



# Delivery of porphrin to cancer cells by organometallic Rh(III) and Ir(III) metalla-cages



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## ABSTRACT

Two pentamethylcyclopentadienyl (Cp\*) rhodium(III) and iridium(III) metalla-prisms of the general formula  $[(Cp^*M)_6(tpt)_2(dhnq)_3]^{6+}$  ( $tpt = 2,4,6$ -tri-(pyridin-4-yl)-1,3,5-triazine,  $dhnq = 5,8$ -dihydroxy-1,4-naphthoquinonato;  $M = Rh$ ,  $[1]^{6+}$ ;  $M = Ir$ ,  $[2]^{6+}$ ) have been synthesized by mixing in methanol the neutral dinuclear complexes  $(Cp^*M)_2(dhnq)Cl_2$  ( $M = Rh, Ir$ ),  $AgCF_3SO_3$  and  $tpt$ . In a similar fashion, the addition of porphrin during the formation of  $[1]^{6+}$  and  $[2]^{6+}$  leads to the carceplex systems  $[porphin \subset 1]^{6+}$  and  $[porphin \subset 2]^{6+}$ . All complexes were isolated as their triflate salts and characterized by different analytical techniques including for the carceplex  $[porphin \subset 1][CF_3SO_3]_6$  by a single-crystal X-ray structure analysis. The hosts were evaluated as drug delivery vectors on the HT-29 cancer cell line, and after internalization, porphrin was photo-activated to generate phototoxicity. Interestingly, the  $IC_{50}$  values of the empty and filled metalla-cages are around  $1 \mu M$ , while upon irradiation ( $20 J/cm^2$ ) only lower nanomolar concentrations of the  $[porphin \subset cage][CF_3SO_3]_6$  systems are necessary to inhibit cell growth by 50%, showing an excellent ratio between cytotoxicity and phototoxicity.

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## Introduction

In recent years, the number of biological studies involving arene ruthenium metalla-assemblies is booming [1]. So far, trinuclear [2], tetranuclear [3], hexanuclear [4], octanuclear [5] and dodecanuclear [6] metalla-assemblies incorporating arene ruthenium(II) complexes have been synthesized and often they have been tested as anticancer agents. In general, these cationic complexes have showed an *in vitro* anticancer activity in the lower micromolar range. Interestingly, some of the hexanuclear and octanuclear arene ruthenium metalla-assemblies possess a hydrophobic cavity which can be filled with a guest molecule [7], and consequently, these cationic water soluble metalla-hosts have been used as drug delivery vectors [8]. Despite the huge popularity of arene ruthenium metalla-assemblies, the biological side of their pentamethylcyclopentadienyl (Cp\*) rhodium(III) and iridium(III) analogues remains unexplored. In fact, several Cp\*Rh and Cp\*Ir metalla-

assemblies can be found in the literature [9], however, only a handful of these metalla-assemblies has been biologically evaluated. Among them, the embelin-derived tetranuclear [10] and hexanuclear complexes [11], and the 5,8-dihydroxy-1,4-naphthoquinonato tetranuclear metalla-cycles with bipyridyl linkers [12]. On the other hand, mononuclear and dinuclear Cp\*Rh and Cp\*Ir complexes have showed great promises as anticancer agents [13], with even in some cases possessing an excellent selectivity for cancerous over noncancerous cell lines [10].

Nevertheless, to obtain temporal and spatial-controlled activation of a drug for an optimal selectivity, and accordingly to alleviate side effects associated to non-specific accumulations, the use of an external stimulus to activate a drug to treat cancers is quite attractive [14]. Among these stimuli, light is commonly employed to activate biologically relevant compounds [15]. The utilization of light for therapeutic purposes is nowadays called photodynamic therapy (PDT), but the concept of using light to cure diseases has been exploited under different terminologies for centuries [16]. Recently, we have inserted a photosensitizer in the hydrophobic cavity of arene ruthenium metalla-assemblies [17]: The metalla-assembly was protecting, transporting and shielding the

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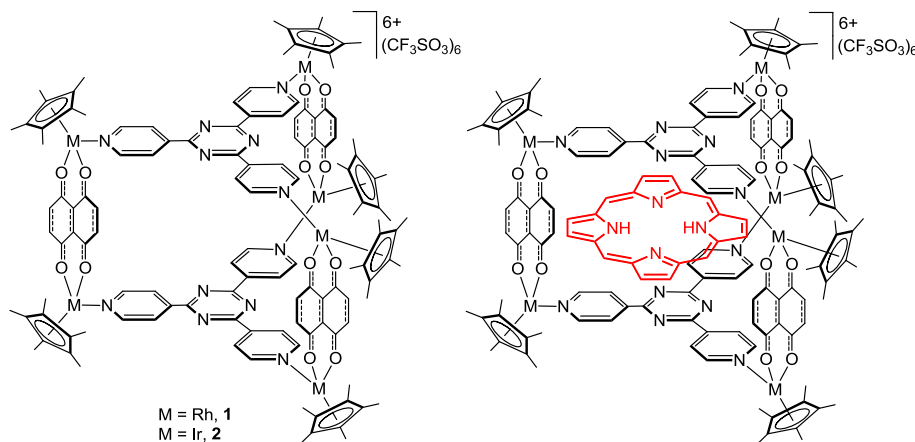


Fig. 1. Molecular structures of **[1]<sup>6+</sup>**, **[2]<sup>6+</sup>**, **[porphin<1><sup>6+</sup>** and **[porphin<2><sup>6+</sup>**.

photosensitizer until it was delivered to its target. Then following the uptake of the host-guest system by cells, the photosensitizer was released and photo-activated, providing a new strategy to deliver hydrophobic photosensitizers to cancer cells [18].

