

(+)-12 α -Hydroxysophocarpine, a new quinolizidine alkaloid and related anti-HBV alkaloids from *Sophora flavescens*

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Abstract—(+)-12 α -Hydroxysophocarpine (**8**), a new quinolizidine alkaloid was isolated from the roots of *Sophora flavescens*, together with 10 known quinolizidine alkaloids, (+)-oxymatrine (**1**), (+)-matrine (**2**), (+)-9 α -hydroxymatrine (**3**), (+)-allomatrine (**4**), (+)-oxysophocarpine (**5**), (–)-sophocarpine (**6**), (–)-9 α -hydroxysophocarpine (**7**), (+)-lehmannine (**9**), (–)-13,14-dehydrosophoridine (**10**), and (–)-anagyrene (**11**). Their structures were elucidated by spectroscopic methods, and the stereochemistry of **8** was confirmed by X-ray analysis. These alkaloids were tested for anti-hepatitis B virus (HBV) activity in vitro, compounds **5**, **6**, **9**, and **10** showed significant anti-HBV activity with inhibitory potency against HBsAg secretion at 48.3–79.3% and that against HBeAg secretion at 24.6–34.6%.

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Sophora species (Leguminosae) are important sources of Chinese herbal drugs. They accumulate quinolizidine alkaloids as principal constituents with potentially useful pharmacological effects such as analgesic, antipyretic, anti-inflammatory, anti-tumor, and notable antiviral activities.^{1,2} The major quinolizidine alkaloids oxymatrine and matrine were reported to exhibit anti-hepatitis B virus (HBV) activity, oxymatrine could downregulate HBV gene expression and decrease HBsAg and HBeAg content in HBV transgenic mice,³ and protect mice from fulminant hepatitis induced by GalN (galactosamine)/LPS (lipopolysaccharide) and block hepatocyte apoptosis as well,⁴ and matrine could protect the D-GalN-treated mice from the development of fatal hepatitis induced by LPS.⁵

The roots of *Sophora flavescens* Ait., a species widely distributed throughout China, are commonly used as the traditional Chinese medicine 'Kushen' for the treatment of skin diseases and gynecological diseases, such as eczema, dermatitis, and colpitis.⁶ Pharmacological studies

showed that the alkaloids of *S. flavescens* inhibited CBV₃ (coxsackie B₃ virus) replication and possessed protective effect on infected myocardial cells,⁷ and its components anagyrene, oxymatrine, and sophoranol also exhibited potent antiviral activities against RSV (respiratory syncytial virus).⁸ During the course of our screening for anti-HBV agents from *Sophora* plants, a phytochemical investigation on the alkaloid constituents of *S. flavescens* was carried out and resulted in the isolation of a new quinolizidine alkaloid, (+)-12 α -hydroxysophocarpine (**8**), together with 10 known quinolizidine alkaloids (**1–7**, **9–11**) (Fig. 1). This paper reports the isolation and structure elucidation of the new alkaloid, as well as the in vitro anti-HBV activity of these isolated alkaloids.

The roots of *S. flavescens* Ait.⁹ (14 kg) were extracted with aqueous 1% (v/v) H₂SO₄, followed by partition with CHCl₃ after being basified with Na₂CO₃ to give the crude alkaloids, which were subjected to repeated silica gel column chromatography and prep. TLC to give a new quinolizidine alkaloid, (+)-12 α -hydroxysophocarpine (**8**), as well as 10 known ones (**1–7**, **9–11**).¹⁰ These known alkaloids were identified as (+)-oxymatrine (**1**),¹¹ (+)-matrine (**2**),¹¹ (+)-9 α -hydroxymatrine (**3**),¹² (+)-allomatrine (**4**),¹³ (+)-oxysophocarpine (**5**),¹⁴ (–)-sophocarpine (**6**),^{15,16} (–)-9 α -hydroxysophocarpine (**7**),¹⁵ (+)-lehmannine

Keywords: *Sophora flavescens*; Quinolizidine alkaloids; (+)-12 α -Hydroxysophocarpine; Anti-HBV.

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	X	R ¹	R ²	R ³	R ⁴	C-12-C-13	C-13-C-14
1	O	α-H	α-H	H	H	single bond	single bond
2	lone pair	α-H	α-H	H	H	single bond	single bond
3	lone pair	α-H	α-H	OH	H	single bond	single bond
4	lone pair	α-H	β-H	H	H	single bond	single bond
5	O	α-H	α-H	H	H	single bond	double bond
6	lone pair	α-H	α-H	H	H	single bond	double bond
7	lone pair	α-H	α-H	OH	H	single bond	double bond
8	lone pair	α-H	α-H	H	OH	single bond	double bond
9	lone pair	α-H	α-H	H	H	double bond	single bond
10	lone pair	β-H	α-H	H	H	single bond	double bond

Figure 1. Chemical structures of compounds 1–11.

