



## Structure guided optimization of a fragment hit to imidazopyridine inhibitors of PI3K



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### ABSTRACT

PI3 kinases are a family of lipid kinases mediating numerous cell processes such as proliferation, migration and differentiation. The PI3 Kinase pathway is often de-regulated in cancer through PI3K $\alpha$  overexpression, gene amplification, mutations and PTEN phosphatase deletion. PI3K inhibitors represent therefore an attractive therapeutic modality for cancer treatment. Herein we describe how the potency of a benzothiazole fragment hit was quickly improved based on structural information and how this early chemotype was further optimized through scaffold hopping. This effort led to the identification of a series of 2-acetamido-5-heteroaryl imidazopyridines showing potent in vitro activity against all class I PI3Ks and attractive pharmacokinetic properties.

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Phosphatidylinositol 3-kinases (PI3Ks) are a family of lipid and serine/threonine kinases involved in a diverse set of cellular functions.<sup>1</sup> PI3Ks can be categorized in class I, II or III, depending on their subunit structure, regulation and substrate selectivity.<sup>2</sup> Class IA PI3Ks are heterodimers composed of a catalytic p110 subunit ( $\alpha$ ,  $\beta$  and  $\delta$  isoforms) constitutively associated with a regulatory subunit that can be p85 $\alpha$ , p55 $\alpha$ , p50 $\alpha$ , p85 $\beta$  or p55 $\gamma$ . The Class IB has one family member, a heterodimer composed of a catalytic p110 $\gamma$  subunit associated with one of two regulatory subunits, p101 or p84. Class IA PI3Ks are activated by tyrosine kinases while Class IB is activated by G protein-coupled receptors, therefore linking upstream receptors with downstream cellular activities including cell growth, proliferation, survival, chemotaxis, cellular trafficking, membrane ruffling and glucose homeostasis,<sup>1</sup> motility, metabolism, inflammatory and allergic responses, transcription and translation.<sup>2,3</sup> PI3Ks catalyze the transfer of phosphate to the D-3' position of inositol lipids to produce phosphoinositol-3-phosphate (PIP), phosphoinositol-3,4-diphosphate (PIP<sub>2</sub>) and phosphoinositol-3,4,5-triphosphate (PIP<sub>3</sub>), second messengers in numerous signaling cascades.<sup>4</sup> In many cases, PIP<sub>2</sub> and PIP<sub>3</sub> recruit AKT to the plasma membrane where it acts as a nodal point for many

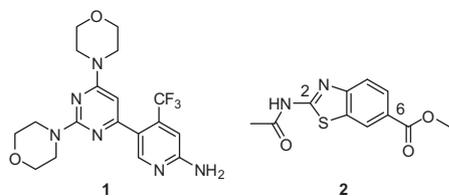
intracellular signaling pathways important for growth and survival.<sup>5,6</sup> Aberrant regulation of PI3K, which often increases survival through AKT activation, is one of the most prevalent events in human cancer<sup>7</sup> and can occur at multiple levels.<sup>8,9</sup> In particular, p110 $\alpha$  pathway deregulation has been demonstrated in ovarian, breast, colon and brain cancers.<sup>10,11</sup> These observations support the inhibition of Class I PI3 kinases as a potential treatment for a variety of tumor types and other proliferative diseases,<sup>12,13</sup> hence the interest during the last 10 years in generating suitable molecules to test this hypothesis in the clinic.<sup>14,15,16,17,18</sup>

In the course of our efforts to identify a backup series for our 2-morpholino pyrimidines front runners<sup>19</sup> and NVP-BKM120 (**1**, Fig. 1)<sup>20</sup> we conducted a second HTS campaign on an expanded compound collection and fragment library. Through this effort, several hits were identified and compound **2** (a fragment with MW = 250.2, Fig. 1) was selected for follow up. Compound **2** had reasonable potency against PI3K $\alpha$  and good ligand efficiency (IC<sub>50</sub> = 2.2  $\mu$ M, LE = 0.47). It appeared to be approximately equipotent against the  $\gamma$  isoform and less potent against the  $\beta$  and  $\delta$  isoforms, PI4K and VPS34.<sup>21</sup> We were therefore intrigued by the opportunity to explore this selectivity potential.

