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Iridoids, flavonoids, and monoterpene diglycoside from the roots of *Triosteum himalayanum* Wall.



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ABSTRACT

The phytochemical investigation of the roots of *Triosteum himalayanum* Wall. led to the isolation of eleven iridoids (1-6 and 8-12), one monoterpene diglycoside (7), and four flavonoids (13-16), of which 4',6'-di-O-isopropylidene sweroside (1) and secologanin diethylacetal (2) were new iridoid glucosides. The existence of iridoids and flavonoids provided further confirmation of the typical profile of the secondary metabolites found in the genus *Triosteum*, suggesting a close relationship among the species of Caprifoliaceae.

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1. Subject and source

The genus *Triosteum* Linn. (Caprifoliaceae) consists of about eight species mainly distributed in East Asia and North America (Hsu et al., 1988), with 3 species in China (Xu et al., 1988). *Triosteum himalayanum* Wall., has been used in Chinese folk medicine for inducing diuresis and promoting blood circulation (Editorial Committee of the Administration Bureau of Traditional Chinese Medicine, 1999). The roots of *T. himalayanum* Wall. were collected from Tanchang County, Gansu Province of China in August 2005, and authenticated by Prof. Guo-Liang Zhang, the College of Life Science, Lanzhou University. A voucher specimen (No. 20050803) was deposited in the Institute of Organic Chemistry, Lanzhou University.

2. Previous work

Until now, only one paper has been published concerning the secondary metabolites of *T. himalayanum* Wall. indicating the presence of one known iridoids (triohima B) and two novel iridoids triohima A and C with an unusual δ -lactone-containing skeleton (Li et al., 2009).

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3. Present study

3.1. Extraction and isolation

Dried roots of *T. himalayanum* Wall. (5.1 kg) were extracted sequentially with acetone and MeOH at room temperature. The acetone extract (153.0 g) was fractioned by silica gel column chromatography (CC) using a step-gradient solvent system from 100% petroleum ether (PE) to 100% EtOAc with a 10% increment and a final MeOH wash to obtain nine fractions (Fr. AA–Fr. AI). Fr. AE was further purified by silica gel CC eluted with PE–EtOAc to yield compound **11** (30 mg) and Fr. AE-1. Fr. AE-1 was further purified by Sephadex LH-20 CC (MeOH) to afford a mixture of compounds **6** (60 mg) and **9** (60 mg). Fr. AH was subjected to macroporous absorption resin (D101) CC and eluted with H₂O, 30%, 50%, 70%, and 95% EtOH to yield six subfractions (AH1–AH6). AH2 was applied to silica gel CC using CHCl₃–MeOH (20:1–5:1) to give three subfractions (AH21–AH23). AH23 was further subjected to silica gel CC and then subjected to Sephadex LH-20 CC to afford compound **1** (55 mg). In the same manner, compounds **2** (60 mg) and **10** (65 mg) were obtained from AH5 and **3** (80 mg) was obtained from AH6. Fr. AG was separated on Sephadex LH-20 (MeOH–H₂O, 1:1) to afford compound **7** (75 mg).

The MeOH extract (249.6 g) was purified by Diaion HP-20 CC using H_2O –EtOH (0:1–1:0) to afford five fractions (Fr. BA–Fr. BE). Fr. BB was subjected to silica gel CC with EtOAc–MeOH (30:1–2:1) to give five subfractions (BB1–BB5). BB3 was separated by repeated silica gel CC with EtOAc–MeOH and then purified by Sephadex LH-20 CC with MeOH to yield compound **4** (100 mg). BB5 was subjected to silica gel MPLC with a gradient of CHCl₃–MeOH (1:0–40:1, 30:1–20:1, 10:1, 2:1, 1:1, 0:1) elution solvent to give six subfractions (BB51–BB56). BB53 was separated by silica gel CC with CHCl₃–MeOH (12:1) to afford compound **12** (12 mg). BB55 was applied to silica gel CC using CHCl₃–MeOH (12:1) to afford **5** (40 mg). BC was subjected to CC on silica gel (CHCl₃–MeOH, 20:1–0:1) to give four subfractions (BC1–BC4). Compound **8** (35 mg) was isolated from fraction BC2 by silica gel CC eluting with CHCl₃–MeOH (10:1). BC4 was subjected to Sephadex LH-20 CC, eluting with CHCl₃–MeOH (20:1, 10:1, 5:1, 3:1, 1:1), to afford compounds **13** (12 mg), **14** (12 mg), **15** (12 mg) and **16** (12 mg).

