



Synthesis and antibacterial activities of a novel alkylide: 3-O-(3-aryl-2-propargyl) and 3-O-(3-aryl-2-propenyl)clarithromycin derivatives

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ABSTRACT

A series of novel 3-O-(3-aryl-*E*-2-propenyl)clarithromycin derivatives **8** and 3-O-(3-aryl-2-propargyl)clarithromycin derivatives **11** were designed, synthesized, and evaluated for their in vitro antibacterial activities. Compared with **8c** and **11c** (Ar was 5-pyrimidyl), 3-O-(3-(5'-pyrimidyl)-*Z*-1-propenyl) counterpart **6c** displayed 4- to 64-fold more potent activities against erythromycin-susceptible *Staphylococcus aureus* and *Streptococcus pneumoniae*. Moreover, the activities of **6c**, **8c**, and **11c** against erythromycin-resistant *S. aureus* and *S. pneumoniae* were in general 4-fold higher than those of the reference compound, clarithromycin and azithromycin.

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The second-generation erythromycin, exemplified by clarithromycin (6-O-methylerythromycin, as shown in Fig. 1), was developed to address the acid instability of erythromycin A, which was associated with an intramolecular ketalization of 6-OH with 9-keto.¹ Clarithromycin was successfully synthesized via regioselective methylation at 6-OH of erythromycin.² As a result, clarithromycin exhibits better bioavailability and improved pharmacokinetics.³

Recently, the resistance against erythromycin and clarithromycin is increasingly prevalent via two main genotypes: ribosome methylation or dimethylation resistance encoded by *erm* gene and efflux resistance encoded by *mef* gene.⁴ Meanwhile, the *erm*-mediated resistance exists in two phenotypes: inducible and constitutive resistance.

To combat the resistance, a series of ketolides, derived from erythromycin by substitution of 3-O-cladinose sugar by a 3-keto group, were synthesized and evaluated.⁵ Consequently, ketolides are distinguished by telithromycin (HMR3647),⁶ cethromycin (ABT-773),⁷ and TE-802,⁸ as shown in Fig. 2. Telithromycin and cethromycin were shown to effectively address *erm*- and *mef*-containing resistance, and TE-802 proved to effectively overcome *mef* resistance but possessed weak activities against *erm* resistance. However, the discovery of ketolides disproved the long held belief that 3-O-cladinose was indispensable moiety for the antibacterial activity.

Encouraged by the important breakthrough, a novel acylide characterized by 3-O-acetyl group was prepared after removal of cladinose. Among the candidates, 3-O-(3-pyridyl)acetyl acylide (TEA-0929) possessed better activity against *mef* resistant strains^{9,10} (Fig. 3). In addition to continuous focus on the synthesis of new ketolides and acylides,^{11–13} other macrolides with various substituents at C-3 instead of cladinose have also been designed and investigated over the past decade, such as 3,6-bicyclolide¹⁴ (Fig. 3), 2,3-anhydrolide,¹⁵ 2,3-enol ether,¹⁶ 3-deoxy,¹⁷ 3-O-phenyl ether,¹⁸ 3,6-ketal,¹⁹ and 3,6-ether.²⁰

The structure–activity relationship study indicated that ketolides, acylides, and 3,6-bicyclolides presented good activities against *mef* resistance. Furthermore, an aryl group tethered to macrolides contributed much to overcome *erm* resistance via additional contacts with domain II of bacterial ribosome.

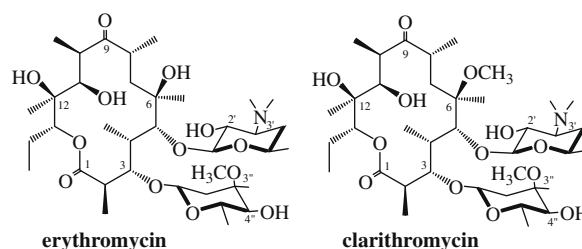


Figure 1. Structure of erythromycin and clarithromycin.

