



# Determination of Prostaglandin E<sub>1</sub> in dog plasma using liquid chromatography–tandem mass spectrometry and its application to a pharmacokinetic study



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## ABSTRACT

The determination of Prostaglandin (PG) E<sub>1</sub> in plasma is challenged by its low concentration (pg/mL) and endogenous interference. An LC–MS/MS method for the determination of PGE<sub>1</sub> in dog plasma has been developed and validated. Plasma being sampled at 4 °C and treated with indomethacin effectively inhibited interferents synthesized post-sampling. Samples were subjected to one-step extraction and separated by reversed phase HPLC with a short cycle time of 3 min. An LLOQ of 10 pg/mL was achieved with 500 μL plasma. The method was applied to a pharmacokinetic study in beagle dogs involving an intravenous infusion of 3.2 μg/kg PGE<sub>1</sub>. The half-life was recovered at 7 min. The simple, sensitive and rapid method was suitable to be applied to pharmacokinetic studies of PGE<sub>1</sub> at clinically relevant doses.

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## 1. Introduction

Prostaglandin (PG) E<sub>1</sub> (Fig. 1), also known as alprostadil, is a vasodilator widely used for the treatment of ischemic peripheral vascular disease [1–4]. It exerts multiple effects on peripheral vascular system, including increase of peripheral blood flow [5] and viscosity [6], modulation of fibrinolytic system [7], and inhibition of platelet aggregation [8]. The drug is also administered intra-urethral and intracavernosally as a second-line therapy for erectile dysfunction [9]. It has been known that PGE<sub>1</sub> is inactivated rapidly by lung [10,11]. Based on its short half-life, the drug is recommended to be administered by intravenous infusion to maintain

plasma PGE<sub>1</sub> concentrations and thus prolong its action for the treatment of peripheral arterial occlusive disease [12].

Recently, prolonged release preparations of PGE<sub>1</sub> were developed in order to expand its administration routes and facilitate clinical application. However, it was a challenge to evaluate these preparations by pharmacokinetic studies due to the difficulty to determine PGE<sub>1</sub> in plasma as well as in other biological samples. Furthermore, although the drug has been applied in clinical practice for decades, to date there are very few reports on its pharmacokinetics. The determination of plasma PGE<sub>1</sub> is not only challenged by its low levels (pg/mL) following a dose in microgram order [13], but also challenged by the endogenous interferents. Endogenous PGE<sub>1</sub> at normal physiological level is very low in plasma, such as 1–3 pg/mL in human plasma [14], and therefore does not significantly interfere the determination of exogenous PGE<sub>1</sub>. However, great amounts of interferents, probably PGE<sub>1</sub> and other PGs, could be biosynthesized from the phospholipids released from cell membrane residues remaining in plasma. The occurrence of post-sampling interference at high levels made it impossible to quantitation exogenous PGE<sub>1</sub> in low pg/mL order.

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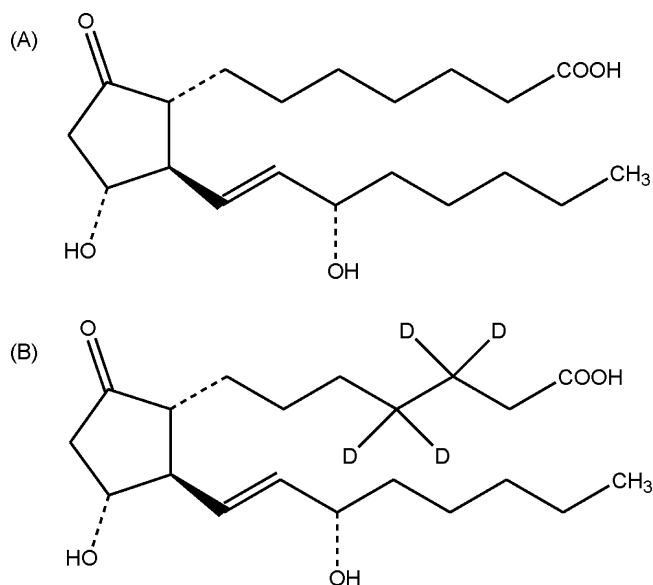


Fig. 1. Prostaglandin E<sub>1</sub> (A) and internal standard, PGE<sub>1</sub>-d<sub>4</sub> (B).

Several methods have been reported concerned with the determination of PGE<sub>1</sub> in biological matrix. Schweer et al. [14] and Hammes et al. [15] reported gas chromatography-tandem mass spectrometry (GC-MS/MS) methods for the determination of PGE<sub>1</sub> in human plasma, with a lower limit of detection (LOD) of 1 pg/mL and a lower limit of quantitation (LLOQ) of 2 pg/mL, respectively. Although both of the methods provided high sensitivity, they suffered from time-consuming sample preparation procedure of three step derivatization. High performance liquid chromatography (HPLC) based methods were developed by Hotter et al. [16] and Tsutsumiuchi et al. [17] with LLOQs of 3.9 pg/mL and 23 pg/mL, respectively. Except for analytical run time (30 min), long time was consumed on either radioimmunoassay (24 h) or derivatization (1 h). None of the methods discussed above was suitable to be applied to pharmacokinetic studies based on their low throughputs, e.g. only 24 samples could be prepared per day in the GC-MS/MS method with LLOQ of 2 pg/mL [15]. Rapid and simple LC-MS methods were provided by Abián et al. [18] and Lee et al. [19] with LLOQs of 50 ng/mL–20 µg/mL. Lin et al. reported an ultra-high performance liquid chromatography-tandem mass spectrometry (UPLC-MS/MS) method for the determination of PGE<sub>1</sub> in plasma and the method was applied to a pharmacokinetic study involving an intravenous injection of PGE<sub>1</sub> [20]. The method was rapid and simple with one-step sample preparation and run time of 2 min. However, it suffered from an LLOQ of 400 pg/mL, which led to a non-clinically-relevant dose of 50 µg/kg PGE<sub>1</sub> for the pharmacokinetic study in rats.