Herein we report the synthesis and characterization of the 5,8-dihydroxy-1,4-naphthoquinonato (dhmq) Cp\**Rh* and Cp\**Ir* metalla-prisms of the general formula [(Cp\**M*)<sub>6</sub>(tpt)<sub>2</sub>(dhmq)<sub>3</sub>]<sup>6+</sup> (tpt = 2,4,6-tri-(pyridine-4-yl)-1,3,5-triazine), and the corresponding complexes in which porphyrin sits inside the cavity. The anti-proliferative activity of the complexes and their photodynamic activity were evaluated on the human colon cancer cell line HT-29.

## Results and discussion

The reaction of the dinuclear pentamethylcyclopentadienyl complexes (Cp\**M*)<sub>2</sub>(dhmq)Cl<sub>2</sub> (M = *Rh*, *Ir*) with AgCF<sub>3</sub>SO<sub>3</sub> followed by the addition of tpt (2,4,6-tri-(pyridine-4-yl)-1,3,5-triazine) affords the cationic hexanuclear metalla-prisms of the general

formula [(Cp\**M*)<sub>6</sub>(tpt)<sub>2</sub>(dhmq)<sub>3</sub>]<sup>6+</sup> (dhmq = 5,8-dihydroxy-1,4-naphthoquinonato; M = *Rh*, **[1]<sup>6+</sup>**; M = *Ir*, **[2]<sup>6+</sup>**), see Fig. 1. However, the synthesis of the carceplexes **[porphin<1><sup>6+</sup>** and **[porphin<2><sup>6+</sup>** is achieved by the addition of porphyrin during the formation of the metalla-prisms **[1]<sup>6+</sup>** and **[2]<sup>6+</sup>**. The empty and filled metalla-prisms are equally obtained in moderate yields as their triflate salts, suggesting no template effect from porphyrin. The metalla-prisms and the host-guest systems were fully characterized by infrared and NMR spectroscopies, ESI mass spectrometry and for **[porphin<1>][CF<sub>3</sub>SO<sub>3</sub>]<sub>6</sub>** by a single-crystal X-ray structure analysis. All complexes are non-hygroscopic and soluble in common polar solvents and insoluble in non-polar solvents. In addition, all the metalla-prisms are moderately soluble in water.

The formation of these complexes was easily monitored by <sup>1</sup>H NMR spectroscopy (Fig. 2). The complexes **[1]<sup>6+</sup>** and **[2]<sup>6+</sup>** show well-defined structures with relatively simple sets of signals in their <sup>1</sup>H NMR spectra, displaying the expected resonances for the metalla-prisms. Yet, in the host-guest systems **[porphin<1><sup>6+</sup>** and

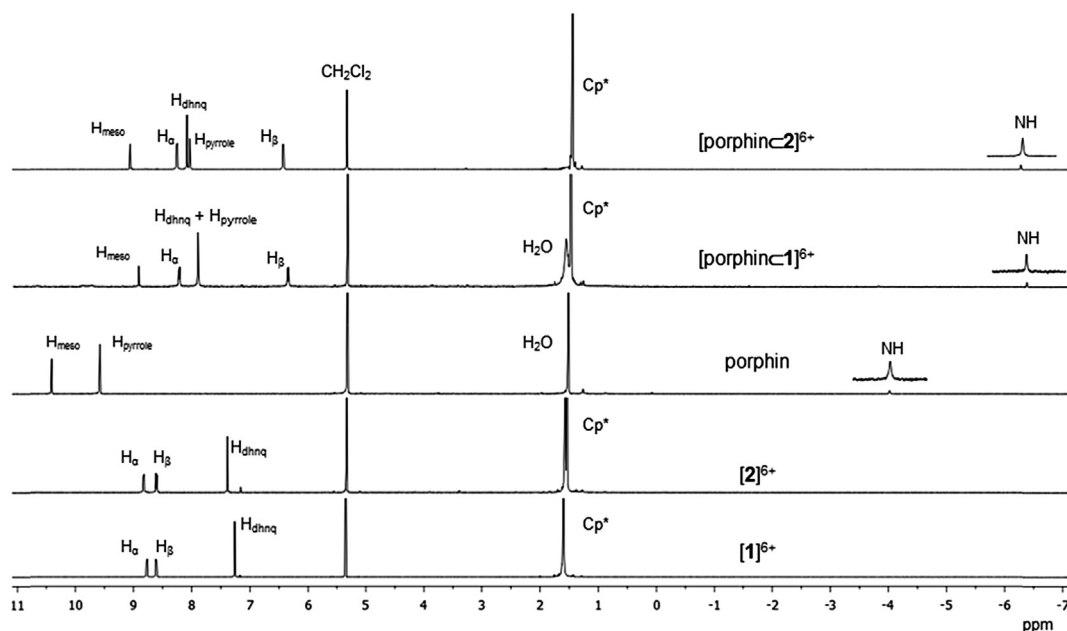


Fig. 2. <sup>1</sup>H NMR spectra (21 °C, CD<sub>2</sub>Cl<sub>2</sub>) of **[1]<sup>6+</sup>**, **[2]<sup>6+</sup>**, porphyrin, **[porphin<1><sup>6+</sup>** and **[porphin<2><sup>6+</sup>**.

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