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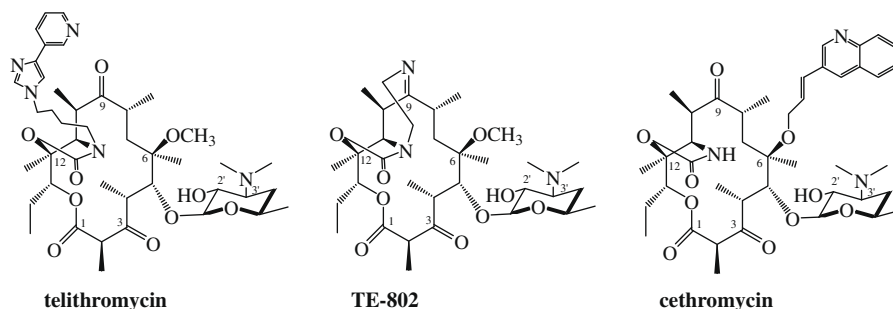


Figure 2. Structure of ketolide.

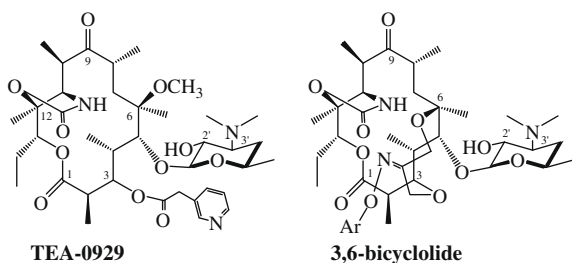
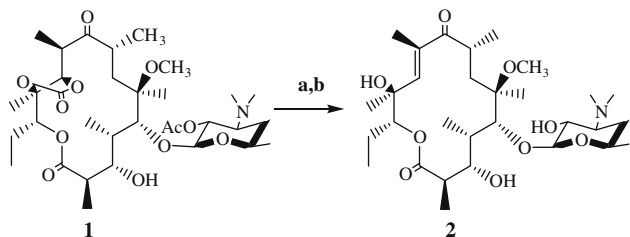


Figure 3. Structure of acylide and 3,6-bicyclolide.

An allyl and a propargyl group have been introduced to 6-OH regioselectively, and the resulting products served as versatile intermediates for a variety of further modification,^{7,21–25} such as the introduction of aryl groups via Heck or Sonogashira reaction. Thus, we presumed that 3-O-allyl clarithromycin and 3-O-propargyl clarithromycin derivatives should be potential lead compounds for further screening. Consequently, we accomplished the regioselective allylation at 3-OH and obtained a series of 3-O-(3-aryl-Z-1-propenyl)clarithromycin derivatives via Heck reaction, which we named alkylides.²⁶ To further study of the structure–activity relationships of alkylides, we herein report the synthesis and the antibacterial activities of 3-O-(3-aryl-*E*-2-propenyl) and 3-O-(3-aryl-2-propargyl)clarithromycin analogues.

A straightforward approach to 3-O-allyl alkylide failed via allylation of 3-OH-9-keto **1**,²⁷ and instead the major product in the resulting mixture was identified as 3-OH-10,11-anhydro **2**, as illustrated in Scheme 1. In contrast, 3-OH-9-O-(2-chlorobenzyl)oxime **3** served as the key precursor to produce corresponding 3-O-allyl **4** in 95.2% yield,²⁶ as presented in Scheme 2.

Interestingly, the Heck reaction of 3-O-allyl **4**, in the presence of palladium(II) acetate and tri(*o*-tolyl) phosphine as catalysts, easily resulted in the allylic double bond isomerization from 2-position to 1-position as well as the configuration isomerization from *E* to *Z* (**5** and **6**),²⁶ presumably due to steric reasons in β -hydrogen elimination (Scheme 2). However, further study in our lab disclosed that the catalyst could efficiently give the desirable sidechain without



Scheme 1. Reagents and conditions: (a) allyl bromide, K₂CO₃, THF/DMSO, 0 °C, 50% (TLC); (b) MeOH, then purified by column chromatography, 11.7% over two steps.

