



ELSEVIER

Contents lists available at ScienceDirect

Data in Brief

journal homepage: www.elsevier.com/locate/dib

Data Article

Data for a comprehensive map and functional annotation of the human cerebrospinal fluid proteome



Yang Zhang^{a,1}, Zhengguang Guo^{b,1}, Lili Zou^b, Yehong Yang^b,
Liwei Zhang^a, Nan Ji^a, Chen Shao^c, Yajie Wang^{d,e,*}, Wei Sun^{b,**}

^a Department of Neurosurgery/China National Clinical Research Center for Neurological Diseases, Beijing Tiantan Hospital, Capital Medical University, 6 Tian Tan Xi Li, Beijing 100050, China

^b Core Facility of Instrument, Institute of Basic Medical Sciences Chinese Academy of Medical Sciences, School of Basic Medicine, Peking Union Medical College, 5 Dong Dan San Tiao, Beijing 100005, China

^c National Key Laboratory of Medical Molecular Biology, Department of Physiology and Pathophysiology, Institute of Basic Medical Sciences Chinese Academy of Medical Sciences, School of Basic Medicine, Peking Union Medical College, 5 Dong Dan San Tiao, Beijing 100005, China

^d Core Laboratory for Clinical Medical Research, Beijing Tiantan Hospital, Capital Medical University, 6 Tian Tan Xi Li, Beijing 100050, China

^e Department of Clinical Laboratory Diagnosis, Beijing Tiantan Hospital, Capital Medical University, 6 Tian Tan Xi Li, Beijing 100050, China

ARTICLE INFO

Article history:

Received 27 January 2015

Received in revised form

3 February 2015

Accepted 9 February 2015

Available online 20 February 2015

ABSTRACT

Knowledge about the normal human cerebrospinal fluid (CSF) proteome serves as a baseline reference for CSF biomarker discovery and provides insight into CSF physiology. In this study, high-pH reverse-phase liquid chromatography (hp-RPLC) was first integrated with a TripleTOF 5600 mass spectrometer to comprehensively profile the normal CSF proteome. A total of 49,836 unique peptides and 3256 non-redundant proteins were identified. To obtain high-confidence results, 2513 proteins with at least 2 unique peptides were further selected as bona fide CSF proteins. Nearly 30% of the identified CSF proteins have not been previously reported in the normal CSF proteome. More than 25% of the CSF proteins were components of CNS cell microenvironments, and

DOI of original article: <http://dx.doi.org/10.1016/j.jprot.2015.01.017>

* Corresponding author at: Core Laboratory for Clinical Medical Research, Beijing Tiantan Hospital, Capital Medical University, 6 Tian Tan Xi Li, Beijing 100050, China.

** Corresponding author.

E-mail addresses: zhangyang8025@yeah.net (Y. Zhang), gzg0625@sina.com (Z. Guo), tiantanwyj@aliyun.com (Y. Wang), sunwei1018@hotmail.com (W. Sun).

¹ These authors contributed equally to this work.

<http://dx.doi.org/10.1016/j.dib.2015.02.004>

2352-3409/© 2015 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

network analyses indicated their roles in the pathogenesis of neurological diseases. The top canonical pathway in which the CSF proteins participated was axon guidance signaling. More than one-third of the CSF proteins (788 proteins) were related to neurological diseases, and these proteins constitute potential CSF biomarker candidates. The mapping results can be freely downloaded at <http://122.70.220.102:8088/csf/>, which can be used to navigate the CSF proteome. For more information about the data, please refer to the related original article [1], which has been recently accepted by Journal of Proteomics.

© 2015 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

| | |
|----------------------------|---|
| Subject area | <i>Biology</i> |
| More specific subject area | <i>Proteomics</i> |
| Type of data | <i>Tables</i> |
| How data was acquired | <i>Instruments including Waters Acquity nano-UPLC system, AB SCIEX Triple TOF 5600 system</i> |
| Data format | <i>Analyzed</i> |
| Experimental factors | <i>Protein samples were reduced with 10 mM DTT, alkylated with 55 mM iodoacetamide, digested using sequencing-grade modified trypsin.</i> |
| Experimental features | <i>CSF pooled from 14 individuals (7 women and 7 men) was subjected to the depletion of 14 high-abundance proteins with an immunoaffinity column. The flow-through proteins, bound proteins, and original proteins were collected separately, digested, and then separated into 30 fractions each by high-pH RPLC. Total 90 fractions were subjected to nano-RPLC-MS/MS analysis.</i> |
| Data source location | <i>Beijing, China</i> |
| Data accessibility | <i>The data in the data in brief, in the related original article [1] and at http://122.70.220.102:8088/csf/ can be freely downloaded.</i> |

Value of the data

- This study identified the largest high-confidence dataset of the human CSF proteome.

- Some CSF proteins' abundances are quantified by the iBAQ method.
- High proportion of the CSF proteins is microenvironment components of CNS cells.
- High proportion of the CSF proteins participate in neurocyte connectivity.
- A large part of CSF proteins are biomarker candidates of neurological diseases.

1. Data, experimental design, materials and methods

1.1. Data

Table 1 lists all the CSF proteins with at least 2 unique peptide identifications. Table 2 lists the proteins and their abundances, which were quantified by the iBAQ [2] method. Table 3 lists the CSF proteins that participate in the axon guidance signaling pathway. Table 4 lists the CSF proteins involved in neurological diseases.

Download English Version:

<https://daneshyari.com/en/article/175197>

Download Persian Version:

<https://daneshyari.com/article/175197>

[Daneshyari.com](https://daneshyari.com)