(**9**),^{17,18} (–)-13,14-dehydrosophoridine (**10**),¹³ and (–)-anagyridine (**11**),¹⁹ respectively, by comparison of their $[\alpha]_D$, IR, EIMS, ¹H NMR, and ¹³C NMR spectroscopic data with those reported.

Compound **8**²⁰ was obtained as white plates, mp 90–92 °C, $[\alpha]_D^{24} +137.3^\circ$ (*c* 0.10, MeOH). The quasi-molecular ion $[M+H]^+$ was detected by HRESIMS at *m/z* 263.1761, consistent with the formula of C₁₅H₂₂N₂O₂. Its IR spectrum showed absorption bands characteristic of a hydroxyl group (3311 cm^{−1}), an α,β-unsaturated lactam system (1665 and 1598 cm^{−1}), and *trans*-quinolizidine functionalities (2811 and 2783 cm^{−1}). The EI mass spectrum of **8** showed a base peak at *m/z* 245 (100), corresponding to $[M-OH]^+$, and fragmentations similar to those of (–)-sophocarpine (**6**) and (–)-12β-hydroxysophocarpine,¹⁵ indicating it might be a hydroxyl derivative of **6**. The ¹H NMR spectrum of **8** agreed well with that of **6**, except that there was an additional isolated signal at δ_H 4.17 (1H, m) in the spectrum of **8**

which could be assigned to a methine proton bearing a hydroxyl group because of its low chemical shift, and that the signals due to the olefinic protons H-13 and H-14 simplified in splitting pattern from octets and sextets in **6** to quartets and doublets in **8**, and shifted to downfield for 0.29 and 0.10 ppm, respectively (Table 1). These differences were very similar to those of (–)-12β-hydroxysophocarpine as compared with **6**.¹⁵ The hydroxyl group was thus deduced to be located on C-12, which was further supported by the HMBC correlations of the methine proton at δ_H 4.17 (H-12) bearing a hydroxyl group with C-13 (δ_C 138.7) and C-14 (δ_C 126.6), and of the olefinic protons at δ_H 6.74 (H-13) and δ_H 5.99 (H-14) with the carbon at δ_C 60.9 which correlated with the methine proton at δ_H 4.17 in the HMQC spectrum. In the ¹³C NMR spectrum of **8**, the signals corresponding to C-2–C-6, C-8–C-10, C-15, and C-17 were consistent with those of **6** with $\Delta\delta_C \leq 1.2$ ppm, and the olefinic carbons C-13 and C-14 shifted to downfield for 1.3 and 1.9 ppm,

Table 1. Selected ¹H NMR data of compounds **6** and **8** (in CDCl₃, δ_H in ppm, *J* in Hz)

Compound	11β	12β	13	14	17α	17β
6 ^a	3.98 (dd, 16.9, 9.6)	— ^c	6.45 (ddd, 9.8, 5.0, 3.8)	5.89 (dt, 9.8, 2.2)	4.14 (dd, 12.9, 4.7)	3.17 (t, 12.9)
8 ^b	3.73 (br d, 10.4)	4.17 (m)	6.74 (dd, 9.6, 6.0)	5.99 (d, 9.6)	3.98 (dd, 13.2, 4.6)	3.22 (t, 13.2)

^a 400 MHz.

^b 500 MHz.

^c Not assigned.

Table 2. ¹³C NMR data of compounds **6** and **8** (in CDCl₃, δ_C in ppm)

Compound	2	3	4	5	6	7	8	9	10	11	12	13	14	15	17
6 ^a	57.3	21.1	27.8	34.7	63.6	41.6	26.6	20.8	57.3	51.5	27.4	137.4	124.7	165.7	42.1
8 ^b	57.4	21.2	27.7	33.5	62.7	34.7	26.3	21.0	57.3	54.7	60.9	138.7	126.6	165.8	41.0
$\Delta\delta$ (8–6)	+0.1	+0.1	−0.1	−1.2	−0.9	−6.9	−0.3	+0.2	0	+3.2	+33.5	+1.3	+1.9	+0.1	−1.1

^a 100 MHz.

^b 125 MHz.

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