In this context, other bacterial groups and species studied, such as thermoduric and psychrotrophic flora, coliforms, and *Escherichia coli* or mastitis-causing pathogens would be of great interest for ovine milk hygiene, safety, quality, and marketing. In all cases, knowledge of the influence of storage and preservation on sheep milk microbiological quality is important both to the farmer and the dairy industry to standardize sampling protocols, to ensure accuracy in test results and to optimize milk storage conditions.

Azidiol (**AZ**) is a widely used preservative to keep milk samples for several tests in dairy laboratories, although its effects on the viability of major bacterial groups and pathogens in sheep milk need to be well established when such samples are going to be used for microbiological purposes. In this sense, other preservatives (e.g., bronopol) significantly decreased the viability of milk bacterial species and groups (Shepherd et al., 1988; Amores et al., 2010) so AZ effect on sheep milk microbiology should be investigated. Similarly, milk freezing could be of remarkable interest in micro-

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biological studies on sheep milk and in mastitis-control plans due to its potential to decrease processing and collection costs and extend the analytical operation thereof. Thus, studies on goat milk (Sánchez et al., 2003; Amores et al., 2010) show that freezing may be used as a storage method for the study of some bacterial species or groups but not for others. Nonetheless, freezing effect on the viability of major bacterial groups and species present in sheep milk has not yet been studied.

The knowledge of preservation and storage effects on recovering bacterial groups and bacterial pathogens over time would allow the optimum analytical conditions for ovine milk to be defined. This knowledge is needed to implement adequate operational strategies and sampling protocols for practical analysis and research in this species. Silo milk is especially suitable for this purpose as it consists of large mixtures of milk from a large number of herds, which makes the study of different bacterial groups and pathogens possible at the same time. Additionally, such large mixes of milk are the raw material of all dairy products produced by the industry so its microbiological quality has a great economical, technological, hygienic, and marketing interest.

The aim of this study was to determine the effects of the most common storage and preservation conditions on the following microbiological quality variables: aerobic mesophilic, psychrotrophic, thermoduric, coliforms, *E. coli*, *Streptococcus agalactiae,* and *Staphylococcus aureus* bacteria.

Between November 2009 and February 2010, a total of 10 samples of silo sheep milk were collected at a milk plant, which includes milk from 10 different tank milk collection routes, from a total of 400 dairy sheep herds. According to standards recommended by the American Public Health Association (White et al., 1992), samples (500 mL) were aseptically collected in sterile containers from each of the silos immediately after milk tankers were unloaded. For this experiment, milk stored in each silo corresponded to a single milk tanker from a single collection route. Milk collection frequency in farms was always 48 h, during which the milk was kept at a temperature lower than 6°C in cooling tanks in the farms. Milk collection was carried out at the same time in each flock. Silo milk temperature was 4°C, maintaining that temperature until the bacteriological analysis, which was carried out immediately after arrival in the laboratory in the Department of Food Hygiene and Technology, University of León, Spain. Bulk tank milk of all flocks was periodically checked for antimicrobial detection by Eclipse-100ov screening test (ZEU-Immunotec, Zaragoza, Spain; Montero et al., 2005) in the Dairy Interprofessional Laboratory of Castilla-León region (Spain). In addition, before unloading in silos, tanker milks were always checked for β-lactams and tetracycline drugs by Rosa Charm rapid screening test (Charm Sciences, Inc., Lawrence, MA). Negative results were always obtained during the experiment.

The initial homogenized sample was divided into 13 aliquots of 40 mL each: 5 aliquots of unpreserved milk were kept refrigerated at 4°C, 5 aliquots of milk were preserved with AZ (Panreac Quimica S.A., Castellar del Vallès, Barcelona, Spain) and kept refrigerated at 4°C, and 3 aliquots of unpreserved milk were kept frozen at −20°C. Azidiol concentration in preserved samples was always 3.3 μL/mL (i.e., 133 μL/40 mL). Azidiol composition was 75 mg of chloramphenicol, 1 mL of ethanol, 1.8 g of sodium azide, 4.5 g of trisodium citrate·$5H_2O$, and 35 mg of bromophenol blue in 100 mL of distilled water. Bacteriological analysis of refrigerated aliquots was carried out at 2, 24, 48, 72, and 96 h post-collection. Frozen aliquots were defrosted at 4°C overnight and analyzed at 7, 15, and 30 d post-collection.

Total aerobic plate count determination was performed following the standards recommended by the American Public Health Association (APHA) for milk and dairy products (White et al., 1992). The total number of viable bacterial cells was determined by the SPC method. Milk samples were subjected to serial dilution in the 10^{-1} to 10^{-5} range and inoculated into plate count agar (PCA; Oxoid Limited, Cambridge, UK) petri plates. The inoculated plates were incubated at 30 ± 1°C for 48 h. Thermoduric count was carried out by the SPC method after laboratory pasteurization at 62.8 ± 0.5°C for 30 min following APHA recommendations (White et al., 1992). Psychrotrophic bacteria count was also performed by SPC, plates being incubated at 7°C between 7 and 10 d (White et al., 1992). The enumeration of coliforms and *E. coli* was carried out using 3M Petrifilm *E. coli*/coliform count plates (3M, St. Paul, Minnesota) according to the manufacturer instructions. In all cases plates were inoculated with 1 mL of milk sample dilution in the range of 10^{-1} and 10^{-3}. Plates were incubated at 37 ± 0.5°C for 24 to 48 h. Enumeration of each group consisted of considering as confirmed coliforms red and blue colonies with associated gas bubbles. Confirmed *E. coli* were considered as blue colonies with associated gas bubbles. Results were expressed as cfu/mL. Regulation UNE-EN ISO 6888–2:1999/ Amd 1:2003 (ISO, 2003) was used as the reference method for *Staph. aureus* enumeration. Finally, Edwards Medium Modified (Oxoid Limited, Cambridge, UK) supplied with 5 to 7% defibrinated sheep blood (Oxoid Limited) was used for *Strep. agalactiae* detection and enumeration (Zadoks et al., 2004). Incubation was at 35°C for 48 h. Hemolytic and nonhemolytic, esculin-negative blue colonies were suspected of being *Strep. agalactiae*. These colonies were tested for confirmation

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