

Contents lists available at ScienceDirect

Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy

journal homepage: www.elsevier.com/locate/saa

Influence of charge and metal coordination of *meso*-substituted porphyrins on bacterial photoinactivation



SPECTROCHIMICA ACTA

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Tamara Zoltan^{a,*}, Franklin Vargas^a, Verónica López^a, Valery Chávez^b, Carlos Rivas^a, Álvaro H. Ramírez^c

^a Laboratorio de Fotoquímica, Centro de Química, Instituto Venezolano de Investigaciones Científicas I.V.I.C., Apartado 20632, Caracas 1020-A, Venezuela

^b Laboratorio de Bioquímica Celular, Centro de Microbiología y Biología Celular, Instituto Venezolano de Investigaciones Científicas I.V.I.C., Apartado 20632, Caracas 1020-A, Venezuela ^c Laboratorio de Biología de virus, Centro de Microbiología y Biología Celular, Instituto Venezolano de Investigaciones Científicas I.V.I.C., Apartado 20632, Caracas 1020-A, Venezuela

HIGHLIGHTS

- Were synthesized and characterized 8 *meso* substituted porphyrins with different substituents.
- Effect of charges and metal coordination in their photochemical characteristics was studied.
- We performed a detailed study of photoinduced antibacterial structure–activity.
- It was obtained a relation among the structural integrity, ROS production and antibacterial activity.

ARTICLE INFO

Article history: Received 23 May 2014 Received in revised form 7 July 2014 Accepted 17 July 2014 Available online 1 August 2014

Keywords: Photoinactivation Water-soluble metallic porphyrins Reactive oxygen species Escherichia coli Metal and charge effect

G R A P H I C A L A B S T R A C T

A: The series of TNPS_4 have higher structural deformity, low production of ROS and reduced antibacterial activity. **B**: The series of TPPS_4 have a low structural deformity, high production of ROS and the greater antibacterial activity.



ABSTRACT

The photodynamic effect of meso-substituted porphyrins with different charges and metal ions: meso-tetraphenylporphyrin tetrasulfonate 1, its nickel 2 and zinc complexes 3; meso-tetranaphthylporphyrin tetrasulfonate 4, and its zinc complex Zn 5; and tetra piridyl ethylacetate porphirins 6 and their nickel 7 and zinc 8 complexes, were synthesized and studied their antimicrobial activity against Escherichia coli. Fluorescence quantum yields (Φ_F) were measured in water using reference TPPS₄, obtaining higher values for complexes **3** and **4**. The singlet oxygen Φ_A were measured using histidine as trapping singlet oxygen and Rose Bengal as a reference standard. Complexes 1, 2 and 6 have the highest quantum yields of singlet oxygen formation, showing no relation with the peripheral charges and efficiency as Type II photosensitizers. Meanwhile complexes 3, 8 and 4 were the most efficient in producing radical species, determined with their reaction with NADH. The photoinduced antibacterial activity of complex was investigated at different concentrations of the photosensitizers with an irradiation time of 30 min. The higher antibacterial activities were obtained for the complexes 1-3 that are those with greater production of ROS and minor structural deformations. Complexes **7** and **8** had moderate activity, while **4–6** a low activity. Thus, in this work demonstrates that the production of ROS and structural deformations due to peripheral substituents and metal coordination, influence the activity of the complexes studied. Therefore, is important to perform comprehensive study physics and structurally when predicting or explain such activity. © 2014 Elsevier B.V. All rights reserved.

* Corresponding author. *E-mail address:* tzoltan@ivic.gob.ve (T. Zoltan).

