



## Mini Review

## Redox biology of hydrogen sulfide: Implications for physiology, pathophysiology, and pharmacology

Asaf Stein, Shannon M. Bailey\*

Departments of Environmental Health Sciences and Pathology, Center for Free Radical Biology, University of Alabama at Birmingham, Birmingham, AL, USA

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## ABSTRACT

Hydrogen sulfide ( $H_2S$ ) has emerged as a critical mediator of multiple physiological processes in mammalian systems. The pathways involved in the production, consumption, and mechanism of action of  $H_2S$  appear to be sensitive to alterations in the cellular redox state and  $O_2$  tension. Indeed, the catabolism of  $H_2S$  through a putative oxidation pathway, the sulfide quinone oxido-reductase system, is highly dependent on  $O_2$  tension. Dysregulation of  $H_2S$  homeostasis has also been implicated in numerous pathological conditions and diseases. In this review, the chemistry and the main physiological actions of  $H_2S$  are presented. Some examples highlighting the cytoprotective actions of  $H_2S$  within the context of cardiovascular disease are also reported. Elucidation of the redox biology of  $H_2S$  will enable the development of new pharmacological agents based on this intriguing new redox cellular signal.

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**Abbreviations:** ARE, Antioxidant response element; CO, Carbon monoxide; CBS, Cystathionine- $\beta$ -synthase; CGL, Cystathionine- $\gamma$ -lyase; CcO, Cytochrome c oxidase; GSH, Glutathione; HSP, Heat shock protein;  $H_2S$ , Hydrogen sulfide; HIF, Hypoxic inducible factor; IL-1 $\beta$ , Interleukin 1 beta; IL-6, Interleukin 6; 3-MST, 3-mercaptopyruvate S-transferase; NO, Nitric oxide; NF- $\kappa$ B, Nuclear factor light chain enhancer of activated B cells; oxLDL, Oxidized low density lipoprotein; PAG, Propargylglycine; PGE2, Prostaglandin E2; NaHS, Sodium hydrosulfide;  $Na_2S$ , Sodium sulfide; SQR, Sulfide quinone oxido-reductase; TNF- $\alpha$ , Tumor necrosis factor alpha; VEGF, Vascular endothelial growth factor; VSMC, Vascular smooth muscle cells

\* Correspondence to: Department of Pathology, University of Alabama at Birmingham, 1670 University Blvd, Volker Hall G019B, Birmingham, AL 35294-0019, USA. Tel.: +1 205 934 7070; fax: +1 205 975 1126.

E-mail address: [sbailey@uab.edu](mailto:sbailey@uab.edu) (S.M. Bailey).

## 1. Introduction

Hydrogen sulfide ( $H_2S$ ); a toxic gas, is endogenously produced, bioactive, and contributes to numerous physiological functions in mammalian systems. Studies support the possibility that  $H_2S$  has therapeutic potential for treating multiple diseases including cardiovascular diseases. For example, experimental animal studies show that  $H_2S$  may be effective in treating atherosclerosis and protecting against ischemia-reperfusion injury [1–3]. Interest in the cytoprotective actions of  $H_2S$  has grown since the discovery that it can induce a hypometabolic state characterized by decreased  $O_2$  consumption, heart rate, and body temperature in non-hibernating rodents [4]. Although not discussed in this review,  $H_2S$ -dependent hypometabolism is an  $O_2$ -dependent phenomenon [5]. The proposed mitochondrial and signaling actions of  $H_2S$  make this molecule an attractive intervention for preventing and treating diseases and trauma-associated injuries. In this review article, we provide an overview of  $H_2S$  redox biology as it relates to the biological and pharmacological actions of this interesting new signaling molecule in mammalian systems.

## 2. Historical benefits of $H_2S$

The ancient Greeks, Egyptians, and Romans regularly bathed in natural sulfur springs as treatments for disease [6]. Depending on the microbiota and oxygen content, sulfur springs typically contain  $H_2S$  concentrations ranging from 1 to 500  $\mu M$  [7] with anti-inflammatory, anti-bacterial, vasodilatory, and anti-fungal properties attributed to the sulfur-containing water [8]. Epidemiological studies report that a diet rich in organosulfur species is associated with longevity and decreased morbidity [9]. Members of the *Allium* genus (garlic and onions), which contain organosulfur compounds have a well-documented history of health benefits [10]. Indeed, garlic-derived compounds such as diallyl trisulfide release  $H_2S$  in the presence of cellular reductants like glutathione (GSH) [11]. Populations that consume garlic regularly have low blood pressure, low cholesterol, and less vascular disease [12]. While administration of exogenous sulfur-containing compounds shows strong promise as therapies,  $H_2S$  is also endogenously produced in many different human tissues.