In order to solve these problems, a liquid chromatography-tandem mass spectrometric (LC-MS/MS) method for the determination of PGE<sub>1</sub> in dog plasma was developed and it has been successfully applied to a pharmacokinetic study in beagle dogs involving an intravenous infusion of PGE<sub>1</sub> at a therapeutically relevant dose of 3.2 µg/kg. The method involved one-step sample preparation, a short analytical time (cycle time 3 min) and high sensitivity (LLOQ 10 pg/mL). Indomethacin was added to plasma to inhibit post-sampling synthesis of endogenous interferences and low baselines were obtained. The sensitivity of this method (LLOQ 10 pg/mL using 0.5 mL plasma) was comparable with the GC-MS/MS method with an LLOQ of 2 pg/mL using 2 mL plasma, but the former method was much more simple and rapid. This method is being validated with human plasma and will be applied

to a phase I clinical study for PGE<sub>1</sub> micelles. An LLOQ of 1 pg/mL has been achieved using 2 mL human plasma.

## 2. Experimental

### 2.1. Chemicals and reagents

Prostaglandin E<sub>1</sub> (97.2% purity) and indomethacin (98% purity) were provided by Jilin Yinglian Biopharmaceutical Co., Ltd., (Changchun, PR China). PGE<sub>1</sub>-d<sub>4</sub> (9-oxo-11α, 15S-dihydroxyprost-13E-en-1-oic-3,3,4,4-d<sub>4</sub> acid) (99% purity) was purchased from Amyjet Scientific Inc. (Wuhan, PR China). Methanol and acetonitrile were HPLC grade and obtained from Fisher Scientific (Fair Lawn, NJ, USA). Ultra-high purity water, prepared using the Milli-Q system, was used through-out the study. All other chemicals were analytical grade and used without further purification. Alprostadil injection (Lot No. 2088S, 2 mL: 10 µg) was purchased from Beijing Tide Pharmaceutical Co., Ltd., (Beijing, PR China).

### 2.2. LC-MS/MS conditions

Chromatography was performed using an Agilent 1100 HPLC system (Agilent Technologies, CA, USA) equipped with an Agilent Zorbax Eclipse XDB C18 column (150 mm × 4.6 mm, 5 µm) maintained at 30 °C. The mobile phase consisted of acetonitrile-0.5% formic acid in water (50:50, v/v) delivered at a flow rate of 1 mL/min. Mass spectrometric detection employed an API 4000 mass spectrometer (AB Sciex, Ont., Canada) equipped with an electrospray ionization (ESI) source operated in the negative ion mode. The curtain gas, Gas 1 and Gas 2 were nitrogen set at 20, 60, and 60 psi, respectively. The ion spray voltage was adjusted to -4500 V and the source temperature was set at 500 °C. Detection was by multiple reaction monitoring (MRM) at unit resolution for Q1 and unit resolution for Q3 with a dwell time of 200 ms per channel. Transitions of [M-H]<sup>-</sup> ions for multiple reaction monitoring (MRM) were at *m/z* 353.4 → 317.3 and 353.4 → 235.0 for PGE<sub>1</sub> and *m/z* 357.2 → 321.1 for IS. Declustering potential and collision energy were -55 V and -80 eV for PGE<sub>1</sub> and IS. Mass spectrometric parameters were tuned to maximize the response of the precursor/product ion combinations. Data acquisition and integration were controlled by Applied Biosystems Analyst version 1.3 software.

### 2.3. Preparation of calibration standards and QC samples

A stock solution of PGE<sub>1</sub> (1 mg/mL) was prepared in methanol. Calibration standards of PGE<sub>1</sub> were prepared by diluting the stock solutions with blank dog plasma (prepared by separating plasma at 4 °C and transferring 1.2 mL plasma to a tube coated with 6 µg indomethacin, see Section 2.6) to concentrations of 10, 30, 100, 300, 1000 and 3000 pg/mL. Quality control (QC) solutions were prepared independently at concentrations of 30, 300 and 2400 pg/mL in the same way. A stock solution (4 µg/mL) of the internal standard (IS), PGE<sub>1</sub>-d<sub>4</sub> in methanol was diluted in methanol-water (50:50, v/v) to give a 500 pg/mL working PGE<sub>1</sub>-d<sub>4</sub> solution. All solutions were stored at 4 °C.

### 2.4. Sample preparation

To 500 µL plasma or calibration standard or QC sample in a glass tube, 100 µL 50% formic acid and 100 µL IS working solution were added. The mixture was then shaken with 3 mL diethyl ether-dichloromethane (3:2, v/v), centrifuged for 5 min at 3500 × *g* and the organic layer transferred to another glass tube and evaporated to dryness at 40 °C under nitrogen. The residue was reconstituted in

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