Heck isomerization under the precondition of 2,3-dehydro-3-O-allyl analogues, and the results in detail will be published elsewhere. Thus, the Heck reaction of 3-O-allyl clarithromycin derivatives may warrant further study.

To prevent the unexpected Heck isomerization of **4**, a new kind of palladacycle catalyst, *trans*-di(μ -acetato)bis[O-(di-*o*-tolylphosphino)benzyl]dipalladium(II)²⁸ was utilized in our synthesis of the alkylides (**7** and **8**), as presented in Scheme 2. The catalyst was extensively used in the synthesis of 3-O-aryl-*E*-2-propenyl leucomycin analogues, a 16-membered macrolide.²⁹ As expected, the application of the palladacycle catalyst successfully furnished the desirable 3-O-[3-(3'-pyridyl)-*E*-2-propenyl] (**8b**) and 3-O-[3-(5'-pyrimidyl)-*E*-2-propenyl] sidechain (**8c**). Nonetheless, it was noted that the palladacycle catalyst could not afford 3-O-[3-(4'-isoquinolyl)-*E*-2-propenyl] sidechain (**8e**) efficiently, and the isomerization product (**6e**) still proved to be the majority, as was attributed to the bulky hindrance of 4-isoquinolyl group.

As for the introduction of a propargyl group at 3-OH, it was first reported by Omura group that treatment of 3-OH-6,9-hemiketal derivative, accessible from the acid degradation product of erythromycin, with propargyl bromide and NaH gave 3-O-propargyl-6,9-hemiketal counterpart in 72% yield.³⁰ However, no reaction and activities were reported concerning the Sonogashira coupling products. To further explore the effect of the sidechain's configuration on the activities, the key intermediate 3-O-propargyl alkylide **9** was synthesized, in the presence of propargyl bromide and K₂CO₃, from the precursor 3-OH-9-O-(2-chlorobenzyl)oxime **3** in 89.5% yield. Next, treatment of **9** with a variety of aryl halides under the Sonogashira coupling conditions efficiently led to the expected coupling products **10**. The aromatic rings included 4-nitrophenyl (**a**), 3-pyridyl (**b**), 5-pyrimidyl (**c**), 3-quinolyl (**d**), 4-isoquinolyl (**e**), 5-indolyl (**f**) group. Finally, hydrolysis of the 2'-O-Ac by heating in methanol provided the desired **11** (Scheme 3).

The antibacterial activities of 3-O-(3-aryl-Z-1-propenyl) alkylide **6**, 3-O-(3-aryl-*E*-2-propenyl) alkylide **8** and 3-O-(3-aryl-2-propargyl) alkylide **11** were assessed against a panel of erythromycin-susceptible and erythromycin-resistant bacteria. The selected strains are *Staphylococcus aureus* ATCC 29213 (erythromycin-susceptible strain), *S. aureus* PU-32 (erythromycin-resistant strain bearing inducible *erm* (A) gene), *S. aureus* PU-19 (erythromycin-resistant strain bearing constitutive *erm* (A) gene), *Streptococcus pneumoniae* ATCC 49619 (erythromycin-susceptible strain), *S. pneumoniae* PU-11 (erythromycin-resistant strain bearing both *erm* and *mef* genes), and *S. pneumoniae* PU-13 (erythromycin-resistant strain bearing both *erm* and *mef* genes). Data are listed in Table 1 as the minimal inhibitory concentration (MIC), which is determined by the broth microdilution method as recommended by Clinical and Laboratory Standards Institute (CLSI).³¹

Actually, the activities of the alkylides **6**, **8**, **11** were comparable to those of the corresponding 9-O-(2-chlorobenzyl)oxime ketolides³² and some triazole derivatives started from 3-O-propargyl-6,9-hemiketal.³⁰ The alkylides exhibited improved activities

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