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Review 1

Involvement of thiol-based mechanisms in plant development $\stackrel{ ightarrow}{ ightarrow}$ 02

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ABSTRACT

Background: Increasing knowledge has been recently gained regarding the redox regulation of plant develop- 24 mental stages. 25

Scope of view: The current state of knowledge concerning the involvement of glutathione, glutaredoxins and 26 thioredoxins in plant development is reviewed.

Major conclusions: The control of the thiol redox status is mainly ensured by glutathione (GSH), a cysteine- 28 containing tripeptide and by reductases sharing redox-active cysteines, glutaredoxins (GRXs) and thioredoxins 29 (TRXs). Indeed, thiol groups present in many regulatory proteins and metabolic enzymes are prone to oxidation, 30 ultimately leading to post-translational modifications such as disulfide bond formation or glutathionylation. 31 This review focuses on the involvement of GSH, GRXs and TRXs in plant development. Recent studies showed 32 that the proper functioning of root and shoot apical meristems depends on glutathione content and redox status, 33 which regulate, among others, cell cycle and hormone-related processes. A critical role of GRXs in the formation 34 of floral organs has been uncovered, likely through the redox regulation of TGA transcription factor activity. TRXs 35 fulfill many functions in plant development via the regulation of embryo formation, the control of cell-to-cell 36 communication, the mobilization of seed reserves, the biogenesis of chloroplastic structures, the metabolism of 37 carbon and the maintenance of cell redox homeostasis. This review also highlights the tight relationships 38 between thiols, hormones and carbon metabolism, allowing a proper development of plants in relation with 39 the varying environment and the energy availability. 40

General significance: GSH, GRXs and TRXs play key roles during the whole plant developmental cycle via their 41 antioxidant functions and the redox-regulation of signaling pathways. This article is part of a Special Issue 42 entitled: Redox regulation. 43

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1. Introduction 49

50Thiols, the sulfur analogues of alcohols, are organo-sulfur compounds that contain a carbon-bonded sulfhydryl group (R-SH). Thiols are readily 51prone to oxidation especially in a basic environment since thiol depro-52tonation leads to the formation of thiolate (R-S⁻), a nucleophilic form sensitive to oxidation. In the presence of reactive oxygen species (ROS), such as hydrogen peroxide, oxidation leads to the formation of a sulfenic acid form (R-SOH) that can be further oxidized to sulfinic 57and sulfonic forms (R-SO₂H and R-SO₃H, respectively) [1]. Of note, thiol groups can also be oxidized by reactive nitrogen species resulting

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for instance in S-nitrosylation (R-SNO) [2]. In other respects, when an 59 oxidized cysteine residue is brought near a reduced cysteine residue, it 60 generates a cystine unit with a disulfide bond (R-S-S-R') that can alter 61 protein folding, contributing to tertiary structure if the cysteines belong 62 to the same polypeptide or to quaternary structure if they belong to 63 different polypeptides. Owing to the physico-chemical properties of 64 thiol groups and to their capacity to bind metals, proteins having these 65 reactive cysteines play key roles in biology. They contribute for instance 66 to ROS detoxification, to the control of cell redox homeostasis or 67 to signaling transduction pathways in particular via redox post- 68 translational modifications.

In most living organisms, thiol groups are mainly present in the 70 sulfur-containing amino acid, cysteine, which is present in most 71 proteins and is also a component of glutathione, a γ -Glu-Cys-Gly 72 tripeptide existing either in a reduced form (GSH) or in oxidized 73 forms, such as glutathione disulfide (GSSG) or nitrosoglutathione 74

