

Review

Trace mycotoxin analysis in complex biological and food matrices by liquid chromatography–atmospheric pressure ionisation mass spectrometry

Peter Zöllner^{a,*}, Bernhard Mayer-Helm^b^a Bayercropscience GmbH, Product Technology, Industriepark Höchst, G836, D-65926 Frankfurt/Main, Germany^b Department of Clinical Pharmacology, Medical University of Vienna, Währinger Gürtel 18-20, A-1090 Wien, Austria

Received 23 March 2006; received in revised form 14 September 2006; accepted 19 September 2006

Available online 7 November 2006

Abstract

Mycotoxins are toxic secondary metabolites produced by filamentous fungi that are growing on agricultural commodities. Their frequent presence in food and their severe toxic, carcinogenic and estrogenic properties have been recognised as potential threat to human health. A reliable risk assessment of mycotoxin contamination for humans and animals relies basically on their unambiguous identification and accurate quantification in food and feedstuff. While most screening methods for mycotoxins are based on immunoassays, unambiguous analyte confirmation can be easily achieved with mass spectrometric methods, like gas chromatography/mass spectrometry (GC/MS) or liquid chromatography/mass spectrometry (LC/MS). Due to the introduction of atmospheric pressure ionisation (API) techniques in the late 80s, LC/MS has become a routine technique also in food analysis, overcoming the traditional drawbacks of GC/MS regarding volatility and thermal stability. During the last few years, this technical and instrumental progress had also an increasing impact on the expanding field of mycotoxin analysis. The aim of the present review is to give an overview on the application of LC–(API)MS in the analysis of frequently occurring and highly toxic mycotoxins, such as trichothecenes, ochratoxins, zearalenone, fumonisins, aflatoxins, enniatins, moniliformin and several other mycotoxins. This includes also the investigation of some of their metabolites and degradation products. Suitable sample pre-treatment procedures, their applicability for high sample through-put and their influence on matrix effects will be discussed. The review covers literature published until July 2006.

© 2006 Elsevier B.V. All rights reserved.

Keywords: Mycotoxin; Metabolite; Degradation product; Bioadduct; Bioconjugate; Trichothecene; Ochratoxin; Zearalenone; Zeranone; Fumonisin; Aflatoxin; Patulin; Moniliformin; Enniatin; Beauvericin; Mycophenolic acid; Mass spectrometry; High-performance liquid chromatography; LC/MS; Electrospray ionisation; Atmospheric pressure ionisation; Atmospheric pressure chemical ionisation; Atmospheric pressure photoionisation; ESI; APCI; APPI

Abbreviations: 3-/15-AcDON, 3-/15-acetyldeoxynivalenol; ACN, acetonitrile; AcOH, acetic acid; AFB₁₋₂, aflatoxin B₁₋₂; AFG₁₋₂, aflatoxin G₁₋₂; AFM₁, aflatoxin M₁; AFQ₁, aflatoxin Q₁; AFP₁, aflatoxin P₁; APCI, atmospheric pressure chemical ionisation; API, atmospheric pressure ionisation; APPI, atmospheric pressure photo ionisation; BEA, beauvericin; CID, collision induced dissociation; CRM, consecutive reaction monitoring; DAS, diacetoxyscirpenol; DOM-1, deepoxydeoxynivalenol; DON, deoxynivalenol; EA, EA₁, enniatin A, enniatin A₁; EB, EB₁, enniatin B, enniatin B₁; ELISA, enzyme-linked immuno sorbent assay; ESI, electrospray ionisation; FAB, fast atom bombardment; FA₁₋₃, fumonisin A₁₋₃; FAK₁, fumonisin AK₁; FB₁₋₅, fumonisin B₁₋₅; FC₁₋₄, fumonisin C₁₋₄; FD, fumonisin D; FP₁₋₃, fumonisin P₁₋₃ = 3-hydroxypyridinium FC₁₋₃; FDA, food and drug administration; FL, fluorescence detection; FUS, fusaproliferin; F-X, fusarenone X; GC–ECD, gas chromatography–electron capture detection; GC/MS, gas chromatography/mass spectrometry; HFB₁, fully hydrolysed fumonisin B₁; HT-2, HT-2 toxin; IAC, immunoaffinity chromatography; LC/MS, liquid chromatography/mass spectrometry; LOD, limit of detection; LOQ, limit of quantification; MAS, monoacetoxyscirpenol; MeOH, methanol; MON, moniliformin; MPA, mycophenolic acid; MS/MS, tandem mass spectrometry; NanoESI, nano electrospray ionisation; NEO, neosolaniol; NIV, nivalenol; OTA, ochratoxin A; OTB, ochratoxin B; OTC, ochratoxin C; OTα, ochratoxin α; OTβ, ochratoxin β; PB, particle beam; PHFB₁, partially hydrolysed FB₁; QTrap, combination of triple quadrupole and ion trap mass analyser; RP, reversed-phase; SFC, supercritical fluid chromatography; SFE, supercritical fluid extraction; SIM, selected ion monitoring; SPE, solid-phase extraction; SRM, selected reaction monitoring; T-2, T-2 toxin; TFA, trifluoro acetic acid; TLC, thin-layer chromatography; TSP, thermospray; VER, verrucarol; WHO, World Health Organisation; ZAN, zearalenone; α-ZAL, α-zearalanol = zeranone; β-ZAL, β-zearalanol = taleranol; α-ZOL, α-zearalenol; β-ZOL, β-zearalenol; ZON, zearalenone

