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Total synthesis of (+)-cladospolide D

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ABSTRACT

Article history: Received 21 January 2016 Accepted 25 January 2016 Available online 10 February 2016 The asymmetric total synthesis of cladospolide D, a natural 12-membered macrolide antibiotic, has been accomplished in 12-steps. Sharpless asymmetric dihydroxylation, Grubb's cross metathesis, and Shiina lactonization have been employed as the key reactions for the installation of the desired stereocenter and the construction of the macrolactone framework.

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1. Introduction

In 2001, Omura et al. have isolated a new 12-membered γ -oxo-lactone, namely cladospolide D **1**, along with two known cladospolides A 2 and B 3 from the fermentation broth of Cladosporium sp. FT-0012.¹ Species belonging to the genus Cladosporium have also produced a family of other fungal secondary metabolites, cladospolides A-C 2-4 and iso-cladospolide B 5, which are plantgrowth regulators (Fig. 1).² Unlike these members, cladospolide D exhibited antimicrobial properties with IC_{50} values of 0.1 and 29 µg/mL against Mucor recemosus and Pyricularia oryzae, respectively. Although, the initial structural determination of cladospolide D was carried out during the isolation, the absolute and relative stereochemistry as well the stereochemical assignment of the C2-C3 double bond were correctly determined only after studies on its total synthesis independently by Hou et al.³ and O'Doherty et al.⁴ Later, Kaliappan and Si et al. also achieved the total synthesis of naturally occurring (+)-cladospolide D.⁵ In continuation of our research program on the total synthesis of bioactive natural macrolides,⁶ in particular the cladospolides,⁷ we herein report the asymmetric total synthesis of (+)-cladospolide D 1.

A retrosynthetic analysis suggests TBS-protected alkenol **7** could be a suitable starting material. Cladospolide D **1** could be prepared by Shiina lactonization of hydroxy acid **6**, which in turn could be obtained from alkenol **7** involving Sharpless asymmetric dihydroxylation for the installation of the C5-hydroxyl group and Grubbs olefin metathesis to obtain the desired C2–C3 olefin functionality of the target molecule. The above reaction sequence has not been utilized previously for the synthesis of cladospolide D (Scheme 1).

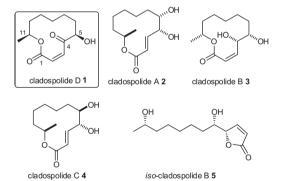
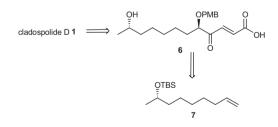


Figure 1. Structures of cladospolides A-D and iso-cladospolide.



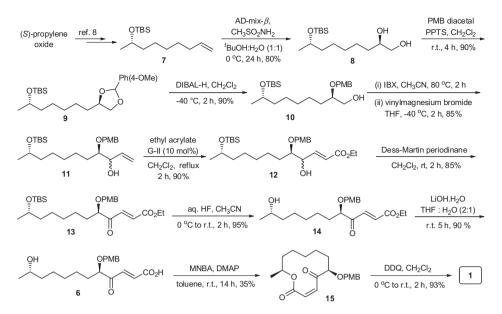
Scheme 1. Retrosynthetic analysis of cladospolide D 1.

2. Results and discussion

As shown in Scheme 2, the synthesis of cladospolide D **1** began from the known alkenyl *tert*-butyldimethylsilyl ether **7**, which was obtained readily in two-steps from (*S*)-propylene oxide following



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Scheme 2. Synthesis of (+)-cladospolide D 1.

the literature.⁸ Sharpless asymmetric dihydroxylation⁹ of alkene **7** with AD-mix- β , CH₃SO₂NH₂ and tBuOH/H₂O (1:1) gave diol **8** in 80% yield as a separable diastereomeric mixture (95:5). Diol 8 was protected as its para-methoxybenzylidene acetal 9 in 90% vield, which was subsequently subjected to reductive opening under DIBAL-H in CH₂Cl₂ at -78 °C to provide the primary alcohol 10 in 81% yield. Primary alcohol 10 was oxidized using IBX in CH₃-CN to give the aldehyde, which upon treatment with vinylmagnesium bromide afforded allylic alcohol 11 as a diastereomeric mixture. To obtain the desired α , β -unsaturated ester, allylic alcohol 11 was subjected to a cross-metathesis reaction with ethyl acrylate using Grubb's second-generation catalyst (G-II),¹⁰ which provided 12 in 90% yield. The hydroxyl group of 12 was oxidized under Dess-Martin periodinane conditions to yield keto-ester 13 in 85% yield. Toward macrolactonization, deprotection of tert-butyldimethyl silyl (TBS) was carried out under aq HF in CH₃CN to obtain hydroxy keto-ester 14 in 95% yield. Hydrolysis of the ester group of **14** using LiOH in THF/H₂O gave the hydroxyl acid **6** (90%), a key precursor for macrolactonization. Treatment of 6 under Shiina lactonization¹¹ with 2-methyl-6-nitrobenzoic anhydride (MNBA) in the presence of DMAP in toluene provided keto-lactone 15 in 35% yield. Finally, DDQ-mediated deprotection of the para-methoxy benzyl group of 15 provided the target molecule, cladospolide D, in 93% yield. The spectroscopic data (IR, ¹H and ¹³C NMR) of **1** were identical and the specific rotation observed for 1, $[\alpha]_D^{20}$ = +55.5 (c 0.8, CH₃OH), is comparable with the reported data {lit: $[\alpha]_{\rm D} = +56.0 \ (c \ 1.00, \ {\rm CH}_3 {\rm OH}) \}.$

