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## 2-Alkylamino- and alkoxy-substituted 2-amino-1,3,4-oxadiazoles—O-Alkyl benzohydroxamate esters replacements retain the desired inhibition and selectivity against MEK (MAP ERK kinase)

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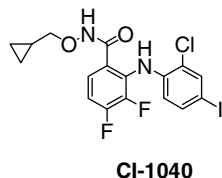
CI-1040

### ABSTRACT

This paper reports a second generation MEK inhibitor. The previously reported potent and efficacious MEK inhibitor, PD-184352 (CI-1040), contains an integral hydroxamate moiety. This compound suffered from less than ideal solubility and metabolic stability. An oxadiazole moiety behaves as a bioisostere for the hydroxamate group, leading to a more metabolically stable and efficacious MEK inhibitor.

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The mitogen-activated protein kinase pathway is thought to be essential in cellular growth and differentiation. The family of mitogen-activated protein (MAP) kinases can be activated in response to a wide variety of extracellular stimuli.<sup>1,2</sup> In particular, MEK (MAP kinase kinase) is an attractive target.<sup>3</sup> This kinase is activated by a cascade of phosphorylation events downstream from ras. Its only known substrates are the MAP kinases, ERK-1 and ERK-2. These kinases phosphorylate a number of transcription factors that control cell growth (e.g., ELK-1). Because of its substrate selectivity in the ras pathway, MEK is an ideal target for intervention of proliferative diseases.<sup>3</sup>



C26 cellular IC<sub>50</sub> = 53 nM  
solubility = <1 µg/mL  
HLM = 8–15 min

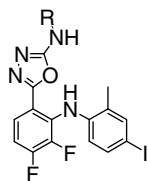
CI-1040

An orally active, potent inhibitor of MEK (CI-1040) was reported by this group.<sup>4</sup> This compound inhibits MEK in a colon 26 cell based assay with an IC<sub>50</sub> of 53 nM. CI-1040 was shown to not be competitive with ATP and is exquisitely selective for MEK (IC<sub>50</sub> > 10 µM against 27 other kinases).<sup>4a</sup> However, this compound suffers from poor solubility and also shows a metabolic liability (human liver microsomes). It was believed that these problems were due in a large part to the key hydroxamic ester moiety. In order to circumvent this liability, a survey of five-membered heterocycles was undertaken. After investigating a number of heterocyclic systems, a 5-amino-1,3,4-oxadiazole was found to have moderate activity against MEK (Table 1).

The synthesis of this oxadiazole<sup>11</sup> is shown in Scheme 1. The anion of 4-iodo-2-methyl aniline was prepared using LiHMDS. This anion was added to the Li salt of 2,3,4-trifluorobenzoic acid in the presence of excess LiHMDS to afford the anthranilic acid **4**. This substitution occurs exclusively at the 2-position of the benzoic acid due to coordination of the aniline anion with the carboxylate. Formation of the hydrazide followed by condensation with cyanogen bromide affords the amino-oxadiazole **1**. Attempts to reductively aminate the oxadiazole **1** proved to be difficult. A number of aldehydes and conditions were tried, but no generally applicable set of conditions were found.

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**Table 1**  
Initial 1,3,4-oxadiazole data

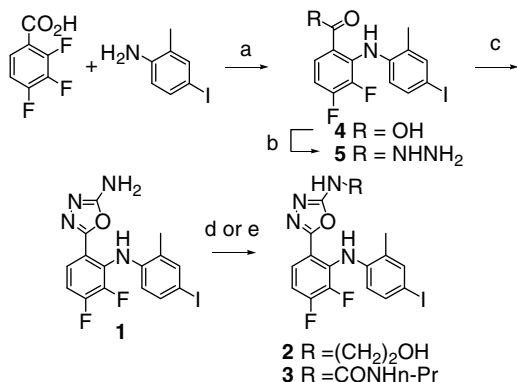
Compound	R	C26 IC <sub>50</sub> <sup>a</sup> (M)
<b>1</b>	H	0.21
<b>2</b>	CH <sub>2</sub> CH <sub>2</sub> OH	1.50
<b>3</b>	CONHCH <sub>2</sub> CH <sub>2</sub> CH <sub>3</sub>	10

<sup>a</sup> Western blot analysis was used to measure MEK inhibitory activity in cultured C26 colon carcinoma cells.

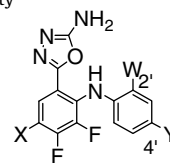
Two derivatives were made, compounds **2** and **3**, in rather low yields after extended reaction times. Neither of these substitutions was found to be advantageous. During this investigation, it was noted that a fluorine in the 2'-position of the anthranilic acid greatly increased potency in the hydroxamate series.<sup>4b,4c</sup> We investigated this substitution and found that a similar effect occurred in the oxadiazole series (Table 2). Simple replacement of the 2' methyl with a fluorine, compound **6**, provided a 5-fold increase in activity, and provided a compound of similar potency to CI-1040. Addition of a chlorine **7** increased potency slightly. The importance of the 4'-iodo is exemplified by **8**, a simple bromo substitution gives only moderate activity.