In silico profiling of **2** (PSA = 96, cLogP = 2.0, logD<sub>7.4</sub> = 1.5, 1 HBD, 3 HBA, good predicted passive permeability) did not show any significant alerts. One significant advantage in the follow up

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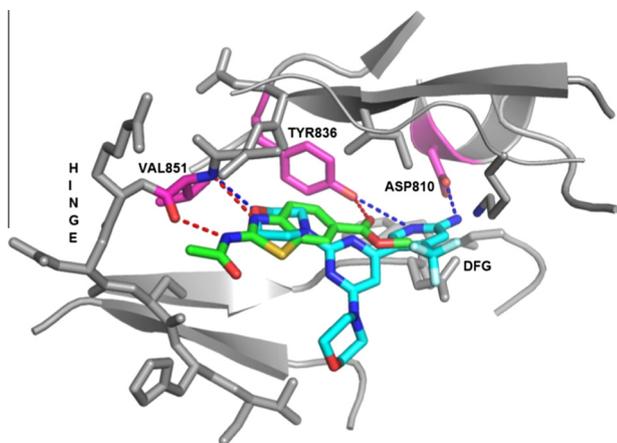
E-mail address: [wooseok.han@novartis.com](mailto:wooseok.han@novartis.com) (W. Han).



**Figure 1.** Clinical candidate **1** (NVP-BKM120) and benzothiazole fragment hit **2**.

was the availability of a co-crystal structure of **2** in p110 $\gamma$  obtained shortly after hit triaging.

**Figure 2** shows an overlay of compound **2** with NVP-BKM120 (**1**, **Fig. 1**), the lead compound from the 2-morpholinopyrimidine series,<sup>20</sup> in a homology model of p110 $\alpha$ . This model was generated based on the co-crystal structures of both compounds in p110 $\gamma$ .<sup>22</sup> It is worth noting that the 2-acetamidobenzothiazole scaffold provides one additional contact with the hinge binding domain of the ATP binding site in comparison to the morpholine. The acetamide NH acts as a H bond donor and the benzothiazole N acts as a H bond acceptor with VAL851 backbone carbonyl and NH, respectively. The third hydrogen bond interaction is provided by the carbonyl of the ester substituent in the 6-position with the OH of TYR836. The benzothiazole spans two key regions of the active site (hinge domain and catalytic region) more efficiently than the 2-morpholinopyrimidine series, where the central pyrimidine ring has mainly a scaffolding function. The ester interaction is likely inefficient, but the 6-position of the benzothiazole provides a good vector for growth in the direction of the ‘affinity pocket’, accessing the same area of the binding site as the one occupied by the aminopyridine in the 2-morpholinopyrimidine series. This indicated the potential for hybridizing the two chemotypes.

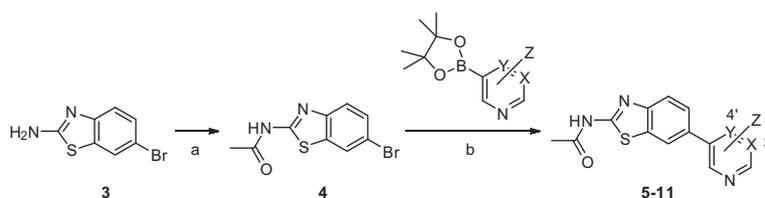


**Figure 2.** Model of compound **2** and NVP-BKM120 (**1**) in p110 $\alpha$ .

We therefore proceeded to synthesize a series of benzothiazoles having six-membered rings at the 6-position. The analogs were easily obtained from commercially available 2-amino-6-bromo benzothiazole **3** by acetylation and Suzuki coupling with various heterocyclic boronic esters, either commercially available or synthesized as previously described<sup>23</sup> (**Scheme 1**). Some of the results obtained are summarized in **Table 1**.

