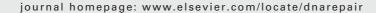


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Adenovirus mediated transduction of the human DNA polymerase eta cDNA

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ABSTRACT

Xeroderma pigmentosum (XP) is an autosomal recessive photosensitive disorder with an extremely high incidence of skin cancers. Seven complementation groups, corresponding to seven proteins involved in nucleotide excision repair (NER), are associated with this syndrome. However, in XP variant patients, the disorder is caused by defects in DNA polymerase η ; this error prone polymerase, encoded by POLH, is involved in translesion DNA synthesis (TLS) on DNA templates damaged by ultraviolet light (UV). We constructed a recombinant adenovirus carrying the human POLH cDNA linked to the EGFP reporter gene (AdXPV-EGFP) and infected skin fibroblasts from both XPV and XPA patients. Twenty-four hours after infection, the DNA polymerase n-EGFP fusion protein was detected by Western blot analysis, demonstrating successful transduction by the adenoviral vector. Protein expression was accompanied by reduction in the high sensitivity of XPV cells to UV, as determined by cell survival and apoptosis-induction assays. Moreover, the pronounced UVinduced inhibition of DNA synthesis in XPV cells and their arrest in S phase were attenuated in AdXPV-EGFP infected cells, confirming that the transduced polymerase was functional. However, over-expression of polymerase η mediated by AdXPV-EGFP infection did not result in enhancement of cell survival, prevention of apoptosis, or higher rate of nascent DNA strand growth in irradiated XPA cells. These results suggest that TLS by DNA polymerase η is not a limiting factor for recovery from cellular responses induced by UV in excision-repair deficient fibroblasts.

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1. Introduction

Cellular DNA is continuously exposed to a large variety of DNA-damaging agents, and all living organisms are equipped with DNA repair systems responsible for removal of lesions and maintenance of genetic integrity [1]. The repair processes, however, are in some cases slow and incomplete, forcing cells to replicate the genome containing persistent damage, and special cell machineries have also evolved to undertake this function.

Ultraviolet (UV) light, present in solar radiation, is known to induce two major types of DNA lesions: cyclobutane-pyrimidine-dimers (CPDs) and pyrimidine-(6-4)-pyrimidone photoproducts (6-4PPs). These lesions cause distortions in the DNA double helix and inhibit essential cellular processes, such as DNA replication and RNA transcription. UV-induced DNA photoproducts have been extensively studied and implicated in skin carcinogenesis.

Xeroderma pigmentosum (XP) is an autosomal recessive syndrome characterized by hypersensitivity to UV-induced DNA damage, resulting in high frequency of skin cancer, even in moderate exposure to solar radiation [2]. Most XP patients are defective in nucleotide excision repair (NER), the pathway that recognizes and removes bulk lesions from the genome, such as UV-induced DNA photoproducts [3]. XP variant (XPV) patients, accounting for approximately 25% of XP cases, do not present any abnormality in NER pathway, but are affected in their capacity to replicate DNA after UV-irradiation [4]. Although the existence of the variant group of XP patients has been known for more than 3 decades [5,6], only recently the gene responsible for the XPV phenotype has been identified: POLH, the gene that encodes DNA polymerase η (pol η) [7,8].

Pol $_{\eta}$ belongs to the Y-family of mammalian DNA polymerases involved in translesion DNA synthesis (TLS), a damage-tolerance mechanism that supports the direct bypass of template DNA lesions, which cannot be accomplished by the high-fidelity DNA polymerases [9,10]. Polymerases belonging to Y-family (pol $_{\eta}$, pol $_{\kappa}$ and Rev1) are characterized by lower processivity and high frequency of nucleotide mis-incorporation in vitro [11,12]. Interestingly, XPV cells are known for their hypermutable phenotype [13], suggesting that few errors are made by pol $_{\eta}$ during the TLS process and favoring the hypothesis that pol $_{\eta}$ is a relatively accurate DNA polymerase when replicating UV-damaged templates [14].

Pol η is a protein of 713 aminoacids, but its polymerase activity is contained within 511 aminoacids of the N-terminal region [7]. However, XPV patients expressing pol η with intact polymerase activity and truncated C-terminus were found, indicating that the latter region plays an important functional role. Analysis of this region of pol η has shown the presence of at least two important domains: the first one corresponds to approximately 70 aminoacids and contains a nuclear localization signal and the other, with approximately 50 aminoacids, is responsible for directing pol η to replication foci after UV-irradiation [15]. A putative third motif in the C-terminus region of pol η seems to include a zinc-finger. Although the presence of this motif still needs to be con-

firmed, it is conserved from yeasts to humans. In vitro, this motif is not required for the polymerase activity of pol_{η} , but it could have an important role in vivo, increasing its binding to DNA when replication forks are blocked by UV photolesions [15].

Although in vitro experiments have shown that pol_{η} can replicate DNA templates containing other types of lesions, CPDs appear to be its main target lesions [7,8,14]. In contrast to most TLS polymerases, pol η has been found to bypass CPDs efficiently and accurately. Poln accuracy in vitro seems to result from both its capacity to select the correct nucleotide during TLS and its failure to elongate DNA chains with mispaired 3'-termini [14]. Interestingly, structural analysis of the Rad30 protein, polη homologue in Saccharomyces cerevisiae, revealed the presence of a much more open active site in this protein, compared to replicative DNA polymerases. This site might accommodate two nucleotides at the same time and this characteristic could explain why this polymerase can replicate through CPDs [16]. Additional experiments showed that pol_{η} usually inserts two adenines opposite thymine dimers, the CPD most frequently formed in UV-irradiated DNA; these findings provide an explanation for why mutagenesis at TT sites is low and why mutations induced by UV-irradiation are preferentially located at dipyrimidine sites containing cytosines [17,18].

In recent years, adenoviral vectors have been used for research and therapeutic purposes. Adenoviral vectors have some potential advantages when compared to other vectors, such as they can be easily manipulated in vitro and are normally produced at very high titers. Furthermore, these viruses are able to infect a wide variety of cell types, including differentiated and non-dividing cells. These characteristics make adenoviral vectors powerful tools for efficient transduction of foreign DNA into human cells [19,20]. We have constructed a recombinant adenovirus carrying human POLH cDNA linked to the EGFP reporter gene (AdXPV-EGFP). XPV cells infected with this adenoviral vector displayed higher UV-resistance than non-infected ones, as determined by increased cell survival, reduced apoptosis, decreased S-phase arrest and increased rate of DNA elongation after UV-irradiation. Taken together, these results indicated that poln-EGFP is functional and efficiently corrected the XPV cells' defect. On the other hand, XPA cells over-expressing pol η (endogenous pol η plus pol η -EGFP from AdXPV-EGFP) did not show any improvement in UV-sensitivity (apoptosis and cell survival) or DNA synthesis after UV-irradiation. These results confirmed that pol_{η} contributes to preventing cell death triggered by UV-DNA damage in human cells, but it cannot overcome the deficiency in the removal of those lesions by nucleotide excision repair.

2. Materials and methods

2.1. Cell lines and culture conditions

SV40-transformed skin fibroblasts were isolated from skin biopsies of XPV (XP30RO-SV) and XPA (XP12RO-SV) patients. MRC5-V1 fibroblasts display normal TLS on UV-damaged templates, as well as NER, and were used as a positive con-

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