## 3. Endogenous production of $H_2S$

In the early 1990s, it was discovered that  $H_2S$  is enzymatically produced by two cytosolic enzymes; cystathionine  $\beta$ -synthase (CBS) and cystathionine  $\gamma$ -lyase (CGL) [13,14]. Seminal work of Abe and Kimura showed, for the first time, that  $H_2S$  enhances long-term potentiation in the hippocampus [15]. Specifically, they demonstrated that  $H_2S$  was produced by CBS and that exogenous  $H_2S$  enhanced NMDA receptor-mediated responses. Since then many studies have shown that CBS and CGL are expressed in human tissues with  $H_2S$  contributing to physiological and pathophysiological processes (Table 1). In addition to CBS and CGL, there are other enzymes that produce  $H_2S$  with several utilizing

cysteine as a substrate. The enzyme 3-mercaptopyruvate S-transferase (3-MST) is found in mitochondria and cytosol and produces  $H_2S$  [16]. Several  $H_2S$  producing enzymes are pyridoxal-5'-phosphate (PLP) dependent enzymes [17]. Moreover, other sulfur-containing amino acids, such as cystine and homocysteine, can be metabolized to generate  $H_2S$ . The enzymatic mechanisms of  $H_2S$  production are shown in Fig. 1. Many of these enzymes participate in the cellular sulfur cycle and have multiple enzymatic activities, including  $H_2S$  generation. Because the concentration of reduced sulfur species has an effect on many cellular processes [18], the activity of these enzymes is tightly regulated. Much of this regulation is linked to substrate availability [19]. Moreover, these are redox-sensitive enzymes, which exhibit increased activity under oxidative conditions [20]. Considering that  $H_2S$ ; a reductant, is a product of these enzymes, it is conceivable that enzymatic activity may also be subject to negative feedback regulation. Finally, work by Wang et al. suggests that under certain conditions, such as oxidative stress,  $H_2S$ -producing enzymes translocate from the cytosol to mitochondria [21]. This dynamic regulation bolsters the argument that  $H_2S$  may function as a redox signaling molecule.

## 4. Catabolism of $H_2S$

Several regulated and unregulated non-enzymatic processes participate in  $H_2S$  catabolism. These pathways maintain *in vivo*  $H_2S$  concentrations, most likely, in the nM to low  $\mu M$  range.  $H_2S$  can react with heme proteins in mitochondria and therefore  $H_2S$  can function as a mitochondrial respiratory toxicant [22,23]. Fatal industrial accidents have been documented in individuals exposed to high concentrations of  $H_2S$  gas (e.g., > 1000 pm) [24]. Therefore, the toxicological profile of  $H_2S$  has been well-studied and documented [24]. The mechanism of toxicity is through the binding of  $H_2S$  to cytochrome c oxidase (CcO) mediating respiratory inhibition [22]. However, this interaction is complex and poorly understood because  $H_2S$  can act as both an inhibitor and an electron donor for CcO [25].  $H_2S$  binds to the oxidized states of the heme  $a-a_3$  binuclear center, resulting in the reduction of the heme molecules [26]. Excess  $H_2S$  can also reduce  $Cu_8$  [27]. While the stoichiometry may vary, Cooper and Brown reported that 3 molecules of  $H_2S$  bind per inhibited CcO [28]. In this inhibitory reaction,  $H_2S$  is oxidized to sulfane sulfur and this is coupled to consumption of molecular  $O_2$  [28]. Unlike nitric oxide ( $^*NO$ ), the inhibition of CcO by  $H_2S$  is noncompetitive with  $O_2$  [27,29]. In addition,  $H_2S$  can also directly reduce the electron carrier cytochrome c producing the one electron oxidation product, the thiyl radical ( $^*SH$ ) [28].

Rhodanese, a mitochondrial sulfur transferase enzyme, catalyzes the oxidation of  $H_2S$  [30]. It is one part of three enzymatic activities characterized as a major pathway for  $H_2S$  catabolism. This pathway consists of a sulfide quinone oxido-reductase (SQR), a sulfur dioxygenase, and the sulfur transferase enzyme rhodanese (Figs. 2 and 3).  $H_2S$  reduces the external disulfide on the SQR to form a thiol (RSH) and a perthiol (RSSH). This two electron oxidation of  $H_2S$  reduces the FAD prosthetic group, which uses ubiquinone (Q) as an electron acceptor [31]. The second sulfur

**Table 1**  
Biological and therapeutic actions of  $H_2S$  are  $O_2$ -dependent.

Biological context	Low $O_2$	High $O_2$
<b>Inflammation</b>	Anti-inflammatory: IL-10; $\downarrow$ IL-6, ICAM	Pro-inflammatory: $\uparrow$ NF- $\kappa$ B, TNF- $\alpha$
<b>Vasoactivity</b>	Vasodilatory: $\uparrow$ $K_{ATP}$ channel conductance	Vasoconstrictive
<b>Angiogenesis</b>	Pro-angiogenic: VEGF, Hif1- $\alpha$	No effect
<b>Respiratory inhibition</b>	Higher [ $H_2S$ ] results in more inhibition	$O_2$ acts as an $H_2S$ antagonist: $\uparrow$ SOx
<b>Ischemia-reperfusion</b>	Ischemic tissue has higher [ $H_2S$ ]: mito $K_{ATP}$ , ARE genes	Unknown

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