

## Ammonium tetrathiomolybdate as a water-soluble and slow-release hydrogen sulfide donor



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### ABSTRACT

Ammonium tetrathiomolybdate (TTM) was found to be a slow hydrogen sulfide (H<sub>2</sub>S) releasing agent. Its H<sub>2</sub>S generation capability in aqueous solutions was confirmed by UV–vis and fluorescence assays. TTM also showed H<sub>2</sub>S-like cytoprotective effects in hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>)-induced oxidative damage in HaCaT cells.

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Hydrogen sulfide (H<sub>2</sub>S) is newly recognized as a nitric oxide (NO)-like gaseous transmitter that plays regulatory roles in many physiological and pathological processes.<sup>1–5</sup> Endogenous production of H<sub>2</sub>S involves both enzymatic pathways (mediated by cystathionine β-synthase (CBS), cystathionine γ-lyase (CSE), and 3-mercaptopyruvate sulfurtransferase (3-MST)) and non-enzymatic pathways (from the sulfane sulfur pools). Current knowledge strongly suggests that modulation of H<sub>2</sub>S could have potential therapeutic values for certain disease states, including vasodilation, anti-inflammation, anti-oxidation, and down regulation of cellular metabolism under stress.<sup>1–5</sup> Because of this, the search for compounds that can release H<sub>2</sub>S and mimic the beneficiary activities of H<sub>2</sub>S has become an attractive area in medicinal chemistry.<sup>6–10</sup> So far, a number of H<sub>2</sub>S releasing compounds (also known as H<sub>2</sub>S donors), such as GYY4137, dithiothiones, *N*-mercapto-based donors, persulfide-based donors, *gem*-dithiol-based donors, etc., have been reported (Fig. 1).<sup>6–10</sup> These compounds are normally small organic molecules which can be triggered by certain biologically relevant or compatible reactions to release H<sub>2</sub>S. Some of them have shown promising bioactivities.<sup>6–10</sup> On the other hand, inorganic molecule-based donors, especially metal complexes, which can slowly hydrolyze to release H<sub>2</sub>S have not been reported. Inspired by the fact that sodium nitroprusside (SNP), with the formula Na<sub>2</sub>[Fe(CN)<sub>5</sub>NO], is a widely used NO donor,<sup>11–13</sup> we

suspected that certain sulfide-containing inorganic compounds might be able to serve as interesting H<sub>2</sub>S donors in aqueous solutions. Like SNP, the release of H<sub>2</sub>S from inorganic molecule-based donors is likely to be the result of simple hydrolysis. As such, the donors would be suitable for many biological studies. Herein we wish to report the discovery of ammonium tetrathiomolybdate (TTM) as an effective inorganic H<sub>2</sub>S donor.

TTM, with the formula (NH<sub>4</sub>)<sub>2</sub>MoS<sub>4</sub>, is a commonly used building block in the chemistry of molybdenum.<sup>14,15</sup> As an excellent copper chelator, TTM has been used therapeutically in the treatment of copper toxicosis, especially for Wilson's disease.<sup>16</sup> It was previously noted that under strong acidic condition (5% H<sub>2</sub>SO<sub>4</sub>) H<sub>2</sub>S could be generated from TTM.<sup>17</sup> We first wondered if TTM could produce H<sub>2</sub>S under mild and biologically friendly conditions,

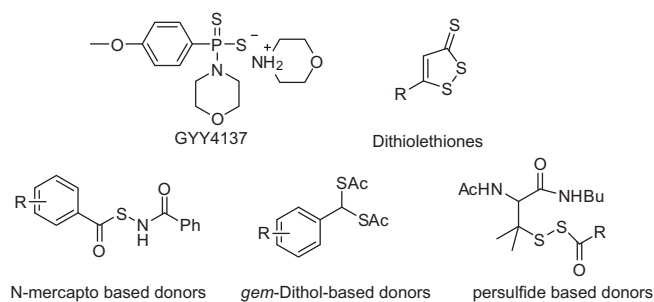


Figure 1. Representative H<sub>2</sub>S donors.

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especially under physiological pH. To this end, we measured  $\text{H}_2\text{S}$  release from TTM in four different pH buffers (5, 6, 7.4, and 8). Normally the release of  $\text{H}_2\text{S}$  from the donors can be determined by the standard methylene blue (MB) method.<sup>18</sup> However, a strong acidic condition was involved in this method. As it is known that acidic media would facilitate TTM hydrolysis, we envisioned that the standard MB method was not appropriate. In this study, a zinc-sulfide precipitation based MB method was used.<sup>19</sup> This method should avoid the false positive signals caused by acid-promoted TTM hydrolysis. Briefly, the solutions of 500  $\mu\text{M}$  TTM were freshly prepared in phosphate buffers under different pH. At different time intervals, aliquots were taken to Eppendorf vials containing a mixture of zinc acetate and NaOH solution. After 15 min, solid ZnS was formed and collected by centrifugation. ZnS was then treated by a MB cocktail. The resulted  $\text{H}_2\text{S}$  concentrations were obtained by UV-vis measurements and calculated based on a calibration curve. As shown in Figure 2, we observed immediate  $\text{H}_2\text{S}$  formation in all of these TTM solutions. The level of  $\text{H}_2\text{S}$  in pH 5 was significantly higher than the levels in other pH values. In all solutions we found  $\text{H}_2\text{S}$  concentrations were maintained in a stable level for a long time (up to 15 h). These results indicate TTM is a stable and slow release  $\text{H}_2\text{S}$  donor.