Introduction

Due to the wide variety of pathogens and how fast their evolutionary changes to become resistant, the field of antimicrobial chemotherapy is an area of study in constant development. Photodynamic therapy (PDT) has been successfully applied in treating cancer as well as in non-cancerous condition [1,2]. Furthermore, there has also been an increasing interest in the application of PDT in the treatment of infectious diseases [3–5]. PDT involves the use of non-toxic dyes, which act as photoactive drugs called photosensitizers (PS) with the visible light of the proper wavelength to excite the PS. The light raises the PS to an excited state, which then interacts with oxygen, leading to the formation of reactive oxygen species (ROS) such as singlet oxygen, resulting in cytotoxicity and direct cell kill [6].

New developments in PDT applications are several approaches, which innovate in the design of PS is very wide. The chemical structure of a photosensitizer, charge and hydrophobicity influences their efficiency of ROS productions and determines how it interacts with itself and with its environment. Theoretically, photosensitizers can be properly designed a function of its chemical structure, but more realistically their effectiveness is determined as a function of the targets to be confronted [7].

The design of porphyrins as PS with microbial activity PDT has been extensively studied mainly focused on develop cationic complexes, due to possibility of interaction with the cell membrane [8,9]. The development of neutral or anionic porphyrins, regardless of their efficiency to produce reactive oxygen species, does not receive great attention because of its low interaction on the membrane [10,11]. Many of those studies report that the latter porphyrins have negligible activity against Gram-negative bacteria such as Escherichia coli [12]. It is important to note that the experimental method, bacterial washes are performed system before irradiation, thereby eliminating the PS not attached to the bacteria [13,14]. Taking into account that in large-scale applications, such as in wastewater treatment, these washings not performed, the efficiency of ROS production in the entire system plays an important role in bacterial photoinactivation. Thus, evaluating new photosensitizers on bacterial cultures without pre-irradiation washes, would allow obtaining information about the system under real operating conditions. In this sense, in this work the synthesis and photochemical characterization 8 hydrophilic porphyrins were performed. The aim of this paper is to examine their efficiency as PS in cultures of *E. coli* (no previous washes), and the impact of structural changes in this efficiency.

In this paper the synthesis of the following water-soluble porphyrins is presented: *meso*-tetraphenylporphyrin (TPP) tetrasulfonate (S_4): (TPPS₄, **1**), its nickel (TPPNiS₄, **2**) and zinc complexes (TPPZnS₄, **3**) as well as the *meso*-tetranaphthylporphyrin tetrasulfonate (TNPS₄, **4**) its Zn complexes (TNPZnS₄, **5**) and tetra piridyl ethylacetate porphirins (TPyEtAcP, **6**) and their Ni and Zn complexes (TPyEtAcPNi, **7**; TPyEtAcPZn, **8**). Studied photophysical properties of absorption, fluorescence emission and the photochemical characteristic of singlet oxygen generation and oxygen free radicals by synthetized porphyrins are compared in dependence on the chemical structure. Finally, a comparison of the photochemical properties of the synthesized compounds and its efficiency in the photoinduced antibacterial activity is performed.

Materials and methods

Chemicals

All analytical or HPLC grade solvents were obtained from Merck (Darmstadt, Germany) and used without further purification.

Benzaldehyde, α -naftaldehyde, chloro ethyl acetate, pyrrole, propionic acid, tetra-pyridyl porphyrin, chloroform, histidine, Rose Bengal, horseradish peroxidase (HRP), nicotinamide adenine dinucleotide phosphate reduced form (NADPH), and *p*-nitrosodimethyl aniline were purchased from Sigma–Aldrich (St. Louis, MO, USA). 3-Aminophthalhydrazide (Luminol) and hydrogen peroxide, 30 wt.% solution in water, were purchased from Aldrich (Milwaukee, USA). Phosphate buffered saline solution (PBS) pH 7.4 (0.01 mol l⁻¹ phosphate buffer and 0.135 mol l⁻¹ NaCl) was prepared daily before each experiment.

