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Influences of operating conditions on biocatalytic activity and reusability of Novozym 435 for esterification of free fatty acids with short-chain alcohols: A case study of palm fatty acid distillate



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ABSTRACT

In the present study, the effects of operating conditions on biocatalytic activity and stability of Novozym 435 for repeated-batch biodiesel production from free fatty acid (FFA) were investigated. Thermal deactivation caused by increased operating temperature from 45 to 50 °C could seriously affect the reusability of Novozym 435. The deactivation of Novozym 435 during the esterification of oleic acid with ethanol tended to be stronger than that in the system with methanol. Under the optimal conditions, considering both biocatalytic activity and stability of the enzyme, Novozym 435 could be reused for 13 cycles for biodiesel productions from oleic acid and absolute alcohols (methanol and ethanol) with FFA conversions of at least 90%. The presence of 4%–5% water in ethanol significantly affected the reusability of Novozym 435. Changes in the surface morphology of Novozym 435 during the esterification with various conditions were observed. It was revealed that the reduction in catalytic activity was related to the swelling degree of the catalyst surface. Additionally, biodiesel production from low cost renewable feedstocks, such as palm fatty acid distillate (PFAD) and 95% ethanol was examined. The esterification of PFAD with 95% ethanol catalyzed by Novozym 435 in 10-repeated batch operation showed the similar results in FFA conversion as compared to those using oleic acid. Novozym 435 remained active and could maintain 97.6% of its initial conversion after being used for 10 batches.

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1. Introduction

The energy supply shortage is currently one of the most important issues that continuously and directly affect humans. In Thailand, biodiesel is one of the most attractive alternative fuels. Biodiesel is an environmentally compatible product that can be produced from renewable resources, such as vegetable oils, free fatty acids (FFAs), animal fats [1,2] or waste cooking oil (WCO) [3] with short-chain alcohols, such as methanol, ethanol, propanol and butanol [4]. The catalysts used in the production of biodiesel can be classified as alkaline catalysts (NaOH or KOH), acid catalysts (H₂SO₄) [5], solid acid and base catalysts [6] and biocatalysts (lipase enzymes).

Lipase can selectively and effectively catalyze both transesterification and esterification reactions with low energy consumption at mild operating temperatures of less than 50 °C [7]. Moreover, this process is more environmentally friendly than chemical processes [8]. Because the use of enzymatic catalysts does not form soaps, it can esterify both FFA and TAG in one step without the need of a subsequent washing step [9]. Since there is no discharge of chemicals and wastewater, the enzymatic

process is considered to be a clean and environment friendly technique. However, the high cost of enzymatic catalysts is considered to be the limiting factor for their commercialization. Therefore, immobilization techniques are used to enhance the potential for industrial-scale enzymatic processes. Immobilization allows for the easy recovery of enzymatic catalysts and for reuse of the catalyst several times without significant losses in activity or stability [10].

Immobilized lipase from *Candida antarctica* lipase B (Novozym 435) is an attractive biocatalyst for the production of biodiesel from many types of oil-containing seed plants in Thailand, such as palm (*Elaeis guineensis*), physic nut (*Jatropha curcas*), papaya (*Carica papaya*) and rambutan (*Nephelium lappaceum*) [11]. The operating conditions, such as the temperature, initial molar ratio of FFA to alcohol, mixing rate and enzyme concentration, have important roles in the enzymatic conversion of FFAs [12–14]. Influence of alcohol structure on the enzymatic activity was also reported [15].

Oleic acid is a major component of various oils, such as palm oil, rapeseed oil and used frying oil [16]. In our previous studies on enzymatic esterification using Novozym 435 in a batch solvent-free system, the optimal conditions for the conversion of oleic acid with methanol and ethanol were reported, and the conversion yields were greater than 90% [13]. In this study, we focused on the study of effects of the operating conditions on the reaction progress and reusability of Novozym

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435 for the esterification of oleic acid with short-chain alcohols. Moreover, for further development of “green technology” for a commercial biodiesel production from low-cost feedstocks, the esterification of palm fatty acid distillate (PFAD), a byproduct from palm oil refining and 95% ethanol using Novozym 435 as the catalyst was evaluated with repeated batch experiments.

2. Materials and Methods

2.1. Materials and chemicals

Novozym 435 (lipase B from *C. antarctica*, EC 3.1.1.3), a nonspecific lipase immobilized on macroporous acrylic resin was purchased from S.M. Chemical suppliers Co., Ltd, Bangkok, Thailand. The diameters of the particle beads are in a range of 0.3–0.9 mm with approximate density of $0.4 \text{ g}\cdot\text{ml}^{-1}$. The catalytic activity was $10000 \text{ PLU}\cdot\text{g}^{-1}$. All other chemicals used in this study were purchased from local suppliers in Thailand.

2.2. Enzymatic esterification reaction

The batch esterification process using 40 g of oleic acid and short-chain alcohols catalyzed by Novozym 435 was performed in a 250 ml Erlenmeyer flask. The optimal operating conditions obtained from our previous work were used, as follows: operating temperature of $45 \text{ }^\circ\text{C}$, FFA to alcohol molar ratio of 1:2, and enzyme loading of 5% (w/w of oleic acid) [13]. To minimize the effect of external mass transfer limitations and maintain the enzyme activity, the mixtures were shaken at a constant rate of $250 \text{ r}\cdot\text{min}^{-1}$ in a shaking incubator (Innova 4000, New Brunswick Scientific Co., Inc., Germany). Samples were collected from the mixtures during the reactions (0–8 h) for the determination of FFA conversion. Water, a reaction byproduct and residual alcohols were removed from the samples via thermal evaporation. The samples were then analyzed using the titration method to determine the FFA conversion. On the study of the reusability of Novozym 435, after each batch, the products and the remaining substrates were removed, and fresh substrates were added for the next cycle.

