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Characterizing urinary hCGBcf patterns during pregnancy

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A R T I C L E I N F O

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ABSTRACT

Objective: Elevated concentrations of hCG beta core fragment (hCG β cf) are known to cause false-negative results in qualitative urine pregnancy test devices, but the pattern of urinary hCG β cf during normal pregnancy has not been well characterized. Here, we evaluate the relationship between urine hCG, hCG β cf, and hCG free β subunit (hCG β) during pregnancy.

Design and methods: Banked second trimester urine specimens from 100 pregnant women were screened for high concentrations of hCG β cf using a qualitative point-of-care device known to demonstrate false-negative results in the presence of elevated hCG β cf concentrations. Additional first and third trimester specimens from the same pregnancy were obtained from 10 women who generated negative/faint positive results, 5 women who generated intermediate positive results, and 10 women who generated strong positive results on the point-of-care device. Intact hCG, hCG β cf, hCG β , and specific gravity were quantified in these 75 specimens.

Results: Urinary hCG β cf concentrations were greater than intact hCG concentrations at all times. A strong correlation ($r^2 = 0.70$) was observed between urine intact hCG and hCG β cf concentrations. A poor correlation was observed between specific gravity and intact hCG ($r^2 = 0.32$), hCG β ($r^2 = 0.32$), and hCG β cf ($r^2 = 0.32$). The highest hCG β cf concentrations were observed between 10 and 16 weeks gestation but individual women demonstrated very different patterns of hCG β cf excretion.

Conclusions: Urine specimens with elevated hCG β cf are frequently encountered during pregnancy but hCG β cf excretion patterns are unpredictable. Manufacturers and clinicians must appreciate that hCG β cf is the major immunoreactive component in urine during pregnancy and must design and interpret qualitative urine hCG test results accordingly.

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1. Introduction

Human chorionic gonadotropin (hCG) is a well-characterized glycoprotein hormone secreted by the trophoblast cells of the placenta that maintains the corpus luteum and supports fetal growth [1]. Several hCG variants, including intact hCG, nicked hCG (hCGn), free β subunit (hCG β), and nicked hCG β (hCG β n), can be detected in both serum and urine while the core fragment of hCG β (hCG β cf) is detected only in urine [1,2]. Importantly, the concentrations and relative proportions of these variants change throughout pregnancy [2,3].

High concentrations of hCG β cf (\geq 500,000 pmol/L) have been shown to cause false-negative results in certain pregnancy tests i.e. qualitative

¹ UHS has served as a consultant to PerkinElmer Wallac.

point-of-care (POC) hCG devices [4–6]. These pregnancy tests are often used to exclude pregnancy in women subjected to administration of radioactive isotopes for therapeutic or diagnostic purposes. Recent work has demonstrated that the majority of qualitative POC hCG devices are susceptible to false negatives due to high concentrations of hCG β cf [7]. Unfortunately, efforts to predict which women are likely to generate elevated hCG β cf concentrations have been largely unsuccessful [8].

While previous publications have reported mean urinary intact hCG, hCG β cf, and hCG β concentrations from multiple women during early pregnancy [2] or hCG and hCG β cf concentrations from a single woman throughout pregnancy [1], little has been published about the individual excretion patterns of urinary intact hCG, hCG β cf, and hCG β in large numbers of women at multiple time points throughout pregnancy. It is unknown whether all women display similar excretion patterns of hCG variants or if the changes in the absolute and relative concentrations of these immunoreactive forms are specific to each woman.

Here, we characterize the relationship between urinary intact hCG, hCG β cf, and hCG β in first, second, and third trimester specimens from 25 women.

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² AMG has served as a consultant and expert witness to Church and Dwight and has received income from Church & Dwight for work that is not a part of this study.

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2. Materials and methods

2.1. Patient samples

One hundred randomly selected second trimester urine specimens were obtained through the Washington University Women and Infant's Health Specimen Consortium (WIHSC), a biobank of specimens from

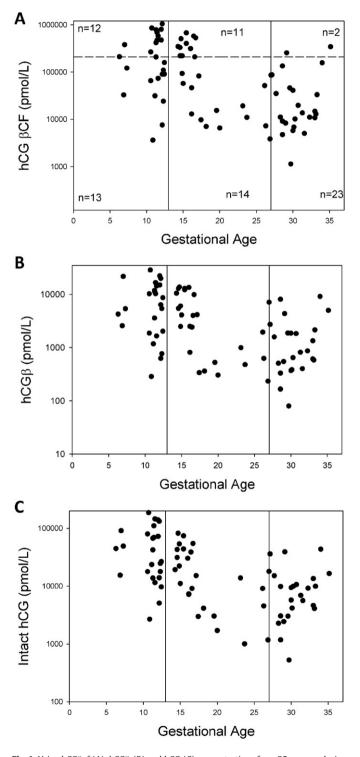


Fig. 1. Urine hCG β cf (A), hCG β (B) and hCG (C) concentrations from 25 women during pregnancy. Samples were collected during the first, second, and third trimesters. Solid vertical lines represent the trimester cutoffs of 13 weeks and 26 weeks. Dotted horizontal line represents hCG β cf concentration of 209,000 pmol/L. False-negative/low-positive results were observed using the OSOM device at this concentration and above.

pregnant women. Additional first and third trimester specimens were obtained from 25 of these patients for subsequent analysis. All samples were stripped of identifiers and coded by the WIHSC. Urine specimens were collected during routine physician office visits and were not necessarily first morning collections. Samples were refrigerated within 4 h of collection, aliquoted and frozen within 12 h of collection, and were stored for up to 4 years at -80 °C. Institutional review board approval was obtained for this study.

2.2. Qualitative hCG measurement

The OSOM POC device (OSOM hCG Combo Test, Genzyme Diagnostics) was used (according to the manufacturer's instructions) as a crude screening device for elevated concentrations of hCG β cf. This device has previously been shown to produce false-negative results at hCG β cf concentrations > 500,000 pmol/L [4,7]. One hundred second trimester urine samples were screened and ten urine samples that showed negative/weak positive, five urine samples that showed intermediate positive and ten samples that showed strong positive results were selected for further analysis. This approach was taken to ensure the widest range of hCG β cf concentrations.

2.3. Quantitative hCG, hCG_βcf, and hCG_β measurement

Samples were shipped on dry ice and stored frozen at -80 °C for quantitative measurement of intact hCG, hCG β , and hCG β cf by time-resolved immunofluorometric assays as described previously [9,10]. Briefly, 25 μ l of urine sample was incubated with 200 μ l assay buffer

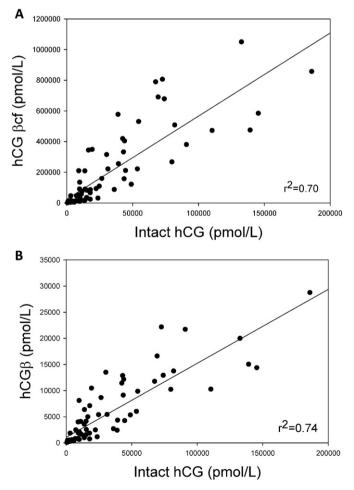


Fig. 2. Correlation between urine hCG and hCG β cf (A) and hCG β (B) concentrations in 75 samples from 25 pregnant women. Solid line represents line of regression.

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