



Research paper

A natural “GA” insertion mutation in the sequence encoding the 3′UTR of CXCL12/SDF-1 α : Identification, characterization, and functional impact on mRNA splicing

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ABSTRACT

The CXCL12 gene produces a series of transcript variants through alternative splicing at the 3′ end of its pre-mRNA. This study explores the biological activities of these alternative transcripts and the mechanisms involved in the regulation of CXCL12 transcription and RNA splicing. We identified a “GA” insertion mutation in the region of CXCL12 α DNA encoding the conserved 3′UTR. This variant transcript was named CXCL12-3′GA+. The mutation occurred at a frequency of 13.2% in healthy Chinese individuals. However, its frequency in healthy Caucasians was 22.6%, significantly higher than what was observed in the Chinese. Genomic analysis indicated that the GA+ mutation likely encodes a G-quadruplex structure in close proximity to a cluster of important AU-rich elements (AREs) that are well-established regulators of mRNA stability at the 3′UTR. Experiments using molecular constructs encoding the 3′UTR of CXCL12 revealed that the GA+ allele can significantly increase gene expression compared to the WT allele. Further studies uncovered that the WT allele was associated with the production of a 225-bp minor transcript isoform (MTI) through alternative splicing resulting in the deletion of exon 2. ARMS-PCR using samples collected from cultured PBMCs of WT/GA+ genotype carriers indicated that the GA+ allele was preferentially transcribed compared to the WT allele. In summary, the study demonstrates that a GA insertion in the region encoding the 3′UTR of CXCL12 α may affect gene expression through alternative mRNA splicing. This finding provides a basis for understanding how multiple elements in the sequence encoding the 3′UTR of the CXCL12 gene regulates its transcription and may lead to insights about diseases involving abnormal CXCL12 α expression.

1. Introduction

The C-X-C motif chemokine ligand (CXCL12), also known as stromal cell derived factor (SDF)-1, and pre-B-cell growth stimulating factor precursor (PBSF), was originally isolated from a murine bone marrow stromal cell line (Nagasawa et al., 1996; Aiuti et al., 1997). This chemokine, along with its receptors CXCR4 and CXCR7, is believed to play

a critical role in HIV-1 infection, the metastatic spread of tumor cells, and the development and maintenance of the nervous, hematopoietic, and cardiovascular systems (Burger and Kipps, 2006; Petit et al., 2007; Sierro et al., 2007; Wang et al., 2008). A previous study showed that the CXCL12 gene is located on human chromosome 10q and spans over 88 kb. In contrast, other CXC chemokine genes are clustered in human chromosome 4q. The amino acid (aa) sequence of the CXCL12 protein

Abbreviations: aa, amino acid(s); AREs, AU-rich elements; ARMS, amplification refractory mutation system; bp, base pair(s); cDNA, DNA complementary to RNA; cpm, counts per minute; CXCL, C-X-C motif chemokine ligand; CXCR, C-X-C chemokine receptor; d, deoxyribo; DMSO, dimethyl sulfoxide; dNTP, deoxyribonucleoside triphosphate; HIV, human immunodeficiency virus; MTI, minor transcript isoform; MU, mutant; ORF, open reading frame; PBL, peripheral blood lymphocytes; PBMC, peripheral blood mononuclear cells; PBSF, pre-B-cell growth stimulating factor precursor; PBS, phosphate buffer saline; PCR, polymerase chain reaction; PHA, phytohemagglutinin; PTC, premature termination codon; QGRS, G-rich quadruplex sequences; RIPA, radio immune-precipitation assay; RT, reverse transcription; SDF, stromal cell derived factor; Tat, trans-activator of transcription; UTR, untranslated region(s); WT/wt, wild type; Δ , deletion

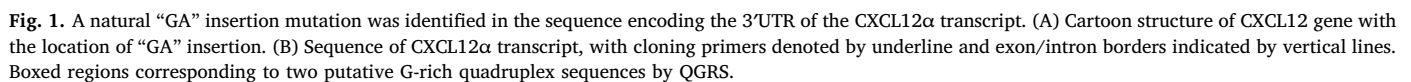
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To date, six functional transcript variants of the CXCL12 gene have been identified. The cDNA of CXCL12 α encodes a protein that is 89 aa long. The remaining variants, CXCL12 β , CXCL12 γ , CXCL12e, CXCL12 δ , and CXCL12 θ , are generated by alternative splicing, with different exons spliced with the same first three exons (Yu et al., 2006; Altenburg

et al., 2007; Laguri et al., 2007). This implies that the first 88 residues (encoded by the first 3 exons) of these splicing variants of CXCL12 are identical and that they are likely essential for binding to CXCR4 (Altenburg et al., 2007; Laguri et al., 2007; Sierro et al., 2007). These CXCL12 splicing variants have been detected in a variety of tissues, including the liver, pancreas, spleen, and heart. The specific mechanisms governing their biological activities, such as in hematopoiesis, chemotaxis, and HIV-1 blocking, remain under investigation (Schols et al., 1997; Wang et al., 2000; Altenburg et al., 2007; Petit et al.,

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