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Enhanced bio-hydrogenesis by co-culturing photosynthetic bacteria with acidogenic process: Augmented dark-photo fermentative hybrid system to regulate volatile fatty acid inhibition

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ARTICLE INFO

Article history: Received 21 April 2013 Received in revised form 23 January 2014 Accepted 28 January 2014 Available online 25 February 2014

Keywords: Volatile fatty acid (VFA) Bacteriochlorophyll Pheophytinization Acetic acid

ABSTRACT

To overcome induced fatty acid inhibition during dark-fermentative hydrogen (H₂) production process, a hybrid strategy was designed and evaluated by co-culturing photosynthetic bacteria with acidogenic microflora. Augmented dark-photo fermentation system (ADPFS) illustrated 40% increment in cumulative H₂ production (CHP, 250 ml) compared to dark-fermentation system (DFS) along with 10% enhancement in COD removal efficiency. Co-culturing helped to reduce VFA accumulation by 40% which supports the functional role of photosynthetic organisms in reducing the fatty acid concentration in association to additional H₂ production. Relatively higher reduction in individual fatty acids viz., acetic acid (43%), butyric acid (57%) and propionic acid (65%) was observed with AD-PFS operation. Increment in bacteriochlorophyll (Bchl) after augmentation corroborated well with results. At lower pH, pheophytinization was observed which hindered H₂ production. Voltammograms illustrated dominant oxidation behavior during hybrid AD-PFS operation and provides viable option for enhancing performance by regulating system buffering microenvironment.

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Introduction

Diversification of energy resources is an essential requirement in the present-day energy scenario [1]. Rapid development of alternative, renewable, carbon-neutral, and eco-friendly fuels is essential to fulfill the burgeoning energy demands. Hydrogen (H₂) gas is an important and promising energy carrier that could play a significant role in the reduction of greenhouse gas emissions. Biologically produced H₂ is a natural and transitory by-product of various microbial-driven biochemical reactions [2–4]. Several strategies have been employed for biological H₂ production which include, direct and indirect biophotolysis of water, photo-fermentation,

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dark-fermentation or hybrid (photo-dark) fermentation. H_2 generation in light dependent (photo-fermentation) and lightindependent (dark-fermentation) processes is of practical importance as it can combine both H_2 production and elimination of organic waste materials in a single step [5,6].

The dark-fermentation of H₂ production generates organic acid metabolites which inhibits the H₂ production process. The generation of these organic acids is one of the major problems as they make the H₂ production process unfavorable by limiting the substrate degradation. Various integration strategies were reported for the utilization of acid-rich effluents as the primary substrate for energy recovery e.g., anaerobic dark fermentation for H2 integrated with the methanogenesis [7], photo fermentation [8-10], bioelectricity through microbial fuel cell [11] and recovery of value added products like bio-plastics [12] at secondary stage. Further utilization of the organic acids towards H₂ production is thermodynamically feasible only if there is an additional energy input. This energy input can be in from of electricity in microbial electrolysis cell (MEC) [13] or in the form of light in 2 stage photofermentation where it allows maximum conversion of the organic carbon to H₂ [14]. Diverse group of photosynthetic bacteria (PSB) are capable of utilizing organic acids as carbon and light as energy sources for H₂ production. Reports are available with two stage process for integrating heterotrophic dark-fermentation with photo-heterotrophic/ fermentation processes for additional H₂ production [15,16]. Combination of dark and photo-fermentation could achieve a theoretical maximum yield of 12 mol H₂/mol Hexose [2,3]. A two-stage process i.e., the integration of dark and photofermentation has been considered as an effective and efficient system to increase H₂ yield and enhance energy recovery from organic wastewater and lower chemical oxygen demand (COD) in the process effluents [8].

In this communication, we have made an attempt to use hybrid process by integrating dark and photo fermentative processes in a single system for enhancing H_2 production along with wastewater treatment. The hybrid strategy facilitates in situ utilization of metabolic intermediates formed during the acidogenic H_2 production simultaneously by coculturing photosynthetic bacteria. Experiments were designed for evaluating the relative performance of darkfermentative process and photosynthetic-dark fermentative hybrid process on H_2 production and substrate degradation at two organic loads (OL) i.e., 1 kg COD/m³-day (OL1) and 1.6 kg COD/m³-day (OL2).

Materials and methods

Biocatalyst

Anaerobic culture

Anaerobic consortia acquired from a full scale operating anaerobic treatment unit was used as dark fermentative inoculum in the experiments. It was initially enriched in designed synthetic wastewater (DSW) [NH₄Cl - 0.5, KH₂PO₄ - 0.25, K₂HPO₄ - 0.25, MgCl₂.6H₂O - 0.3, FeCl₃ - 0.025, NiCl₄ - 0.016, CoCl₂ - 0.025, ZnCl₂ - 0.0115, CuCl₂ - 0.0105, CaCl₂ - 0.005, MnCl₂ - 0.015, C₆H₁₂O₆-3.00 (g/l)] for a period of 72 h comprising 3 cycles each with 24 h under anaerobic microenvironment at pH 6.0 (100 rpm; 48 h). After enrichment of the inoculum it was subjected to sequential pretreatment with chemical, heat-shock and acid-shock to enrich H₂ producers (hydrogenic bacteria) as well as to suppress methanogenic bacteria (MB) [8].

Photosynthetic culture

An indigenous mixed photosynthetic consortium was acquired from existing photosynthetic fuel cell (PhFC) reported in our previous experiments [9]. This culture was enriched in a succinate salt broth, consisting of $KH_2PO_4 - 0.33$ g, MgSO₄.7H₂O- 0.33 g, NaCl- 0.33 g, NH₄Cl- 0.5 g, CaCl₂.2H₂O-0.05 g, sodium succinate- 1.0 g, Yeast extract- 0.02 g, Distilled H₂O- 1 L, 1 ml trace metal solution (ZnSO₄.7H₂O- 10 mg, MnCl₂.4H₂O- 3 mg, H₃BO₃- 30 mg, CoCl₂.6H₂O- 20 mg, CuCl₂.2H₂O- 1 mg, NiCl₂.6H₂O- 2 mg, Na₂MoO₄- 3 mg, Distilled H₂O- 1.0 L, pH 3–4) and 0.02% FeSO₄.7H₂O solution- 0.5 ml. This composition works well for enrichment of

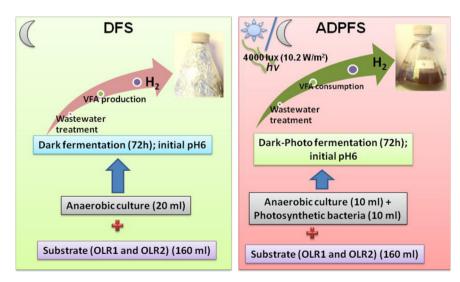


Fig. 1 – Schematic representation of experimental methodology followed during the operation dark-fermentation system (DFS) and augmented dark-photo fermentative system (ADPFS).

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