3. Conclusion

In conclusion, the chemistry described herein defines an asymmetric approach for the construction of natural cladospolide D based on a Shiina lactonization strategy involving Sharpless asymmetric dihydroxylation and Grubb's olefin cross metathesis reactions as key steps.

4. Experimental

4.1. General

All reagents and solvents were of reagent grade and used without further purification unless otherwise stated. All of the reactions were performed under N₂ in flame or oven dried glassware with magnetic stirring. Reactions were monitored by thin-layer chromatography carried out on silica plates (silica gel 60 F254, Merck) using UV-light and anisaldehyde or potassium permanganate or β-naphthol for visualization. Column chromatography was performed on silica gel (60-120 mesh) using hexanes and ethyl acetate as eluents. Evaporation of the solvents was conducted under reduced pressure at temperatures less than 45 °C. IR spectra were recorded on Perkin-Elmer 683, Nicolet Nexus 670 spectrometer. ¹H and ¹³C NMR spectra were recorded in CDCl₃ solvent on a 300 MHz, 500 MHz and 600 MHz NMR spectrometer. Chemical shifts δ and coupling constants I are given in ppm (parts per million) and Hz (hertz) respectively. Chemical shifts are reported relative to a residual solvent as an internal standard for ¹H and ¹³C (CDCl₃: δ 7.26 ppm for ¹H and 77.0 ppm for ¹³C). Mass spectra were obtained on Finnigan MAT1020B, micromass VG 70-70H or LC/MSD trapSL spectrometer operating at 70 eV using direct inlet system. Optical rotations were measured on an Anton Paar MLP 200 modular circular digital polarimeter by using a 2-mL cell with a path length of 1 dm.

4.1.1. (2R,8S)-8-((tert-Butyldimethylsilyl)oxy)nonane-1,2-diol 8

To a 500 mL round bottom flask, were added 85 mL of ^tBuOH, 85 mL of H_2O and AD-mix- β (27.3 g, 1.4 g/mmol) and methane sulfonamide (2.6 g, 19.5 mmol). The mixture was stirred at room temperature for 5 min, then cooled to 0 °C. To this cooled solution, was added compound 7 (5 g, 19.5 mmol) and stirred for 24 h at 0 °C. The reaction was guenched with saturated sodium sulfite at room temperature. The mixture was diluted with EtOAc (50 mL) and after separation of the layers, the aqueous layer was further extracted with EtOAc (2×50 mL). The combined organic layers were washed with brine solution (40 mL) and dried over Na₂SO₄. The crude mixture was purified by column chromatography (silica gel, $20\% \rightarrow 30\%$ EtOAc in hexanes) to give compound 8 (4.37 g, 80% yield) as a clear colorless oil. $[\alpha]_{D}^{25} = +4.4$ (c 0.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 3.84–3.51 (m, 3H), 3.37 (dt, J = 37.7, 18.8 Hz, 1H), 2.27–2.03 (m, 2H), 1.59–1.14 (m, 10H), 1.07 (d, J = 6.1 Hz, 3H), 0.84 (s, 9H), 0.00 (s, 3H), -0.00 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 72.3, 68.64, 66.9, 39.6, 33.2, 29.7, 25.9, 25.6, 23.8, 21.9, 18.2, -4.4, -4.7; IR (KBr): 3365, 2932, 2858, 1464, 1374, 1253, 1134, 1066, 835, 774 cm⁻¹; MS (ESI): m/z 313 (M+Na)⁺; HRMS (ESI): m/z calcd for C₁₅H₃₄NaO₃Si (M+Na)⁺: 313.2168, found: 313.2181.

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