* Corresponding author. Tel.: +49 69 305 12248; fax: +49 69 305 21802.

E-mail address: peter.zoellner@bayercropscience.com (P. Zöllner).

Contents

1.	Introduction	124
2.	Trichothecenes	126
2.1.	Conventional trichothecene analysis	126
2.2.	Early LC/MS technology in trichothecene analysis	126
2.3.	Modern LC/MS analysis of trichothecenes	127
2.3.1.	Investigated trichothecene analytes and typical matrices	127
2.3.2.	Sample preparation	127
2.3.3.	Typical LC conditions for LC/MS analysis	127
2.3.4.	Mass spectrometry	129
2.3.5.	Method validation and matrix effects	131
3.	Ochratoxins	131
3.1.	Conventional ochratoxin analysis	132
3.2.	Early LC/MS technology in ochratoxin analysis	133
3.3.	Modern LC/MS analysis of ochratoxins	133
3.3.1.	Investigated ochratoxin-type analytes and typical matrices	133
3.3.2.	Sample preparation	133
3.3.3.	Typical LC conditions for LC/MS analysis	135
3.3.4.	Mass spectrometry	136
3.3.5.	Method validation and matrix effects	136
4.	Zearalenone and metabolites	137
4.1.	Conventional ZON analysis	138
4.2.	Early LC/MS technology for ZON analysis	139
4.3.	Modern LC/MS analysis of ZON	139
4.3.1.	Investigated zearalenone-type analytes and typical matrices	139
4.3.2.	Sample preparation	139
4.3.3.	Typical LC conditions for LC/MS analysis	140
4.3.4.	Mass spectrometry	140
4.3.5.	Method validation and matrix effects	142
5.	Fumonisin	145
5.1.	Conventional fumonisin analysis	145
5.2.	Early LC/MS technology for fumonisin analysis	146
5.3.	Modern LC/MS analysis of fumonisins	146
5.3.1.	Investigated fumonisin-type analytes and typical matrices	146
5.3.2.	Sample preparation	146
5.3.3.	Typical LC conditions for LC/MS analysis	149
5.3.4.	Mass spectrometry	149
5.3.5.	Method validation and matrix effects	151
6.	Aflatoxins	152
6.1.	Conventional aflatoxin analysis	152
6.2.	Modern LC/MS analysis of aflatoxins	152
6.2.1.	Investigated aflatoxin analytes and typical matrices	152
6.2.2.	Sample preparation	152
6.2.3.	Typical LC conditions for LC/MS analysis	152
6.2.4.	Mass spectrometry	153
6.2.5.	Method validation and matrix effects	157
7.	LC/MS analysis of other mycotoxins	157
7.1.	Enniatins and beauvericin	157
7.2.	Moniliformin	160
7.3.	Mycophenolic acid	160
7.4.	Patulin	161
8.	Multitoxin LC/MS analysis	164
9.	Conclusion	164
	References	165

1. Introduction

Mycotoxins are toxic secondary metabolites produced by about 200 identified filamentous fungi, as e.g. *Fusarium*,

Aspergillus and *Penicillium* species, growing under a wide range of climatic conditions on agricultural commodities (grains, spices, fruits, coffee, nuts, etc.) in the field and during storage [1,2]. Their occurrence in food, beverages and feed has

Download English Version:

<https://daneshyari.com/en/article/1209427>

Download Persian Version:

<https://daneshyari.com/article/1209427>

[Daneshyari.com](https://daneshyari.com)