



Inhibition of human arginase I by substrate and product analogues ^{☆,☆☆}

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ARTICLE INFO

Article history:

Received 25 November 2009
and in revised form 8 February 2010
Available online 12 February 2010

Keywords:

Metalloenzymes
Enzyme inhibition
Crystallography
SPR
ITC

ABSTRACT

Human arginase I is a binuclear manganese metalloenzyme that catalyzes the hydrolysis of L-arginine to generate L-ornithine and urea. We demonstrate that *N*-hydroxy-L-arginine (NOHA) binds to this enzyme with $K_d = 3.6 \mu\text{M}$, and nor-*N*-hydroxy-L-arginine (nor-NOHA) binds with $K_d = 517 \text{ nM}$ (surface plasmon resonance) or $K_d \approx 50 \text{ nM}$ (isothermal titration calorimetry). Crystals of human arginase I complexed with NOHA and nor-NOHA afford 2.04 and 1.55 Å resolution structures, respectively, which are significantly improved in comparison with previously-determined structures of the corresponding complexes with rat arginase I. Higher resolution structures clarify the binding interactions of the inhibitors. Finally, the crystal structure of the complex with L-lysine ($K_d = 13 \mu\text{M}$) is reported at 1.90 Å resolution. This structure confirms the importance of hydrogen bond interactions with inhibitor α -carboxylate and α -amino groups as key specificity determinants of amino acid recognition in the arginase active site.

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Human arginase I is a binuclear manganese metalloenzyme that catalyzes the hydrolysis of L-arginine to generate L-ornithine and urea. This reaction is the key step of the urea cycle in the liver that allows for the excretion of nitrogenous waste resulting from protein catabolism; healthy adults excrete approximately 10 kg urea per year [1–4]. In extrahepatic tissues arginase serves to regulate L-arginine concentrations for other metabolic pathways. For example, arginase activity can decrease L-arginine concentrations utilized by nitric oxide synthase to generate NO; arginase inhibitors can increase L-arginine concentrations and thereby enhance NO biosynthesis and NO-dependent physiological processes such as smooth muscle relaxation [5]. Accordingly, arginase is a pharmaceutical target for the treatment of diseases associated with aberrant smooth muscle physiology such as erectile dysfunction [6,7] and asthma [8,9].

Among substrate analogue inhibitors of arginase, boronic acid and *N*-hydroxyguanidinium derivatives exhibit the highest affinity (selected inhibitors are shown in Table 1) [7,10–19]. The crystal structures of rat arginase I complexed with 2(*S*)-amino-6-boronohexanoic acid (ABH¹) [6], dehydro-ABH [17], and *S*-(2-boronoethyl)-L-cysteine (BEC) [7], human arginase I complexed with ABH

and BEC [18], and human arginase II complexed with BEC [20] reveal that the planar boronic acid moiety of the inhibitor undergoes nucleophilic attack by the metal-bridging hydroxide ion to yield a tetrahedral boronate anion that mimics the tetrahedral intermediate and its flanking transition states in catalysis. In contrast, the low resolution crystal structures of rat arginase I complexed with *N*-hydroxy-L-arginine (NOHA) and an analogue bearing a side chain shortened by one methyl group (designated nor-NOHA) [21] indicate that the N–OH moiety of the inhibitor displaces the metal-bridging hydroxide ion. Although interactions with the binuclear manganese cluster are strikingly different for the best *N*-hydroxyguanidinium and boronic acid inhibitors, each binds with comparable affinity. With regard to product binding, early studies indicated that L-ornithine and L-lysine are competitive inhibitors of rat and bovine arginase I [22–24]. Moreover, hydroxylation of the side chain amino group of L-lysine yields a potent inhibitor of bovine arginase I with $K_i = 4 \mu\text{M}$ [14]. Here, too, the N–OH group likely displaces the metal-bridging hydroxide ion of the native enzyme.

Given the appreciable inhibitory activity observed for amino acid derivatives of the substrate L-arginine and the product L-ornithine, and given the growing pharmaceutical importance of human arginase isozymes, we now report affinity measurements and X-ray crystal structures of the complexes between human arginase I and nor-NOHA, NOHA, and L-lysine. Importantly, crystals of human arginase I afford 1.55 and 2.04 Å resolution structures of complexes with nor-NOHA and NOHA, respectively, which are significantly improved in comparison with previously-determined structures of rat arginase I complexed with these inhibitors at 2.8 and 2.9 Å resolution, respectively [21]. The crystal structure of the complex with inhibitor L-lysine is determined at 1.90 Å resolution. The higher resolution structures of complexes with human

[☆] This work was supported by NIH grant GM49758.