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(GSNO). In plants, glutathione is present in most tissues with subcellu-75 76 lar concentrations in the millimolar range [3,4] and is considered together with ascorbate as a major redox buffer [3,5]. Two enzymes: 77 78 γ -glutamyl-cysteine ligase or γ -glutamyl-cysteine synthase (γ -GCS or GSH1) and glutathione synthetase (GS or GSH2) ensure the production 79 of glutathione at the expense of ATP. The reduction of GSSG is catalyzed 80 by glutathione reductase (GR), a NADPH-dependent flavoprotein [5]. 81 82 The redox status of protein cysteinyl residues is controlled by two 83 main types of enzymes termed thioredoxins (TRXs) and glutaredoxins 84 (GRXs). Many of them, particularly TRXs, display disulfide reductase activity, and some are able to reduce sulfenic acid forms. GRXs are 85 more specifically involved in the deglutathionylation of proteins revers-86 ibly modified by the formation of a mixed disulfide bond between 87 glutathione and a cysteine. However, whereas it is firmly established 88 that all GRXs from class I possess deglutathionylation activities, it is 89 less clear whether GRXs belonging to classes II and III possess such 90 activity [6]. Both TRXs and GRXs function via a catalytic site formed at 91 92least by a redox active cysteine which is generally separated in the primary sequence by two variable residues from another cysteine 93 (often referred to as resolving cysteine) in most TRXs and in about 94 half of GRXs or from a serine in the half remaining GRXs [6,7]. A 95 96 WCGPC amino acid signature is generally quite well conserved among 97 TRXs whereas a greater variability is found among GRXs being often of the CPYC, CPFC, CGFS or CCxC/S type. To obtain a complete view of the 98 active site sequences and current classification of GRXs and TRXs, we 99 invite the reader to refer to the following papers [8,9]. At the structural 100 level, TRXs and GRXs adopt a specific TRX-fold that is usually character-101 102ized by a 4-stranded parallel beta-sheet core enclosed by 4 to 5 alphahelices, and that is shared by several other protein families. In plant 103 cells, the regeneration of oxidized TRXs and GRXs is generally achieved 104 via distinct pathways (Fig. 1). For the large majority, GRXs use reduced 105glutathione as an electron donor but for example, a chloroplastic class II 106 107Grx from Chlamydomonas reinhardtii was shown to be recycled by the ferredoxin-Trx reductase [7,10]. The reduction of TRXs is more complex 108 and further depends on their subcellular localization. Nuclear, cytosolic 109and mitochondrial TRXs are usually reduced by NADPH-thioredoxin 110 reductases (NTR), enzymes with FAD cofactors [11] whereas chloroplas-111 tic TRXs are reduced in a light-dependent manner by a ferredoxin-112 dependent thioredoxin reductase (FTR), an iron-sulfur (Fe-S) enzyme 113 composed of two different subunits and which converts the electron 114 signal coming from photosystem I and relayed by ferredoxins into a 115 116 thiol-reducing cascade [12]. Atypical reduction modes exist for some TRXs. A poplar TRX h4 and its plant orthologs are regenerated by the 117

GSH/GRX system owing to the presence of an additional cysteine in 118 the N-terminal region [13–15]. Some Trx-like and Trx-lilium can be 119 recycled by GSH in a manner analogous to GRXs [16]. 120

Until the last years, glutathione was mainly thought to act as a redox 121 buffer in various processes like responses to environmental stress, 122 plant–pathogen interactions and detoxification of xenobiotics and 123 heavy metals [5]. Plant TRXs have been long presumed to mainly 124 regulate photosynthesis and carbon metabolism enzymes [17] or to 125 participate in the mobilization of seed reserves [18]. They have been 126 also characterized as important actors in the responses of plants to 127 oxidative stress [19]. The information concerning the sequence characteristics, the biochemical functions or the expression patterns of GRXs 129 lag behind, since they have been identified later in plants compared 130 with TRXs [7]. Although some have been reported quite early to play 131 roles in flower development [20,21] and later in stress responses [22, 132 23], the physiological roles of many GRXs remain elusive. 133

A role for TRXs and GRXs as key determinants in plant growth and 134 development has only been recently uncovered together with the 135 involvement of several ROS like NO or H_2O_2 . This is in fact not surprising 136 considering the capacity of the latter molecules to modify protein thiol 137 groups. The cellular redox homeostasis is varying during specific devel- 138 opmental stages or in response to changing environmental conditions. It 139 has thus to be tightly regulated and among the various types of effec- 140 tors, thiol-containing components form an important signaling 141 network. An additional layer of new information concerning the 142 involvement of redox regulated circuits into developmental processes 143 is the complex interplays that have been unveiled between glutathione 144 in particular and several hormones. In this review, we describe the 145 current knowledge and recent advances concerning the roles of gluta- 146 thione and of disulfide reductases, GRXs and TRXs, in the control of 147 development in relation to ROS and hormones. 148

2. Glutathione is essential for plant development

2.1. Direct involvement of glutathione in developmental stages of plants 150

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Glutathione is synthesized in two steps catalyzed by the gammaglutamyl cysteine synthase (GSH1) and the glutathione synthase (GSH2). In higher plants, GSH1 is exclusively located in plastids whereas GSH2 is dual-targeted to plastids and cytosol [24,25]. The crucial role of glutathione for plant development was first demonstrated by the characterization of different mutants in the biosynthesis pathway (Table 1). Unless GSH is added to the growth medium, Arabidopsis 157

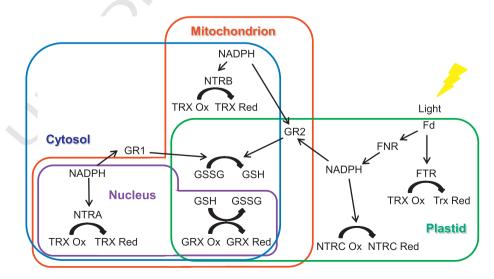


Fig. 1. Reduction pathways of the thiol-containing compounds, glutathione, glutaredoxins and thioredoxins in the main subcellular compartments of plant cells. GSH and GSSG, reduced and oxidized glutathione, respectively; GR, glutathione reductase (2 types, 1 and 2); GRX Ox and Red, oxidized and reduced glutaredoxin, respectively; TRX Ox and Red, oxidized and reduced thioredoxin, respectively; NTR: NADPH thioredoxin reductase (3 types: A, B and C); Fd, ferredoxin; FNR, ferredoxin NADP⁺ reductase; FTR, ferredoxin reductase.

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