



## Gabapentin hybrid peptides and bioconjugates



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### ABSTRACT

Synthetic approaches to gabapentin bioconjugates that overcome the tendency of gabapentin to cyclize into its  $\gamma$ -lactam are studied. Gabapentin was converted by N-acylation at its N-terminus into di-, tri-, and tetrapeptides (L-Ala-Gbp, L-Val-Gbp, L-Ala-L-Phe-Gbp, Gly-L-Ala- $\beta$ -Ala-Gbp). Carboxyl-activated Boc-protected gabapentin was used to N-, O-, and S-acylate small peptides and hormones to give conjugates that could also provide prodrugs containing conformationally constrained gabapentin units.

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## 1. Introduction

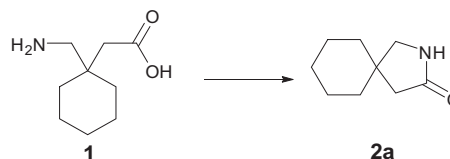
Gabapentin 1-(aminomethyl)cyclohexane acetic acid, Gbp **1**, (trade name Neurontin), a structural analog of  $\gamma$ -aminobutyric acid (GABA), is utilized for the treatment of epilepsy,<sup>1</sup> neuropathic pain,<sup>2–4</sup> restless legs syndrome,<sup>5</sup> anxiety disorders,<sup>6</sup> hot flushes<sup>7</sup> and numerous other conditions.<sup>8,9</sup> Anhydrous gabapentin exists in three polymorphic forms.<sup>10,11</sup> The mechanism of action of gabapentin remains elusive.<sup>12</sup> Apparently, it does not directly interact with GABA receptors.<sup>13–15</sup> The absorption of gabapentin and clearance of the drug depends on the dosage and the patient.

Gabapentin **1** easily undergoes facile intramolecular cyclization to form the five-membered cyclic lactam 2-aza-spiro[4,5]decan-3-one **2a** (Scheme 1). While gabapentin has low toxicity (DL<sub>50</sub>, tested in mice) at more than 8000 mg/kg, lactam **2a** has a much higher toxicity rating of 300 mg/kg.<sup>16</sup> The presence of  $\gamma$ -lactam **2a** as an impurity or the potential formation of **2a** during storage in any active ingredient must therefore be minimized both in solution and solid states.<sup>16,17</sup>

To improve Gbp bioavailability, much attention has been given to: (i) Gbp conformational properties in peptide sequences,<sup>18–25</sup> (ii) Gbp conjugated systems,<sup>26–30</sup> (iii) intermolecular interactions of Gbp,<sup>31</sup> its hydrates,<sup>32</sup> and derivatives,<sup>33</sup> (iv) Gbp multicomponent

co-crystals with various carboxylic acids,<sup>11,34</sup> and (v) the formation of N-terminus bioconjugates of gabapentin, for example, with bile<sup>29,35</sup> acid or gabapentin enacarbil **3** (Fig. 1) (Horizant, XP-13512).<sup>36,37</sup>

The preparation of the Gbp bioconjugate **3** indicated that chemical instability, insufficient oral absorption, rapid pre-systemic metabolism, and toxicity of Gbp could be overcome by formulation of Gbp prodrugs.<sup>38</sup> The development of gabapentin peptides or acylated at N-, O-, S-termini derivatives that could transform in vivo to release the active drug could thus be a feasible strategy to improve the physicochemical, biopharmaceutical and/or pharmacokinetic, and pharmacological properties of Gbp, and thereby increase its usefulness.<sup>39</sup>



**Scheme 1.** Intramolecular cyclization of gabapentin (Gbp) into 2-aza-spiro[4,5]decan-3-one **2a**.

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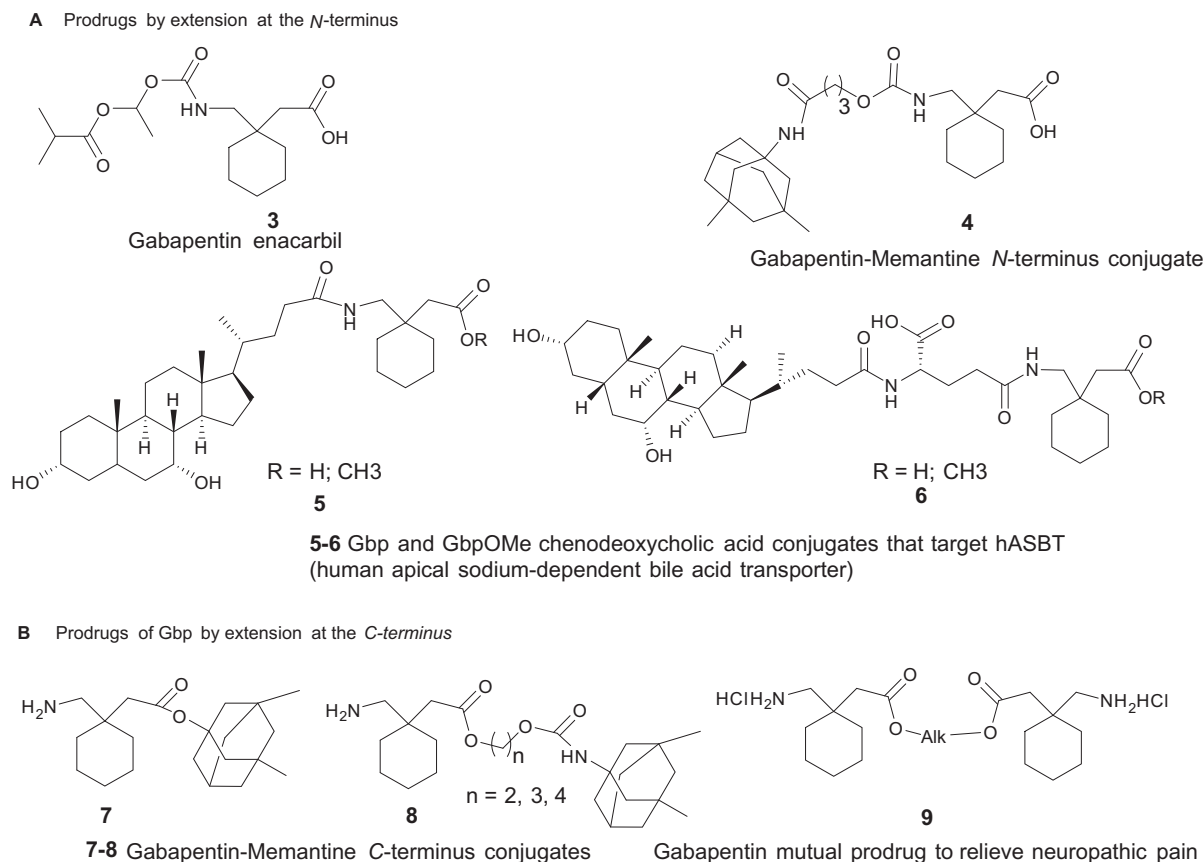


