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Data Article

Data for proteomic profiling of Anthers from a photosensitive male sterile mutant and wild-type cotton (*Gossypium hirsutum* L.)

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ABSTRACT

Cotton is an important economic crop, used mainly for the production of textile fiber. Using a space mutation breeding technique, a novel photosensitive genetic male sterile mutant CCR19106 was isolated from the wild-type upland cotton cultivar CCR1040029. To study the male sterile mechanisms of CCR19106, histological and iTRAQ-facilitated proteomic analyses of anthers were performed. This data article contains data related to the research article titled *iTRAQ-Facilitated Proteomic Profiling of Anthers From a Photosensitive Male Sterile Mutant and Wild-type Cotton (Gossypium hirsutum L.)* [1]. This research article describes the iTRAQ-facilitated proteomic analysis of the wild-type and a photosensitive male sterile mutant in cotton. The report indicated that exine formation defect is the key reason for male sterility in mutant plant. The information presented here represents the tables and figures that detail the processing of the raw data obtained from iTRAQ analysis.

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Specifications table.

Subject area	Biology
More specific subject area	Plant proteomics
Type of data	Table and figure
How data was acquired	Plant phenotype: DP72 light microscope (Olympus, Japan) Scan electron microscopy: scanning electron microscopy S-530 (HITACHI, Japan) Mass spectrometry: AB SCIEX Triple TOF 5600 System (AB SCIEX, USA) Quantitative real-time PCR: ABI 7500 real-time PCR system (Applied Biosystems, USA)
Data format	Processed
Experimental factors	No pretreatment of samples was performed
Experimental features	Total anther protein was extracted from mutant and wild-type plants by triplicate using a TCA–acetone method. Three replicates iTRAQ-facilitated proteomic analysis were conducted for protein identification and quantification. Any protein changed with $\alpha \geq 1.5$ -fold difference and a p -Value ≤ 0.05 in at least two replicates would thus be considered as a significant DEP in our data.
Data source location	Cotton anther samples were collected in Anyang, Henan Province, China. iTRAQ-facilitated proteomic analysis were conducted in Beijing Genomics Institute, Shenzhen, Guangdong Province, China.
Data accessibility	The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium via the PRIDE partner repository with the dataset identifier PXD002209. The reviewer account: username, reviewer23539@ebi.ac.uk; password: 3ts0ERFU.

Value of the data

- An iTRAQ-based proteomic analysis in cotton anthers.
- Identification of 6,121 high-confidence proteins in cotton anther.
- There are 325 proteins show differential expression patterns between WT and MT.
- The data enrich the understanding of the molecular regulatory mechanisms of male sterility.

1. Experimental design

Using a space mutation breeding technique, a novel photosensitive genetic male sterile mutant CCRI9106 was isolated from the wild-type upland cotton cultivar CCRI040029. Histological and iTRAQ-facilitated proteomic analyses of anthers were performed to explore male sterility mechanisms of the mutant.

2. Materials and methods

2.1. Plant growth and anther collection

Two *G. hirsutum* L. genotypes, a PGMS mutant CCRI9106 and its WT line, CCRI040029, were used in this study. CCRI040029 was an elite upland variety bred in our lab, and the mutant line, CCRI9106, was created by space mutation in 2010 [2]. They were grown in an agronomic field in Anyang (Henan, China) from April to October (Fig. S1), and in Sanya (Hainan, China) from October to early April (Fig. S2). Thirty rows (8 m in length \times 0.8 m in width) were prepared for each genotype, and every 10 rows formed one replicate. To test the pollen fertility, anthers were stained with Alexander's solution. Additionally, anthers from both MT and WT at different development stages were collected for further analysis.

2.2. Scan electron microscopy

For SEM (Fig. S3), anthers were infiltrated with 2.5% (v/v) glutaraldehyde in phosphate buffer (0.1 M, pH 7.2), dehydrated in a graded series of ethanol (from 30% to 100%), treated in acetone for

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