^{☆☆} Atomic coordinates of human arginase I complexed with *N*⁹-hydroxy-nor-L-arginine (nor-NOHA), *N*⁹-hydroxy-L-arginine (NOHA), and L-lysine have been deposited in the Research Collaboratory for Structural Bioinformatics (<http://www.rcsb.org/pdb>) with the following accession codes: 3KV2, 3LP7, 3LP4.

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¹ Abbreviations used: ABH, 2(*S*)-amino-6-boronohexanoic acid; BEC, *S*-(2-boronoethyl)-L-cysteine; NOHA, *N*-hydroxy-L-arginine; nor-NOHA, nor-*N*-hydroxy-L-arginine.

Table 1
Arginase inhibitors.

Inhibitor	Structure	Affinity, rat arginase I (nM) (K_d or K_i , pH 8.5)	Affinity, human arginase I (nM) (K_d , pH 8.5)
2(S)-amino-6-boronohexanoic acid (ABH)		100 ^a	5 ^b
(S)-(2-boronoethyl)-L-cysteine (BEC)		2200 ^c	270 ^b
<i>N</i> ^ω -hydroxy-L-arginine (NOHA)		10,000 ^d	3600 ^e
<i>N</i> ^ω -hydroxy-nor-L-arginine (nor-NOHA)		500 ^d	517 ^e ; 47, 51 ^f
L-lysine		1,100,000 ^g	13,100 ^e

^a Ref. [10].^b Ref. [18].^c Ref. [7].^d Ref. [12].^e This study, surface plasmon resonance determination.^f This study, isothermal titration calorimetry determination.^g Ref. [24].**Table 2**
Data collection and refinement statistics.

Human arginase I complex	nor-NOHA	NOHA	L-lysine
<i>Data collection</i>			
Resolution, Å	50.0–1.55	50.0–2.04	50.0–1.90
Total/unique reflections measured	135,232/90,987	81,841/40,118	117,869/49,010
R_{merge} ^{a,b}	0.032 (0.302)	0.089 (0.186)	0.081 (0.433)
$I/\sigma(I)$ ^a	14.4 (2.0)	28.9 (8.7)	17.4 (2.5)
Completeness (%) ^a	98.2 (96.2)	98.6 (91.8)	97.1 (98.9)
<i>Refinement</i>			
Reflections used in refinement/test set	85,281/4185	39,744/1587	46,106/1897
R_{twin} ^c	0.144	0.124	0.155
$R_{\text{twin/free}}$ ^c	0.178	0.174	0.204
Protein atoms ^d	4782	4782	4782
Water molecules ^d	302	269	209
Inhibitor atoms ^d	24	26	20
Manganese ions ^d	4	4	4
<i>R.m.s. deviations</i>			
Bond lengths, Å	0.006	0.006	0.006
Bond angles, °	1.34	1.29	1.33
<i>Average B-factors, Å²</i>			
Main chain	18	26	23
Side chain	20	28	26
Manganese ions	13	17	18
Inhibitors	16	41	27
Solvent	23	30	25

^a Number in parentheses refer to the outer 0.1 Å shell of data.^b $R_{\text{merge}} = \sum |I - \langle I \rangle| / \sum I$, where I is the observed intensity and $\langle I \rangle$ is the average intensity calculated for replicate data.^c $R_{\text{twin}} = \sum (|F_{\text{calc}/A}|^2 + |F_{\text{calc}/B}|^2)^{1/2} - |F_{\text{obs}}| / \sum |F_{\text{obs}}|$ for reflections contained in the working set. $|F_{\text{calc}/A}|$ and $|F_{\text{calc}/B}|$ are the structure factor amplitudes calculated for the separate twin domains A and B, respectively. R_{twin} underestimates the residual error in the model over the two twin-related reflections by a factor of approximately 0.7. The same expression describes $R_{\text{twin/free}}$, which was calculated for test set reflections excluded from refinement.^d Per asymmetric unit.

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