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Non-oxime inhibitors of B-Raf^{V600E} kinase

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ABSTRACT

The development of inhibitors of B-Raf^{V600E} serine-threonine kinase is described. Various head-groups were examined to optimize inhibitor activity and ADME properties. Several of the head-groups explored, including naphthol, phenol and hydroxyamidine, possessed good activity but had poor pharmacokinetic exposure in mice. Exposure was improved by incorporating more metabolically stable groups such as indazole and tricyclic pyrazole, while indazole could also be optimized for good cellular activity.

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The Ras/Raf/MEK/ERK (MAPK) signaling pathway transduces signals from cell surface receptors to the nucleus leading to cellular proliferation, differentiation and survival.¹ Mutations in the BRAF gene may lead to MAPK pathway amplification via constitutive activation of B-Raf kinase and are present in \sim 7% of all cancers.² Mutated B-Raf is most frequently associated with melanoma and has been detected in up to 82% of cutaneous melanocyte nevi,³ 66% of primary melanomas² and 40-68% of metastatic melanomas.⁴ Over 90% of detected mutations in B-Raf are a single glutamic acid for valine substitution at residue 600 (V600E)² which leads to constitutive kinase activity 500-fold greater than B-Raf^{WT} and correlates with increased malignancy and decreased response to chemotherapy.^{5,6} Thus, small-molecule inhibition of B-Raf^{V600E} is a promising strategy in oncology drug development.⁷

In the previous paper we described a series of furopyridine inhibitors of B-Raf^{V600E} (Fig. 1).⁸ While several of these inhibitors demonstrated good enzymatic and cellular inhibition of B-Raf, they displayed insufficient plasma exposure in rodents for in vivo efficacy. The poor pharmacokinetics of this series was attributed to oxime metabolism.^{8,9} In an effort to improve inhibitor pharmacokinetics while maintaining activity against B-Raf, several replacements of the indanone-oxime head-group were explored.

X-ray crystallography revealed several critical interactions between furopyridines and the ATP cleft of the B-Raf catalytic domain (Fig. 2). The template forms several hydrophobic interactions with

residues of the P-loop, the hinge, and the floor of the ATP pocket. The pyridine of the bicyclic core makes a hydrogen bond at the hinge region of the active site to the -NH of Cys532. The indanone moiety occupies the hydrophobic pocket adjacent to the gatekeeper residue Thr529, while the oxime makes two key interactions: the sp² nitrogen acts as an acceptor from the sidechain amine of the catalytic lysine (Lys483) and the hydroxyl group acts as a donor to Glu501 of the C-helix. Initially, we sought to develop a head-group which would make the same interactions with the enzyme as the oxime.

Structure-activity relationships were developed for each head-group by varying the substitution at the 2-position of the furopyridine. Initially, inhibitors were designed that would make one or both of the interactions achieved by the oxime; the activities of these compounds are provided in Table 1. Indanones 2a and 2b were significantly less active against B-Raf in both the



B-Raf IC₅₀ = <2 nM pERK IC50 = 30 nM

Figure 1. Furopyridine inhibitor 1.

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Figure 2. X-ray crystal structure of furopyridine inhibitor 1 (3.7 Å resolution). Hydrogen bonding interactions are illustrated by dashed yellow lines. (a) View of the hingebinding template. For clarity, sidechains are depicted only for those residues that make specific hydrophobic contacts. (b) Close-up of electronic interactions involving the oxime.

Table 1

B-Raf^{VGODE} activity of indanones, hydroxyamidines and pyrazoles



^a Values are means of three experiments.

biochemical (B-Raf^{V600E}) and cellular assays,¹⁰ most likely due to the loss of the hydrogen bond with Glu501. Hydroxyamidines **3a** and **3b** possess the same hydrogen bonding interactions as oximes.¹¹ Isopropyl amide **3a**, while showing excellent activity in the biochemical assay, was poorly active in the cellular assay. Substituting the amide in **3a** for 2-pyrimidine provided compound **3b** and a 70-fold increase in cellular activity. Unfortunately, this compound suffered from rapid clearance in vivo.^{8,12} In an attempt to improve these poor properties while maintaining the interactions of the oxime and hydroxyamidine, the tricyclic pyrazole compounds **4a–c** were prepared. However, none of the analogs provided the necessary level of cellular activity and other headgroup replacements were pursued. Table 2B-RafB-Raf5-9



Compound	R	B-Raf IC_{50}^{a} (nM)	pERK IC_{50}^{a} (nM)
5		179	_
6		30.0	5000
7	NH2	52.0	9100
8	OH	1.1	54
9	F CH	3.0	142

^a Values are means of three experiments.

Naphthyl head-groups were prepared and their biochemical and cellular activities are provided in Table 2. Initial SAR studies conserved the ethyl ester at the 2-position of the furopyridine core while substitutions on the naphthyl group were examined. Unsubstituted naphthyl 5 was weakly active in the biochemical assay. Quinoline 6 and 1-aminonaphthyl 7 were only moderately more active than the naphthyl analog. An appropriately positioned hydroxyl group on the naphthyl head-group (8) provided the naphthol inhibitor class that was of comparable potency to the oximes. Fluorination at the 2-position adjacent to the hydroxyl group (9) led to a small loss of biochemical and cellular activity, likely due to reduced acceptor ability of the hydroxyl. The excellent potency of the naphthol series can be explained by the dual donor/acceptor capability of the hydroxyl group; this group binds to Lys483 and Glu501 similarly to oximes with only a minor movement of the Glu501 sidechain (Fig. 3a).¹³

The naphthol series was substituted with both amides and 2-pyrimidines at the 2-position on the furopyridine core (Table 3). Replacement of the ethyl ester of compound **8** with various amides gave potent compounds bearing both lipophilic

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