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An "off–on" optical sensor for mercury ion detection in aqueous solution and living cells



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ABSTRACT

A rhodamine-based optical sensor Rh–3S has been developed for selective detection of Hg^{2+} in aqueous solution as well as in living cells. UV–vis and fluorescence spectroscopic studies reveal that Rh–3S shows marked sensitivity and selectivity to Hg^{2+} with a chelation-induced "off–on" response in acetonitrile (ACN)/MOPS buffer (10 mM, pH 7.3, v/v, 1:1). The Rh–3S sensor binds Hg^{2+} in a 1:1 stoichiometry with an apparent binding constant $3.71 \times 10^6 M^{-1}$ (log K = 6.57) and displays a distinct change in color and fluorescence upon the alteration of free Hg^{2+} levels in solution with a reversible response and little interference with other biological relevant metal ions. Live cell confocal imaging studies demonstrate that the sensor is also capable of imaging the presence of Hg^{2+} ions as well as its dynamic changes in live cells.

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As one of most toxic metals, mercury is widely distributed in water, air and soil, generated from both natural and artificial sources such as oceanic and volcanic eruption, mining, waste incineration and the combustion of fossil fuels, posing serious problems to human health [1]. Mercuric ion (Hg^{2+}) , the ionic mercury form, shows high affinities to thiol groups, various enzymes and amino acids, leading to the dysfunction of these biomolecules and subsequently causing serious irreversible damage to the organs in cardiovascular, endocrine and central nervous systems [1-3]. It is believed that ionic mercury and its organometallic derivatives play important roles in the pathology of neurodegenerative diseases such as Amvotrophic lateral sclerosis. Parkinson's disease, and Alzheimer's disease [1-3]. Thus, there has been a tremendous interest in developing rapid, efficient and high-throughput methods for monitoring the level of Hg²⁺ in vivo and in vitro. Compared with those costly and time-consuming methods such as atomic absorption spectroscopy, inductively coupled plasma mass spectrometry (ICP-MS), and inductively coupled plasma atomic emission spectrometry (ICP-AES), optical chemosensors are greatly preferred due to its advantages in simple sample pretreatment, low need for expensive instrument as well as in-time and on-site responses.

In recent years, considerable efforts have been devoted to the development of fluorescent and colorimetric sensors for detecting Hg^{2+} ions [4,5]. Rhodamine-based sensors with "off–on" response have attracted

* Corresponding author at: Department of Chemistry and Biochemistry, UMass Cranberry Health Research Center, University of Massachusetts Dartmouth, 285 Old Westport Road, Dartmouth, MA 02747, USA. much attention in chemosensor development due to its striking spectroscopic properties such as large molar extinction coefficients, high quantum yield and long excitation and emission wavelength suitable for live cell imaging [6–16]. However, only a few Hg^{2+} sensors have demonstrated their capability in live cell imaging [9,11,12]. It is still highly



Fig. 1. Titration of 20 μ M Rh–3S with increasing concentrations of HgCl₂ (2, 4, 6, 10, 12, 14, 16, 18, 20, 28, 36, 44, 52, 60 and 80 μ M, respectively) in ACN/MOPS buffer (10 mM, pH 7.3, v/v 2:1). The absorption intensities were measured at 565 nm.

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Fig. 2. (a) Job's plot of Rh–3S and Hg²⁺ in the ACN/MOPS buffer (10 mM, pH 7.3, v/v 1:1). The absorbance was collected at 565 nm. (b) ESI-MS spectrum of the solution of Rh–3S with Hg²⁺ (1 µM each in ACN).



Scheme 1. Proposed reversible 1:1 binding mode between Rh-3S and Hg²⁺.

desirable to develop Hg^{2+} sensors with good aqueous solubility and potential applications in living cell imaging [17]. We have been devoted to designing optical sensors for detecting metal ions such as iron and copper as well reactive oxygen species (ROS) [18–22]. Herein, we have developed a novel optical chemosensor for mercury, Rh–3S, which incorporates a rhodamine fluorophore with an Hg^{2+} -binding receptor moiety composed of one nitrogen and three sulfur atoms by virtue of both the high thiophilicity of Hg^{2+} and strong nucleophilicity of sulfur.

Rh–3S was synthesized in two steps (Scheme S1) by refluxing rhodamine B hydrazine [23] and 5-bromo-2,2'-bithiophene-5'-carboxaldehyde in ethanol, followed by the thionation of carbonyl group of the product obtained in the first step [24]. The detailed synthetic procedures and characterization are described in the Supporting information.

First, we evaluated the optical properties of Rh–3S and its interactions with Hg^{2+} in a mixture solvent of acetonitrile (ACN)/3-(N-morpholino)propanesulfonic acid (MOPS) buffer (10 mM, pH 7.3, v/v, 1:1) (Fig. 1). Rh–3S (20 μ M) in ACN/MOPS buffer solution is yellowish with only very weak absorption over 500 nm, indicating that the

rhodamine moiety was dominant in the spirolactam form. When Hg²⁺ was added to the solution, the solution instantaneously turned pink with an absorption band that appeared at 565 nm and increased in intensity with the gradual addition of Hg²⁺, suggesting the formation of ring-opened rhodamine species [18–20]. This rapid response to mercury is very important in practical application for real-time detection. The titration curve (a plot of Rh–3S versus Hg²⁺ concentration) increased linearly and plateaued at 1:1 ratio of the sensor and Hg^{2+} , suggesting the formation of a 1:1 Hg²⁺/Rh–3S complex. The binding constant of this complex was determined to be $3.71 \times 10^{6} \text{ M}^{-1}$ (log K = 6.57) by following a method reported previously [25], using absorption values at 565 nm (the equations were described in the Supplementary material). The stoichiometry of Rh–3S–Hg²⁺ complex was further confirmed by Job's method (Fig. 2a) and is supported by mass spectrometric analysis. Job's plot, using continuous variation with a total concentration of 20 µM Rh–3S and Hg²⁺, exhibited a maximum absorbance (at 565 nm) when the molecular fraction of Hg^{2+} and Rh-3S was close to 1:1, suggesting a 1:1 binding ratio of Rh-3S



Fig. 3. (a) Absorption responses of 20 μ M Rh–3S to various metal ions (20 μ M for Zn²⁺, Ct³⁺, Ni²⁺, Hg²⁺, Fe³⁺, Mn²⁺, Ag⁺, Pb²⁺, Fe²⁺, Cu⁺, Cu²⁺, and Co²⁺; 100 μ M for Na⁺, K⁺, Mg²⁺, and Ca²⁺) in ACN/MOPS buffer (10 mM, pH 7.3, v/v 1:1). (b) Fluorescence response ($\lambda_{Ex}/\lambda_{Em}$ 510/580 nm, filter 515 nm) of 20 μ M Rh–3S to various metal ions (20 μ M for Ni²⁺, Cu⁺, Cu²⁺, Zn²⁺, Pb²⁺, Fe²⁺, Fe³⁺ Cr³⁺, Hg²⁺, Mn²⁺, Ag⁺, and Co²⁺; 100 μ M for K⁺, Na⁺, Ca²⁺ and Mg²⁺) in the ACN/MOPS buffer (10 mM, pH 7.3, v/v 1:1).

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