To further confirm the production of  $\text{H}_2\text{S}$  from TTM in buffers, a  $\text{H}_2\text{S}$  gas trapping experiment was designed. As shown in Figure 3, a solution of TTM was placed in a sealed glass vial. An Eppendorf vial containing a solution of WSP-5, a  $\text{H}_2\text{S}$  specific fluorescent probe,<sup>20</sup> was also placed in the vial to trap the evaporated  $\text{H}_2\text{S}$  from the TTM solution. After incubation at 37 °C for 3 h, the trapping solution was diluted and the fluorescence intensity (excitation at 502 nm, emission at 525 nm) was measured. Both the positive and negative controls (using the standard  $\text{Na}_2\text{S}$  solution or pure buffer) were also carried out using the same procedure for comparison. As shown in Figure 4, TTM and  $\text{Na}_2\text{S}$  led to significant fluorescence increases in the trapping solutions while the pure buffer did not cause any detectable fluorescence increase. The pH dependence of  $\text{H}_2\text{S}$  release from TTM was also noted as higher fluorescence was observed in pH 5 than in other pHs. These results further demonstrated  $\text{H}_2\text{S}$  generation from TTM in buffers.

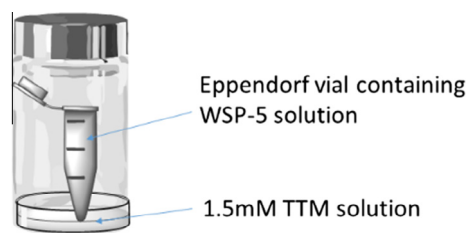


Figure 3.  $\text{H}_2\text{S}$  gas trapping set-up.

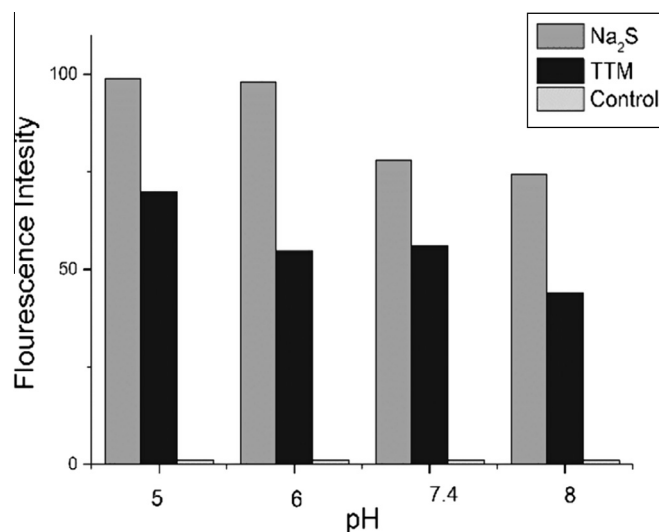


Figure 4.  $\text{H}_2\text{S}$  release of TTM detected by indirect fluorescent assay.

One of the most well-studied biological functions of  $\text{H}_2\text{S}$  is its cytoprotective effects under oxidative stress. Being a  $\text{H}_2\text{S}$  donor, TTM was expected to have similar effects. We then applied TTM

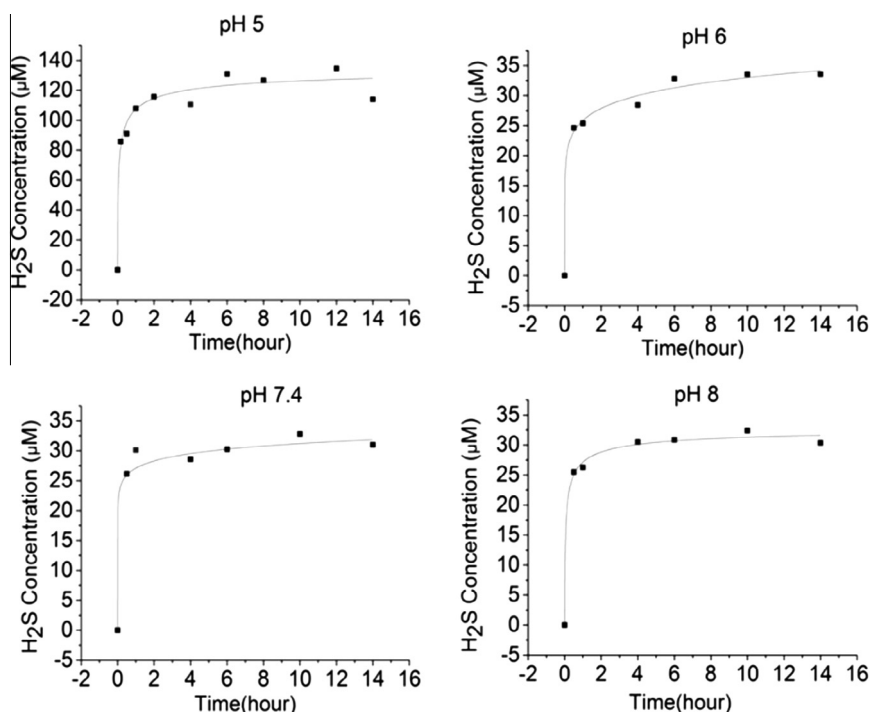


Figure 2.  $\text{H}_2\text{S}$  release from TTM under different pHs.

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