Replacement of the ester in compound **2** with a 3-pyridyl (**5**) gave a 20-fold potency improvement in the biochemical assay, and showed some pathway modulation (IC<sub>50</sub> for the inhibition of AKT phosphorylation on SER473 in the A2780 ovarian carcinoma cell line was 0.5  $\mu$ M). Aminopyridine **6** maintained high Ligand Efficiency (LE = 0.5) and was almost 35-fold more potent than **2**. Consequently, mechanism modulation was enhanced to a level sufficient to observe a signal in our functional assay (inhibition of cell proliferation EC<sub>50</sub> = 2.9  $\mu$ M). Interestingly, aminopyrimidine **7** was comparable to **6**, indicating that the SAR was not tracking with the earlier 2-morpholinopyrimidine series, where 6-aminopyrimidine analogs were 50 to 100-fold more potent than the pyridines.<sup>19</sup> Furthermore, a CF<sub>3</sub> group in the 4'-position of **8**, the preferred position for substitution in the 2-morpholinopyrimidine series,<sup>19,20</sup> was not tolerated. On the other hand, various substituents were tolerated in the 3'-position (e.g. **9–11**). The 3'-CF<sub>3</sub> analog **9** was the most potent with almost 10-fold biochemical potency improvement compared to **6**, and with a two- and seven-fold improvement in mechanism modulation and inhibition of cell proliferation, respectively. It is worth noting, however, that neither LE nor LLE (0.47 and 4.9) were improved for **9** compared to **6** (LLE = 4.8).

Thus, this hybridization approach led to the quick identification of novel, potent PI3K $\alpha$  inhibitors. Potency against the other isoforms was also improved, and the series was not as selective as initially observed for compound **2** (e.g. PI3K $\beta$  IC<sub>50</sub> was 0.030  $\mu$ M for **6**) but was still an interesting starting point for pan Class I PI3K inhibitors. Both compounds **6** and **9** had good microsomal stability and permeability, with low efflux potential in the Caco-2 assay.<sup>24</sup> The compounds were advanced to rat PK (**Table 5**) where they demonstrated oral exposure. However, based on our understanding of PK/PD efficacy relationship in this series, this exposure would not have sufficed for inhibiting tumor growth in our in vivo mouse xenograft model. In addition, low solubility (4.2 and 1.5  $\mu$ M) and CYP450 inhibition (IC<sub>50</sub> for 3A4 = 0.05 and 0.8  $\mu$ M) were significant issues. To address solubility, we introduced basic amines in the 2-position (solvent exposed). Some of the results are shown in **Table 2**. We observed potency loss, but to a lesser extent when more than one carbon tether was introduced on the amide (compare **6** with **13**), and up to 80-fold improvement in solubility (**6** vs **14**). Tracking with the 2-acetamido series, the 3'-CF<sub>3</sub> aminopyridines were more potent than the unsubstituted aminopyridines (10 to 20-fold, **14** vs **13**). Morpholine was also used as solubilizing group (compound **15**), but was less effective. Due to lower efficiency (no potency improvement accompanied the increase in MW) and increased efflux potential (e.g. Caco-2 for compound **11** = A–B 6.7, B–A 18, B/A 2.7), this series was promptly abandoned.



**Scheme 1.** Synthesis of 6-heteroaryl substituted 2-acetamido benzothiazoles. Reagents and conditions: (a) Ac<sub>2</sub>O, THF, rt; (b) 2M Na<sub>2</sub>CO<sub>3</sub>, Pd(dppf)Cl<sub>2</sub>·CH<sub>2</sub>Cl<sub>2</sub>, DME, microwave, 115 °C, 10 min.

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