In order to circumvent the poor reactivity of the 5-amino-1,3,4-oxadiazole, we required a new synthesis. The hydrazide **9** was treated with CDI to provide the intermediate oxazolidinone **10**.<sup>5</sup> This was then treated with various amines to give the semicarbazides **11**,<sup>6</sup> which could be cyclized with either triphenylphosphine or polymer bound triphenylphosphine and carbon tetrachloride<sup>7</sup> to provide the desired 5-amino-oxadiazoles **12**. In the case of **12b–d** (Table 3), the ω-amino BOC was removed using HCl in methanol. In this case, no opening of the oxadiazole was observed (Scheme 2).

The results of the SAR of the substituted amino oxadiazoles is shown in Table 3. The goal of this series of compounds was to maintain or increase potency with a concomitant increase in solubility and metabolic stability. As can be seen, this series proved to be metabolically stable in a human liver microsome assay. Affect on solubility for the most part was disappointing, with the majority of analogs having a solubility on par with CI-1040. Compound **12a** proved to be the most soluble, and retained an activity 2-fold better than CI-1040.



**Scheme 1.** Reagents and conditions: (a) LiHMDS, THF, −78 °C to rt, 90%; (b) hydrazine, PyBop, CH<sub>2</sub>Cl<sub>2</sub>, THF, rt, 94%; (c) CNBr, NaHCO<sub>3</sub>, water, dioxane, rt, 86%; (d) hydroxyacetaldehyde, NaB(OAc)<sub>3</sub>H, DCE, rt, 64 h, 35%; (e) propyl isocyanate, toluene, 130 °C, 52%.

**Table 2**  
Effect of 2' F on cellular activity

Compound	W	X	Y	C26 IC <sub>50</sub> <sup>a</sup> (M)
<b>1</b>	CH <sub>3</sub>	H	I	0.210
<b>6</b>	F	H	I	0.040
<b>7</b>	F	Cl	I	0.018
<b>8</b>	F	H	Br	0.140

<sup>a</sup> Western blot analysis was used to measure MEK inhibitory activity in cultured C26 colon carcinoma cells.

Compound **12a** was further evaluated in an ex vivo pharmacodynamic assay. This assay analyzed p-ERK suppression in tumors following an oral dose of inhibitor. Mice bearing subcutaneously implanted colon 26 tumors were treated with one dose of the inhibitor and after each time point the tumors were excised and Western blots were used to measure p-ERK levels versus vehicle treated controls. As can be seen in Figure 1, **12a** showed superior inhibition of p-ERK levels relative to CI-1040. Most noticeably, at 10 h, **12a** completely suppresses phosphorylation of ERK by MEK at 100 mg/kg dose, and even at 12.5 mg/kg dose, outperforms CI-1040 (100 mg/kg). A high level of inhibition was retained at the highest dose even at 24 h post-dose.

We next investigated the effect of **12a** in vivo. Colon C26 tumors were grown to 200 mg over 8 days, then drug (or vehicle) was administered PO from days 9 to 15 (see Table 4). Compared to CI-1040 (BID, 100 mg/kg), **12a** not only provided 3/3 complete re-

**Table 3**  
SAR for a series a substituted oxadiazoles

Compound	R	n	C26 IC <sub>50</sub> <sup>a</sup> (M)	Solubility <sup>a</sup> (μg/mL)	HLM <sup>b</sup> t <sub>1/2</sub> (min)
<b>6</b>	H	0	0.040	<3	40
<b>12a</b>	NHCH <sub>3</sub>	2	0.027	6.7	40
<b>12b</b>	NH <sub>2</sub>	2	0.015	<3	40
<b>12c</b>	NH <sub>2</sub>	3	0.076	5.4	40
<b>12d</b>	NH <sub>2</sub>	4	0.045	ND	ND
<b>12e</b>	NH(CH <sub>3</sub> ) <sub>2</sub>	2	0.140	ND	ND
<b>12f</b>	OH	2	0.015	<3	40
<b>12g</b>	OH	3	0.028	<3	40
<b>12h</b>	OH	4	0.012	<3	40
<b>12i</b>	OH	5	0.120	<3	40
<b>12j</b>	Morpholine	2	0.032	4.7	40
<b>12k</b>	OMe	3	0.080	<3	35
<b>12l</b>	NH(CH <sub>2</sub> CH <sub>3</sub> ) <sub>2</sub>	3	0.153	<3	40
<b>12m</b>	Morpholine	3	0.311	<3	40
<b>12n</b>	(rac)CH <sub>2</sub> CHOHCH <sub>2</sub> CHOH	0	0.067	<3	40
<b>12o</b>	(S)CH <sub>2</sub> CHOHCH <sub>2</sub> CHOH	0	0.091	3.5	40
<b>12p</b>	CH(CH <sub>2</sub> CHOH) <sub>2</sub>	0	0.770	8.8	40

<sup>a</sup> Solubility determined by measuring the concentration of the compound in a saturated solution over time. In this procedure, increasing aliquots of a 10 mM solution of a compound in DMSO are added to a series of 15 vials containing 1.8 mL of 50 mM sodium phosphate buffer at pH 6.5. The solutions are prepared over the range of 1–60 mg/mL and are allowed to stand overnight at ambient temperature. The samples are vortexed, then HPLC analysis is performed.

<sup>b</sup> Human liver microsomal stability, maximum t<sub>1/2</sub> is 40 min for this assay.

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