2.3. Biodiesel conversion analysis

The percentage of FFA conversion was determined by the titration method with $0.1 \text{ mol}\cdot\text{L}^{-1}$ KOH solution using phenolphthalein as the indicator. The FFA conversions were calculated from the titration volumes of the KOH solution. The reported values were the average values of each duplicate set.

2.4. Characterization of Novozym 435

Scanning electron microscope (SEM) was performed to observe morphology changes of the biocatalyst (Novozym 435) after being used in the esterification reaction. Excess oil and solution at the surface of the biocatalysts was blotted out with Kimwipes paper. The samples of biocatalysts were then sputtered with gold and were examined for morphological structures by the scanning electron microscope JSM-5410LV (Tokyo, Japan).

3. Results and Discussion

3.1. Effects of operating conditions on catalytic activity and reusability of Novozym 435

3.1.1. Effect of operating temperature

Higher operating temperature results in higher initial conversion rate and according to the high rates, the reaction reaches equilibrium sooner. However, thermal deactivation of enzymes might

occur and thereby negatively affect the activity, stability and reusability of enzymes. Our previous study showed that there was no significant difference in the FFA conversion and the reaction rate at temperatures between $45 \text{ }^\circ\text{C}$ and $60 \text{ }^\circ\text{C}$ [13]. In the present study, the stability of Novozym 435 during the enzymatic esterification of oleic acid with short-chain alcohols, such as methanol (99.9%) and ethanol (99.9%) was investigated at the operating temperature of $45 \text{ }^\circ\text{C}$ and $50 \text{ }^\circ\text{C}$. As shown in Fig. 1A, with the use of methanol, the initial FFA conversion slightly increased as the temperature increased from $45 \text{ }^\circ\text{C}$ to $50 \text{ }^\circ\text{C}$. However, it was found that the final FFA conversion was not significantly affected by temperature changes in this temperature range. The effect of the thermal deactivation of Novozym 435 at $50 \text{ }^\circ\text{C}$ was more clearly observed in the esterification of oleic acid with ethanol as shown in Fig. 1B. In the first batch with the fresh enzyme, the rates of FFA conversion and the final FFA conversions at the operating temperature of $45 \text{ }^\circ\text{C}$ and $50 \text{ }^\circ\text{C}$ were quite similar. However, from the second batch to the fifth batch, the rates of FFA conversion significantly decreased as the temperature increased from $45 \text{ }^\circ\text{C}$ to $50 \text{ }^\circ\text{C}$. At the operating temperature of $50 \text{ }^\circ\text{C}$, the FFA conversion after 8 h was approximately 91% in the first batch, and it was less than 80% in the fifth batch, whereas no significant drop in FFA conversion was observed with the operating temperature at $45 \text{ }^\circ\text{C}$. The results indicated that for the long-term use of this biocatalyst, the optimal temperature for the esterification of oleic acid with methanol or ethanol by Novozym 435 should be $45 \text{ }^\circ\text{C}$ for maintaining high catalytic activity and reusability of the enzyme.

3.1.2. Effect of alcohols

The effect of alcohols on the enzyme activity in acyl transfer reactions includes reversible inhibition and irreversible inactivation [17,18]. The type of alcohol might directly affect the reusability of enzymes. In this part, the effects of alcohols, methanol (99.9%) and ethanol (99.9%), on the reusability of Novozym 435 in the esterification of oleic acid were investigated. In Section 3.1.1, to reduce the effects of thermal deactivation on the activity and stability of Novozym 435, the reactions were performed at a constant operating temperature of $45 \text{ }^\circ\text{C}$. As shown in Fig. 2, the FFA conversion rates to produce methyl oleate were higher than those of ethyl oleate. During the production of methyl oleate, the conversion rate slightly decreased as the number of enzyme reuse cycles increased. However, a gradual decrease in the FFA conversion rate with increasing number of Novozym 435 reuse cycles was more clearly seen during the production of ethyl oleate. Nevertheless, for both reactions, Novozym 435 could be reused for 13 repeated batches (104 h) with FFA conversions of at least 90%. Previously, it was reported that among all enzymes tested, Novozym 435 was the most effective biocatalyst for the methanolysis in a continuous process [19]. However, the deactivation of Novozym 435 in the esterification could occur because of the interaction of alcohols with the surface of Novozym 435 through the adsorption of alcohols [20]. The denaturing effect of alcohols on proteins, causing enzyme deactivation is well known. Progressive deactivation after several reuses of the biocatalyst during the esterification was also observed [20]. In previous reports, 85% of the initial Novozym 435 activity was maintained after 9 batches in the transesterification of vegetable oils and ethanol [21], and 90% of the activity of Novozym 435 was maintained over 7 batch reactions in the transesterification of vegetable oils and short-chain alcohols [22].

3.1.3. Effect of initial water content in alcohol

Influence of the initial water content on the esterification and transesterification reactions is another important issue. It was reported that transesterification reactions catalyzed by *Candida rugosa*, *Pseudomonas cepacia*, and *Pseudomonas fluorescens* lipases could not occur in the water-free system [9]. However, Novozym 435 exhibited

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