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## Supercritical fluid extraction of quinolizidine alkaloids from Sophora flavescens Ait. and purification by high-speed counter-current chromatography

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#### **Abstract**

Supercritical fluid extraction (SFE) was used to extract quinolizidine alkaloids from *Sophora flavescens* Ait. (Kushen). An orthogonal test  $L_9(3)^4$  including pressure, temperature, flow rate of  $CO_2$  and the amount of modifier was performed to get the optimal conditions. The process was then scaled up by 30 times with a preparative SFE system under 25 MPa, 50 °C and a flow rate of  $CO_2$  (2 l/min) and the amount of modifier (0.04 ml/min). The crude extracts were separated and purified by high-speed counter-current chromatography (HSCCC) with a two-phase solvent system composed of chloroform–methanol– $2.3 \times 10^{-2}$  M NaH<sub>2</sub>PO<sub>4</sub> (27.5:20:12.5, v/v), and the collected fractions were analyzed by high-performance liquid chromatography (HPLC). Three kinds of quinolizidine alkaloids were obtained, yielding 10.02 mg of matrine, 22.07 mg of oxysophocarpine and 79.93 mg of oxymatrine with purities of 95.6, 95.8, 99.6% in one-step separation, respectively.

Keywords: Supercritical fluid extraction; High-speed counter-current chromatography; Quinolizidine alkaloids; Sophora flavescens Ait

#### 1. Introduction

The dried root of *Sophora flavescens* Ait. (Leguminosae), a typical traditional Chinese medicine, is commonly used for the treatment of viral hepatitis, cancer, viral myocarditis, gastrointestinal hemorrhage and skin diseases (such as colpitis, psoriasis and eczema) [1–3]. The principal bioactive constituents of *S. flavescens* Ait. are the major quinolizidine alkaloids matrine (MT) and oxymatrine (OMT), which were reported to exhibit sedative, depressant, anti-tumor, antipyretic, cardiotonic activities [4,5] and anti-hepatitis B virus (HBV) activity [6]. Oxysophocarpine (OSC), another alkaloid, obviously suppressed the biosynethesis of leukotrienes (LTC4 and LTB4) in dose-dependent manner [7]. OSC, MT and OMT (structure shown in Fig. 1) were also found in other plants of Leguminosae, such as *Sophora tonkinensis*, *Sophora subprostata* and *S. alopecuroides* [8,9].

Due to the high pharmacological activities, alkaloids from the root of S. flavescens Ait. has recently drawn great attention in natural medication researches. A large quantity of pure materials is urgently needed for further studies. Several methods such as high-performance liquid chromatography (HPLC), silica gel, polyamide column and thin-layer chromatography (TLC) have been applied to the separation and purification of matrine-type alkaloids in S. flavescens root. Undoubtedly, HPLC is the most widely used separation technique [10]. However, the conventional preparative separation and purification methods are tedious and time-consuming, requiring multiple chromatographic steps. As for HPLC, reconditioning of the column requires a long time and a large volume of organic solvent. Expensive columns and frequent changing of the columns for fear of loss of elution efficiency are also required. Hence, sensitive, rapid and specific methods for purification of quinolizidine alkaloids are of great interest.

As an alternative, supercritical fluid extraction (SFE) is a particularly suitable method for the research of natural materials. Carbon dioxide is an ideal solvent because it is non-toxic, non-explosive, readily available and easy to remove from extracted

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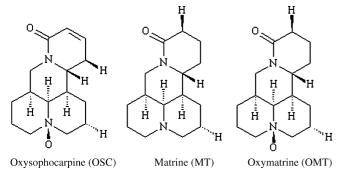


Fig. 1. Chemical structures of alkaloids from Sophora flavescens Ait.

products. SFE has the ability to use low temperatures, leading to less deterioration of the thermally labile components in the extract [11–16]. In addition, SFE using carbon dioxide ensures minimal alteration of the active ingredients, and the curative properties can be preserved.