The structures of the isolated metabolites (compounds 1–16, Fig. 1) were identified on the basis of spectroscopic data (MS, ¹H NMR, ¹³C NMR and 2D NMR) and by comparison of these data with values in the literature. The compounds were identified as two new iridoid glucosides, 4',6'-di-O-isopropylidene sweroside (1), and secologanin diethylacetal (2), together with fourteen known ones, *i.e.*, sweroside (3) (Cambie et al., 1990), secologanin dimethylacetal (4) (Kakuda et al., 2000), loganin (5) (Mpondo and Garcia, 1989), naucledal (6) (Mclean and Murray, 1972), urceolide (7) (Iwagawa and Hase, 1983), vincosamide (8) (Zhang et al., 2001), 3-epinaucledal (9) (Mclean and Murray, 1972), secologanin (10) (Markham et al., 1978), isosecologaninaglycon (11) (Kinast and Tietze, 1976), 7-O-ethyl sweroside (12) (Chen et al., 2007), apigenin (13) (Sun et al., 2007), luteolin (14) (Markham et al., 1978), luteolin-7-O- β -D-glucoside (15) (Zhong et al., 2008), luteolin-4'-O- β -D-glucoside (16) (Bertrand et al., 2006).

The structure of compound **1** with molecular formula being established as $C_{19}H_{26}O_9$ by the molecular ion at m/z 399.1653 $[M + H]^+$ in the HR-ESI-MS, is consistent with the observation of 19 resonances in the ¹³C NMR spectrum. ¹H and ¹³C NMR spectra showed signals due to a vinyl group $[\delta_H$ 7.58 (1H, d, J = 2.4 Hz, H-3); δ_C 153.9 (C-3), 106.2 (C-4)], an oxymethylene $[\delta_H$ 4.47 (1H, m, H-7a) 4.34 (1H, m, H-7b); δ_C 69.8 (C-7)], and a sugar moiety $[\delta_H$ 4.75 (1H, d, J = 7.5 Hz, H-1'), 3.33 (1H, m, H-2'), 3.50 (1H, m, H-3'), 3.52 (1H, m, H-4'), 3.28 (1H, m, H-5'), 3.90 (1H, dd, J = 10.5, 5.4 Hz, H-6'a), 3.77 (1H, dd, J = 10.5, 10.2 Hz, H-6'b); δ_C 100.5 (C-1'), 75.6 (C-2'), 75.0 (C-3'), 74.8 (C-4'), 68.9 (C-5'), 63.2 (C-6')]. The above observations along with signals at δ_C 168.4 (C-11), 153.9 (C-3), and 106.2 (C-4) suggested that **1** possessed an iridoid glucoside skeleton (Suyama et al., 2013).

The comparison of the NMR data between **1** and **3** reveals a similar structure of each other except for the appearance of two methyl groups [δ_{H} 1.37 (3H, s), 1.50 (3H, s); δ_{C} 19.4, 29.5] and a quaternary C-atom (δ_{C} 100.9) in **1** from an isopropylidene unit. On the basis of these data, **1** could be deduced as a sweroside acetonide derivative (Franzyk et al., 1997). The location of the isopropylidene moiety was assigned from the HMBC correlation of isopropylidene protons [δ_{H} 1.37 (3H, s), 1.50 (3H, s)] with C-4' (δ_{C} 74.8) and C-6' (δ_{C} 63.2)]. Therefore, the structure of compound **1** was determined as 4',6'-di-O-isopropylidene sweroside (Fig. 1), and is described here for the first time.

Compound **2** was isolated as colorless oil. The molecular formula $C_{21}H_{34}O_{11}$ was determined by HR-ESI-MS (m/z 480.2441, $[M + NH_4]^+$, calcd for 480.2439). The ¹H NMR spectrum showed resonances corresponding to an iridoid glucoside at δ_H 7.42 (1H, d, J = 1.2 Hz, H-3), 5.51 (1H, d, J = 5.1 Hz, H-1), and 4.67(1H, d, J = 8.1 Hz, H-1'), as well as resonances from a vinyl group at δ_H 5.74 (1H, ddd, J = 17.1, 9.6, 8.4 Hz, H-8), 5.26 and 5.30 (H-10a and b), suggesting that **2** is a secoiridoid glucoside (Kakuda et al., 2000). The structural assignments of compound **2** were supported by comparison of the spectroscopic data of **2** with those of **4**. **2** showed almost the same NMR spectra as **4** (Table 2), except for ethoxy signals in **2** instead of methoxy signals in **4**. This is in accordance with the introduction of a diethyl acetal group at C-7, which was further verified by 2D NMR (HMBC correlations between H-1"/C-7 and C-2"; H-1"'/C-7 and C-2"; and H-2" and H-2"/C-6) to derive the structure of **2** as secologanin diethylacetal that is reported here for the first time.

Tomassini et al. (Tomassini et al., 1995) have reported that compounds like secologanin dimethylacetal (**4**) and 7-O-ethyl sweroside (**10**) can be easily formed as artifacts by the reaction of secologanin (**8**) or secologanin acid with alcoholic solvents. Similarly, **2** was proposed to be formed by reaction of secologanin and ethanol.

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