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## Data Article

## Data for the inhibition effects of recombinant lamprey CRBGP on the tube formation of HUVECs and new blood vessel generation in CAM models



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## ABSTRACT

In the present data article, lamprey cysteine-rich buccal gland protein (CRBGP) which belongs to cysteine-rich secretory proteins (CRISPs) family was recombinant and expressed in *Rosetta blue* cells. After identification, the recombinant protein was purified through affinity chromatograph. The inhibition effects of recombinant lamprey CRBGP (rL-CRBGP) on tube formation of human umbilical vein endothelial cells (HUVECs) and new blood vessel generation in chick chorioallantoic membrane (CAM) models were analyzed. This paper contains data related to research concurrently published in “Anti-angiogenic activities of CRBGP from buccal glands of lampreys (*Lampetra japonica*)” [1].

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## 1. Specifications table

Subject area	Biology
More specific subject area	Biochemistry

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Type of data	Figure
How data was acquired	Microscope, mass spectroscopy, camera
Data format	Raw and analyzed, etc.
Experimental factors	PBS and rL-CRBGP were added in HUVECs and CAM models
Experimental features	Protein recombination, expression, separation, purification and identification. Cell culture, tube formation and CAM model assay
Data source location	Dalian, China
Data accessibility	Data is with this article

## 2. Value of the data

- These data are valuable for the soluble expression of the other CRISP family members.
- These data are valuable for the studies of the relationship between other CRISP family members and angiogenesis.

## 3. Data

As shown in Fig. 1, lamprey CRBGP is a very conservative gene and has 45% sequence identity with the ES-CRISP from the snake venom of *Echis carinatus sochureki*. Subsequently, lamprey CRBGP was subcloned into a pEGX-4T-1 vector and expressed as a Glutathione S-transferase (GST)-tagged fusion protein in *Rosetta blue* cells with the molecular weight of 51.6 kDa (Fig. 2). After identification by matrix-assisted laser desorption/ionization time of flight (MALDI-TOF/TOF) analysis, rL-CRBGP was found to exhibit the anti-angiogenic activities in both tube formation and CAM assays (Figs. 3–5).

## 4. Experimental design, materials and methods

### 4.1. Sequence alignment

Additional 10 CRISP sequences from the other species were obtained from ExPASy (<http://www.expasy.ch/tools/blast>). The multiple sequence alignments of CRISPs were performed by ClustalX (1.81) software using default settings [2].

### 4.2. Expression, purification, and identification of rL-CRBGP

A pair of PCR primers (CRBGP-F: 5'-CCGAATTCGCGAGCGTCGTGGCGGCGACA-3'; CRBGP-R: 5'-AGAAGAATGCGCCGCTGCACATCCGTGC-3') was designed based on the sequence of lamprey CRBGP [3], flanked by an *EcoR* I and a *Not* I restriction site. Lamprey CRBGP was amplified and subcloned into a pEGX-4T-1 vector with a GST-tag. rL-CRBGP was expressed in *Rosetta blue* cells induced with 1 mM isopropyl-1-thio- $\beta$ -D-galactopyranoside (IPTG) for 36 h. The cells were collected by centrifugation, and washed in PBS for twice (pH 7.4). Subsequently, the cells were resuspended in the PBS (pH 7.4) and sonicated on ice for 60 min. After centrifugation, the soluble supernatant was collected and

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