High-speed counter-current chromatography (HSCCC) is a unique liquid–liquid partition chromatography technique that uses no solid support matrix [17]. HSCCC eliminates the irreversible adsorptive loss of samples onto the solid support matrix used in the conventional chromatographic column. This method has been successfully applied to the separation and purification of several natural products [18–22]. No reports on the use of SFE to extract and HSCCC to isolate matrine, oxysophocarpine and oxymatrine from natural plants have been found.

We herein optimized experiment parameters by an analyticalscale SFE system using an orthogonal test design. Then, the extraction was scaled up by 30 times by a prepared-scale SFE system. Subsequently, the crude extract was purified by HSCCC.

### 2. Experimental

#### 2.1. Reagents and materials

Carbon dioxide (99.9% purity) was obtained from Daxing Gas Co., Beijing, China. All organic solvents used for HSCCC were of analytical grade and purchased from Guangcheng Chemical Factory, Tianjin, China. HPLC-grade acetonitrile was obtained from Tedia, USA. The roots of *S. flavescens* Ait. were purchased from a local drug store of Jinan, Shandong Province, in August 2006. Three quinolizidine alkaloids, matrine, oxysophocarpine and oxymatrine, were purchased from National Institute of the Control of Pharmaceutical and Biological Products, Beijing, China, and diluted to the desired concentration prior to use.

Compared with the voucher specimen collected from Bozhou, Anhui Province, the dried roots of *S. flavescens* were identified by Professor Yongqing Zhang (Shandong University of Traditional Chinese Medicine, Jinan, China).

#### 2.2. Optimization of SFE extraction

A Spe-ed SFE system (Applied Separations, Allentown, PA, USA) fitted with a 10 ml stainless-steel extraction vessel was

Table 1 Orthogonal experimental design

	A, pressure (MPa)	B, temperature $(^{\circ}C)$	C, flow rate of CO <sub>2</sub> (l/min)	D, flow rate of modifier (ml/min)
1	20	45	1.0	0.02
2	25	50	1.5	0.03
3	30	55	2.0	0.04

used for optimization of the extraction conditions with modifier adding by a WellChrom K-501 HPLC pump (Knauer, Berlin, Germany). A micro-metering valve was used as restrictor valve to control the flow rate of the supercritical  $CO_2$  to the solvent collection. Extraction temperatures were monitored using a thermocouple and were found to be accurate to within  $\pm 1$  K. The precision of the pressure measurement was  $\pm 1$  Pa.

To get more effective extraction, the roots of *S. flavescens* Ait. were shattered to powder (60–80 mesh) and dipped in 0.1 ml/l ammonia–ethanol at the ratio of 1:4 (v/v) for 24 h. An orthogonal test design  $L_9(3)^4$  was employed where temperature, pressure, flow rate of  $CO_2$  and the flow rate of 75% ethanol and 25% water as a modifier were considered to be the four major factors for effective extraction. Combinations of the three different levels of each factor are listed in Table 1. In each test, 5.500 g *S. flavescens* (60–80 mesh) was placed into the extraction vessel. After 0.5 h of static extraction (no liquid flow), the sample was subjected to dynamic extraction by flowing  $CO_2$  at a set rate for 2 h.

#### 2.3. Scaling-up SFE

Under the optimized SFE conditions determined above, the extraction was scaled up by 30-fold using a 1000 ml vessel. A 165 g amount of sample was extracted statically for 1 h and then dynamic extraction done for 3 h by flowing liquid  $CO_2$  at a rate of 2 l/min; the extract was depressed directly into a collection vessel and stored in a refrigerator for subsequent HPLC analysis and HSCCC separation.

# 2.4. Preparation of two-phase solvent system and sample solution

The selected solvent system, chloroform—methanol— $2.3 \times 10^{-2}$  M NaH<sub>2</sub>PO<sub>4</sub> (27.5:20:12.5, v/v), was prepared by adding all the solvents to a separation funnel according to the volume ratios and thoroughly equilibrated by shaking separately. After being thoroughly equilibrated, the upper phase and lower phase were separated and degassed by sonication for 45 min prior to use. The sample solution was prepared by dissolving the crude sample in 5 ml upper phase of the solvent system.

#### 2.5. HSCCC separation

The extracted alkaloids from *S. flavescens* Ait. were separated by a TBE-300A high-speed counter-current chromatograph

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