¹H NMR and ¹³C NMR spectra were recorded with a Brucker Avance 500 and 300 MHz respectively, in chloroform-*d* with tetramethylsilane (Me₄Si) as an internal standard. Chemical shifts (δ) are given in parts per million. Infrared spectroscopy (IR) spectra were performed using a Nicolet Magna 560 FT-IR spectrometer. ElectroSpray Ionization mass spectra and MS/MS spectra were obtained with a Thermo-Finnigan TSQ Quantum Ultra AM spectrometer coupled to a HPLC Electrospray (ESI). Elemental analyses were performed in a Fisions Instrument EA-1108. The samples were prepared by addition of the compound of interest to water. The absorption spectra were recorded on a Perkin Elmer Lambda-35 UV–Vis spectrophotometer. The fluorescence spectra and the quantum yields were registered in a Perkin Elmer LS-45.

Synthesis of the porphyrins derivatives

Tetraphenyl porphyrins (TPP)

The synthesis of TPP (Scheme 1) was carried out following the experimental procedure reported by Alder and coworkers [15] with some modifications: benzaldehyde (0.67 mol) and pyrrole (0.65 mol) were added simultaneously to refluxing propionic acid (200 ml) and the mixture was refluxed for 1 h before being allowed to cool and stand at room temperature overnight. The product was filtered off and washed with water and methanol to give purple crystals. Yield 20% (20.5 g). **F.T-I.R** (KBr, thin film) v_{max} (cm⁻¹): 3309 (N-H); 3047 (C=C); 2361 (C=N); 1593, 1466 (aromatic C=C): 698 (C-H). UV-vis $(1.0 \times 10^{-5} \text{ mol } l^{-1} \text{ in CHCl}_3)$, λ_{max} (nm): 419, 515, 550, 590, 674; ¹H NMR (500 MHz, CDCl₃) δ = ppm: 8.87 (s, 8H, H-pyrrolic), 8.21–8.28 (d, J = 5.0 Hz, 8H, ophenyl), 7.72–7.85 (m, 12H, H-phenyl), –2.73 (s, NH). ¹³C NMR $(125 \text{ MHz}, \text{ CHCl}_3) \delta = \text{ppm: } 142.21 (2C-\text{phenyl}), 134.6 (CH-o-\text{phe-})$ nyl), 131.15 (CH-pyrrolic), 127.74 (2C-pyrrolic), 126.72 (2CH-phenyl), 120.18 (2C).

Tetranaphthylporphirins (TNP)

The synthesis of TNP (Scheme 2) was carried out following the experimental procedure reported by Regimol and coworkers [16] with some modifications: benzaldehyde (0.10 mol) and pyrrole (0.14 mol) were added simultaneously to refluxing propionic acid (100 ml). The reaction mixture was left at reflux for a ten days, after which it was allowed to cool to room temperature, and then it was kept at 5 °C overnight. The product was filtered off and washed with water and methanol and the solid obtained was purified by preparative chromatography on silica plates, using chloroform as mobile phase, yielding purple crystals. Yield of 19.8% (5.44 g). **F.T-I.R** (KBr, thin film) v_{max} (cm⁻¹): 3432 (N–H), 2357 (C=N), 3048 (C=C), 1568, 1499 (aromatic C=C), 1021 (C-H). **UV–vis** $(1.0 \times 10^{-5} \text{ mol } l^{-1} \text{ in CHCl}_3)$, λ_{max} (nm): 423. ¹H NMR (500 MHz, CDCl₃) δ = ppm: 8.47 (d, 8H, J = 4.5 Hz, H-pyrrolic), 8.28-7.73 (m, 28H, H-naphthalene), -2.76 (s, 2H, NH). MS (APCI) m/z: 815.22 (M + H)⁺, (cal: M:C₆₀H₃₈N₄ = 814.97).

Tetrapiridyl ethylacetate porphirins

(TPyEtAcP, **6**): The synthesis of water-soluble porphyrin TPyEtAcP (Scheme 3) was performed according to changes in the methodology reported by Berezin and coworkers [17]: tetrapiridyl Download English Version:

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