Figure 1. Reported Gbp bioconjugates.

Gabapentin enacarbil (**3**) was designed to improve physicochemical, biopharmaceutical and pharmacokinetic properties of the pharmacologically active gabapentin molecule.<sup>40</sup> The oral bioavailability of gabapentin was increased from 25% to 84% by use of the gabapentin enacarbil prodrug in monkeys, which showed dose-proportional gabapentin exposure in humans.<sup>41</sup> With these promising results in mind, we developed gabapentin conjugates that might increase bioavailability of the drug in similar ways to the gabapentin enacarbil prodrug **3**. An additional search revealed reports on both C- and N-terminus gabapentin conjugated systems (**4–9**, Fig. 1). In the development of bile acid prodrugs of Gbp the authors' strategy was to couple the Gbp drug to a natural substrate to create a molecule that mimics the three-dimensional structure of natural human (apical) sodium-dependent bile acid transporter.<sup>29</sup> Thus, the synthesis of glu-chenodeoxycholic acid and chenodeoxycholic acid Gbp and its methyl ester conjugates **5** and **6** helped overcome the limitations that gabapentin faces in its zwitterionic form and improve gabapentin oral bioavailability. For the purpose of treating neurological diseases, novel Gbp-adamantine (e.g., memantine) combinatorial N-terminus (**4**) and C-terminus (**7** and **8**) compositions were reported.<sup>30</sup> Gabapentin hydrochloride dimeric esters linked via a short C2–C6 aliphatic chain **9** were also synthesized and evaluated as mutual prodrugs to relieve neuropathic pain.<sup>26</sup>

The limited literature describing gabapentin N- and C-terminus bioconjugates is a testimony to the ease of intramolecular cyclization of **1** to form  $\gamma$ -lactam **2a**<sup>10</sup> (also encountered during our work with gabapentin **1**, see later). Coupling Gbp **1** at its N-terminus with other amino<sup>42,33,22,20</sup> or bile acids was reported using DCC/HOBt reagents and N-Boc or N-Cbz protection.<sup>43</sup> Moderate yields (50–73%) were reported with DCC/HOSu in THF.<sup>44</sup>

## 2. Results and discussion

We now report (i) benzotriazole-mediated acylations of Gbp **1** with amino acids, di- and tri-peptides; together with (ii) a series of N-protected carbonyl-activated gabapentin conjugates with biologically important S-, O-, N-nucleophiles.

### 2.1. Synthesis of Gbp-conjugated at the N-terminus with amino acids and peptides

Our first strategy to stabilize conformationally constrained Gabapentin **1** involved the construction of small  $\alpha$ -, $\gamma$ -peptidomimetics by N-acylation of **1** with carbonyl activated N-protected amino acids and small peptides. Benzotriazolyl precursors were constructed by solution-phase step-wise coupling of intermediate benzotriazolides with unprotected  $\alpha$ - and  $\beta$ -amino acids.<sup>45–47</sup> This method allowed us to build up two- and three-amino acid units that were further activated at the C-terminus carbonyl group as benzotriazolides. For the synthesis of **14a,b**, only one CO-activation step was required. The conjugation of benzotriazolyl amino acids and peptides with Gbp employed known methodology (MeCN, 1.5 equiv TEA, 6 h, rt)<sup>47</sup> to give compounds **14a–d** (77–86%); formation of **2** was not detected. Gabapentin **1** was acylated with COBt-activated N-Cbz protected amino acid segments (Cbz-L-Val-OH, Cbz-L-Ala-OH) to form **14a** (77%), **14b** (86%) N-Boc dipeptide (Boc-L-Ala-L-Phe-OH) **14c** (81%), and **14d** N-Cbz-tripeptide (Cbz-Gly-L-Ala- $\beta$ -Ala-OH) (83%) (Scheme 2). N-Cbz-protected amino acids L-alanine and L-valine used for acylation of **1** at its N-terminus are structurally similar to gabapentin enacarbil **3**. For the same reason—Cbz-Gly-L-Ala- $\beta$ -Ala-Bt **12** was employed as acylation reagent. Such substituents should not restrict the

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